# Research article

# **Open Access**

# Comparative venom gland transcriptome surveys of the saw-scaled vipers (Viperidae: *Echis*) reveal substantial intra-family gene diversity and novel venom transcripts

Nicholas R Casewell<sup>\*1</sup>, Robert A Harrison<sup>2</sup>, Wolfgang Wüster<sup>1</sup> and Simon C Wagstaff<sup>\*2</sup>

Address: <sup>1</sup>School of Biological Sciences, Bangor University, Environment Centre Wales, Bangor, UK and <sup>2</sup>Alistair Reid Venom Research Unit, Liverpool School of Tropical Medicine, Liverpool, UK

Email: Nicholas R Casewell\* - n.r.casewell@bangor.ac.uk; Robert A Harrison - r.harrison@liverpool.ac.uk; Wolfgang Wüster - w.wuster@bangor.ac.uk; Simon C Wagstaff\* - simonw@liv.ac.uk

\* Corresponding authors

Published: 30 November 2009

BMC Genomics 2009, 10:564 doi:10.1186/1471-2164-10-564

This article is available from: http://www.biomedcentral.com/1471-2164/10/564

© 2009 Casewell et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<u>http://creativecommons.org/licenses/by/2.0</u>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Received: 14 August 2009 Accepted: 30 November 2009

#### Abstract

**Background:** Venom variation occurs at all taxonomical levels and can impact significantly upon the clinical manifestations and efficacy of antivenom therapy following snakebite. Variation in snake venom composition is thought to be subject to strong natural selection as a result of adaptation towards specific diets. Members of the medically important genus *Echis* exhibit considerable variation in venom composition, which has been demonstrated to co-evolve with evolutionary shifts in diet. We adopt a venom gland transcriptome approach in order to investigate the diversity of toxins in the genus and elucidate the mechanisms which result in prey-specific adaptations of venom composition.

**Results:** Venom gland transcriptomes were created for *E. pyramidum leakeyi, E. coloratus* and *E. carinatus sochureki* by sequencing ~1000 expressed sequence tags from venom gland cDNA libraries. A standardised methodology allowed a comprehensive intra-genus comparison of the venom gland profiles to be undertaken, including the previously described *E. ocellatus* transcriptome. Blast annotation revealed the presence of snake venom metalloproteinases, C-type lectins, group II phopholipases  $A_2$ , serine proteases, L-amino oxidases and growth factors in all transcriptomes throughout the genus. Transcripts encoding disintegrins, cysteine-rich secretory proteins and hyaluronidases were obtained from at least one, but not all, species. A representative group of novel venom transcripts exhibiting similarity to lysosomal acid lipase were identified from the *E. coloratus* transcriptome, whilst novel metallopeptidases exhibiting similarity to neprilysin and dipeptidyl peptidase III were identified from *E. p. leakeyi* and *E. coloratus* respectively.

**Conclusion:** The comparison of *Echis* venom gland transcriptomes revealed substantial intrageneric venom variation in representations and cluster numbers of the most abundant venom toxin families. The expression profiles of established toxin groups exhibit little obvious association with venom-related adaptations to diet described from this genus. We suggest therefore that alterations in isoform diversity or transcript expression levels within the major venom protein families are likely to be responsible for prey specificity, rather than differences in the representation of entire toxin families or the recruitment of novel toxin families, although the recruitment of lysosomal acid lipase as a response to vertebrate feeding cannot be excluded. Evidence of marked intrageneric venom variation within the medically important genus *Echis* strongly advocates further investigations into the medical significance of venom variation in this genus and its impact upon antivenom therapy.

## Background

Snake venoms contain a complex mix of components, with biologically active proteins and peptides comprising the vast majority [1]. Variation in the composition of venom occurs at several taxonomical levels in multiple snake lineages [reviewed in [2,3]]. The view that variation in venom composition evolves primarily through neutral evolutionary processes [4-6] is not supported by other reports that snake venom composition is subject to strong natural selection as a result of adaptation towards specific diets [e.g. [7-10]]. Since the primary role of venom is to aid prey capture [2], it is perhaps unsurprising that variation in the protein composition of venom has been associated with significant dietary shifts in a number of genera [9-12]. Irrespective of the evolutionary forces underpinning venom protein composition, variation in venom components can significantly impact upon the clinical manifestations of snake envenoming [13-15] and, because the clinical efficacy of an antivenom may be largely restricted to the venom used in its manufacture, the success of antivenom therapy [16-18].

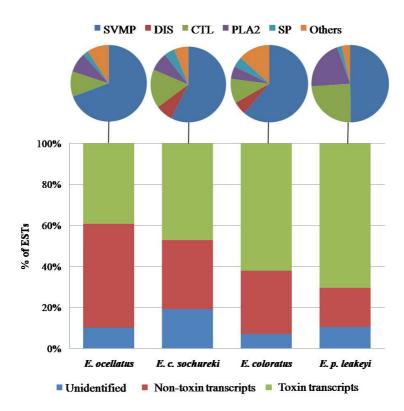
Envenoming by saw-scaled viper (Viperidae: Echis) species is thought to be responsible for more snakebite deaths worldwide than any other snake genus [19]. Envenomed victims typically suffer a combination of systemic and local haemorrhagic symptomatologies and up to 20% mortality rates without antivenom treatment [19-21]. Whilst the clinical symptoms are largely consistent throughout this widely distributed genus [20], cases of incomplete intrageneric antivenom efficacy have been documented, implying substantial inter-species venom variation [18,22-24]. We demonstrated that the four species complexes making up this genus, the E. carinatus, E. ocellatus, E. pyramidum and E. coloratus species groups [10,25], exhibit considerable vertebrate or invertebrate dietary preferences, E. coloratus being a vertebrate specialist whereas invertebrates feature prominently in the diet of the others. Since the proportions of consumed invertebrates correlated strongly with alterations in venom toxicity to scorpions, we believe the toxicity of the venom from these species to have co-evolved alongside evolutionary shifts in diet [10]. A preliminary venom protein analysis using reduced SDS-PAGE failed to identify an obvious link between venom composition and diet [10], justifying the use of a more comprehensive venom composition analysis in order to elucidate the mechanisms driving venom adaptations within the Echis viper genus.

Based on our earlier work with *E. ocellatus* [26], a comparative venom gland transcriptome approach was elected and we generated venom gland cDNA libraries from *E. coloratus, E. pyramidum leakeyi* and *E. carinatus sochureki*. Together with the existing *E. ocellatus* database, these provided DNA sequence data representing the venom gland transcriptomes for each of the four major species groups within the genus. The production of multiple *Echis* venom gland expressed sequence tag databases (vgDbEST) provides an unbiased overview of the transcriptional activity during venom synthesis in the venom glands of four species in this genus. This, the first comprehensive compilation of venom gland transcriptomes of congeneric snake species, was then interrogated to determine whether the mechanisms resulting in prey-specific adaptation of venom composition involve (i) the recruitment of novel prey-specific venom toxin transcripts, (ii) major changes in the expression levels of established toxin families, (iii) the diversification of functional isoforms within established toxin families or (iv) a combination of these factors.

## Results

EST data provides a powerful insight into the transcriptional activity of a tissue at a particular time point. Our protocols for the generation of venom gland EST databases provide a snapshot of transcriptional activity in the venom gland 3 days after venom expulsion, when transcription peaks [27] in preparation for new venom synthesis. Although each individual venom transcript cannot be correlated with the mature venom proteome without considerable extra experimental verification, our own work with E. ocellatus [28] shows there is a good general accordance between the venom proteome and that predicted from the venom gland transcriptome. Thus, whilst a cautionary approach is required when interpreting a correlation between transcriptome and proteome, the sensitivity and unbiased nature of venom gland transcriptome surveys can be valuable in the identification of rare, unusual or potentially novel toxins and their isoforms that are difficult to detect in the proteome [29].

To provide a representative overview of the transcriptional variation in venom components in each species, whilst minimising compositional bias arising from intraspecific variation in venom composition, venom gland cDNA libraries were based on ten specimens of variable size and gender. Generated ESTs were clustered under high stringency conditions to assemble overlapping single sequence reads into full length gene objects where possible. Using BLAST, 80-93% of gene objects for each library were assigned a functional annotation based upon significant (>1e-05) scores against multiple databases. The majority of annotated ESTs (61-74%) were assigned to clusters representing distinct gene objects (additional file 1). The proportion of toxin encoding transcripts (enzymes and nonenzymatic toxins) assigned by BLAST homology, was typically greater than those encoding non-toxin transcripts (for example, those involved in cellular biosynthetic processes) and unidentified components (i.e. with no significant hit against the databases) (Figure 1). There were twice



#### Figure I

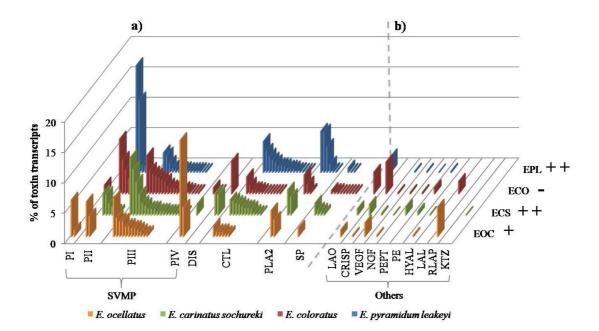
The relative expression of annotated venom gland transcriptomes from four members of the genus Echis. Bar charts represent the proportions of BLAST-annotated ESTs; unidentified = non-significant hits. Toxin encoding transcripts are expanded as pie charts illustrating the proportional representation of snake venom metalloproteinases (SVMP), short coding disintegrins (DIS), C-type lectins (CTL), group II phospholipases  $A_2$  (PLA<sub>2</sub>), serine proteases (SP) and other less represented venom toxins (Others) in the transcriptomes of each *Echis* species

the numbers of unidentified ESTs in the *E. c. sochureki* vgDbESTs than in any of the other *Echis* vgDbESTs. As the bulk of these unidentified ESTs were singletons, not clustered gene objects, we interpret this to result from increases in unidentified 3' untranslated regions rather than unidentified novel toxin transcripts. The annotated venom toxin encoding profiles for the four *Echis* species revealed substantial variation in (i) the inferred expression levels and (ii) the cluster diversity within many toxin families (Figure 2, additional file 2). The details and potential implications of this species-specific variation in the representation of each toxin family will be discussed in turn.

#### Snake venom metalloproteinases (SVMP)

The SVMP transcripts were the most abundant and divergent (in terms of cluster numbers) *Echis* venom toxin family (Figure 2) and comprised roughly half of the total toxin transcripts (Figure 1). The SVMPs are a diverse group of enzymes classified into those comprising only the metalloproteinase domain (PI) and those sequentially extended by a disintegrin domain (PII), a disintegrin-like and cysteine-rich domain (PIII) and the latter co-valently linked to C-type lectin-like components (PIV) [30]. Known and suspected modifications in domain structure are thought to account for the wide range of SVMP pathological activities, including haemorrhage, coagulopathy, fibrinolysis and prothrombin activation [30-32].

There were more PIII SVMP clusters in the genus *Echis* than any other toxin family clusters. The presence of apparent, extensive PIII SVMP gene diversification hints that evolutionary pressures are acting to increase the functional diversity of this SVMP group, highlighting their fundamental biological importance to the genus. In contrast, PI SVMP transcripts were present, albeit at low levels, only in the *E. coloratus* and *E. ocellatus* vgDbESTs. While the diversity of the PII SVMPs was substantially lower than that of the PIII SVMPs, their abundance differed between species. Thus, 80% of total *E. p. leakeyi* SVMP transcripts were PIIs (cluster EPL00005 comprised 38% of all SVMPs) and, although less numerically significant, 38% of the *E. coloratus* SVMPs were also PIIs. Despite intrageneric variation in abundance and diversity, analysis of PII contigu-



#### Figure 2

The relative abundance and diversity of each *Echis* genus venom toxin family. a) Relative expression levels of nonsingleton clusters of the most representative venom toxin families and b) Relative expression levels of total non-singleton clusters and singletons representing the less numerically represented venom toxin families (Others) are expressed as a percentage of total toxin encoding transcripts. Column to the right indicates the proportion of invertebrate prey consumed and the corresponding correlation of venom toxicity to scorpions: ++, high; +, moderate; -, low [adapted from [10]]. Key - PI-PIV: subclasses of snake venom metalloproteinases (SVMP); DIS: short coding disintegrins; CTL: C-type lectins; PLA2: group II phospholipases  $A_2$ ; SP: serine proteases; LAO: L-amino oxidases; CRISP: cysteine-rich secretory proteins; VEGF: vascular endothelial growth factors; NGF: nerve growth factors; PEPT: peptidases - aminopeptidase, dipeptidyl peptidase III and neprilysin; PE: Purine liberators - phosphdiesterase, 5'-nucleotidase and ectonucleoside triphosphate diphosphohydrolase (E-NTPase); HYAL: hyaluronidases; LAL: lysosomal acid lipases; RLAP: renin-like aspartic proteases; KTZ: kunitz-type protease inhibitors.

ous sequences throughout the genus revealed the ubiquitous representation of motifs (RGD, KGD and VGD) involved in binding to the  $\alpha_{IIb}\beta_3$ ,  $\alpha_v\beta_3$  and  $\alpha_5\beta_1$  integrins implicated in platelet aggregation inhibition [33,34]. The RGD-only representation of E. p. leakeyi PII SVMPs implies evolutionary conservation of this particular disintegrin motif, in contrast to the gene diversification observed in the PIIIs. We assigned some PIII SVMP transcripts as putative PIV SVMPs according to the presence of an additional cysteine residue in the cysteine-rich region at positions 397 or 400 [[28,30] (numbering from 30)]. These transcripts also form strongly supported monophyletic groups (data not shown) with homologues of SVMP PIVs previously characterised from venom proteomes; two of the three putative E. coloratus PIVs (ECO00075 & ECO00144) show the greatest sequence similarity to PIV SVMPs characterised from Macrovipera lebetina and Daboia russelii respectively [UniProt:Q7T046 and Q7LZ61], whereas all other Echis PIVs showed greatest similarity to the previously characterised E. ocellatus PIV SVMP, EOC00024 [28]. The relative representation of these putative PIV SVMPs was substantially greater in E.

ocellatus (EOC00024 - 23% and EOC00022 -7%) than *E.* coloratus and *E. c.* sochureki (<4%); no PIV SVMPs were found in the *E. p.* leakeyi vgDbEST. Taken together, this implies that two divergent forms of PIV SVMPs may be uniquely present in *E.* coloratus, despite their low representation in this species.

We (SCW, RAH) recently identified a new *E. ocellatus* cDNA precursor encoding numerous QKW tripeptides and a polyH/G peptide that have potent SVMP-inhibiting activities [35]. Representatives of this SVMP inhibitory transcript were identified in each *Echis* vgDbEST (data not shown), but no correlation was identified between the proportional representation of the *Echis* SVMPs and their SVMP inhibitory transcripts.

#### Disintegrins

Snake venom disintegrins are derived either from proteolytic processing of PII SVMP precursors [36] or are encoded by discreet PII-derived disintegrin-only genes, containing only a signal peptide and a disintegrin domain - previously described as 'short coding' disintegrins [37,38]. Representation of short coding disintegrins in the *Echis* genus is variable; small clusters were found in *E. c.* sochureki (4% and 3% of toxin transcripts) and *E. coloratus* (5%), whilst only a singleton transcript was found in *E. p.* leakeyi. Despite not being represented in the original *E. ocellatus* vgDbEST, we previously identified, by PCR, a sequence encoding the short coding disintegrin ocellatus from this species [39], confirming the presence of short coding disintegrin transcripts throughout the *Echis* genus.

## C-type lectins (CTL)

The CTLs proved to be the next most abundant and diverse (by cluster numbers) group of Echis venom toxin encoding transcripts. As argued for the SVMPs, the substantial CTL cluster diversity and implied functional diversity would be consistent with the known variation in CTL activity. Thus, CTL isoforms typically act synergistically as homologous or heterologous multimers to promote or inhibit platelet aggregation and/or target distinct elements of the coagulation cascade [see [40,41]]. Each of the Echis species showed considerable CTL diversity (10-24% toxin encoding transcripts), with E. p. leakeyi exhibiting both the largest number of ESTs and cluster-diversity. Notably, clusters showing similarity to echicetin  $\alpha$  and  $\beta$ , a platelet aggregation-inhibitor isolated from E. c. sochureki [42,43], were found throughout the Echis genus and are the most represented CTLs in both E. c. sochureki and E. p. leakevi. Recently, E. ocellatus echicetin-like CTLs were demonstrated to be associated with forming the quaternary structure of PIV E. ocellatus SVMPs [28]. However, PIV SVMPs are absent from the E. p. leakeyi vgDbEST and present in only small numbers in E. c. sochureki (2%), implying that PIV-related binding may not be the sole function of echicetin. In contrast, each of the Echis vgDbESTs (except for E. p. leakeyi) contained clusters showing high sequence similarity to another PIV-related CTL, Factor X activator light chain 2 from M. lebetina [44], producing an Echis representational profile of CTLs matching that of the PIV SVMPs.

## Phospholipase A<sub>2</sub> (PLA<sub>2</sub>)

Group II PLA<sub>2</sub>s are ubiquitously expressed in *Echis* species [45]. *Echis* PLA<sub>2</sub>s have been demonstrated to inhibit platelet aggregation and induce oedema, neurotoxicity and myotoxicity through multiple isoforms exhibiting high (Asp<sup>49</sup>) and low (Ser<sup>49</sup>) enzymatic activity [46-49]. Despite low representation and diversity in *E. coloratus, E. ocellatus* and *E. c. sochureki* (5-8% of toxin transcripts), an increase in representation (21%) and cluster diversity was observed in *E. p. leakeyi*, suggesting an important role for PLA<sub>2</sub> activity in the venom of this species. Furthermore, both enzymatic PLA<sub>2</sub> variants are conserved throughout the genus, highlighting the apparent importance of these functionally-distinct isoforms - presumably for prey capture. Given that Ser<sup>49</sup> PLA<sub>2</sub>s have only been isolated from the genera *Vipera* [50] and *Echis* [49], which are not sister taxa [51], we would expect the presence of this isoform in other members of the Viperinae. However, considering the absence of Ser<sup>49</sup> PLA<sub>2</sub>s from a *Bitis gabonica* vgDbEST [38], we cannot rule out convergent evolution of this myotoxic PLA<sub>2</sub> type and its consequent functional importance in these genera.

## Serine proteases (SP)

The snake venom serine proteases are a multi-gene enzyme family acting upon platelet aggregation, blood coagulation and fibrinolytic pathways [reviewed in [41]]. Considering the severe coagulopathy observed in victims of Echis envenoming [19,31], the SPs are represented in amounts lower than predicted (2-5% of toxin encoding transcripts), particularly given their high representation in other, albeit distantly related, Viperidae species [52,53]. Interestingly, variations in cluster diversity are considerable, with nine clusters of low representation identified in E. coloratus compared to one in E. ocellatus. Despite low levels of representation, the unique variation in cluster diversity observed in E. coloratus implies multiple gene duplication events within this lineage; a process that underpins functional diversification in multi-gene venom proteins [8,54].

#### L-amino oxidases (LAO)

Snake venom LAOs have been demonstrated to induce apoptosis and inhibit platelet function [reviewed in [55]]. While the mechanisms for these actions remain predominately uncharacterised, it seems clear that, unlike other snake venom toxin families, isoform diversity is not a requirement. Thus, the low representation (1-4% of toxin transcripts) observed in the *Echis* vgDbESTs is consistent with other viperid venom gland transcriptomes [26,38,52,53,56-59]. Indeed, the atypically high level of sequence conservation between all the *Echis* LAOs and those from other viperid genera (>80%) implies a conserved mechanism of action, whereby evolutionary pressures act to constrain diversification.

#### Cysteine-rich secretory proteins (CRISP)

Members of the snake venom CRISP family interact with ion channels and exhibit the potential to block arterial smooth muscle contraction and nicotinic acetylcholine receptors [e.g. [60,61]]. The relative CRISP expression profiles vary considerably in the genus *Echis*, ranging from 5% of toxin encoding transcripts in *E. coloratus*, less than 2% in *E. c. sochureki* and *E. ocellatus* and none in *E. p. leakeyi*. Given that CRISPs are typically underrepresented toxin transcripts in Viperidae vgDbESTs [26,38,52,56-59], the abundant representation observed in *E. coloratus* implies an unidentified evolutionary pressure favouring transcriptional expression in this species. Its potential biological significance is further highlighted by the apparent absence of these toxins in the transcriptome of the most closely related species, *E. p. leakeyi*, which differs strongly in diet from *E. coloratus* [10].

#### Other toxin components

Clusters encoding vascular endothelial growth factors and nerve growth factors were identified in small numbers (additional file 2) throughout the genus and, like the LAOs, each showed a high degree of sequence conservation. Similarly, and consistent with previous reports [62], the sequence homology of the new hyaluronidase singleton ESTs of E. c. sochureki and E. ocellatus was also considerable, and extended to hylauronidase sequences of other genera. It is apparent that evolutionary forces exist to conserve the sequence of this group of venom proteins, presumably because their role in disseminating venom toxins by reducing the viscosity of the extracellular matrix [29] is a universal requirement for prey 'knock-down'. Another singleton EST from the E. c. sochureki vgDbEST exhibited 81% identity to a kunitz-type protease inhibitor isolated from the elapid snake Austrelaps labialis [63]. Given the phylogenetic distance between these species, homology between these haemostatic disruptors is surprising, particularly since the singleton exhibited only 38% identity to kunitz-type protease inhibitors identified from the Bitis gabonica vgDbEST [38], a species closely related to Echis. An additional number of peptidases and purine liberators were identified as minor components in all but the E. ocellatus vgDbEST (Table 1). Despite their low representation and inconsistent conservation throughout the genus, the distinct biological activities of these components have been reported to play a role in the pathology of viper envenoming (Table 1), although these claims require experimental confirmation.

## Novel venom gland transcriptome components

We identified a cluster from the *E. coloratus* vgDbEST that exhibited 64% identity to mammalian lysosomal acid lipase/cholesteryl ester hydrolase (LAL) [Uni-Prot:Q4R4S5]. The most critical function of LAL is to modulate intracellular cholesterol metabolism by degrading cholesterol esters and triglycerides derived from low density lipoproteins that are transported, via specific receptors, into most cells [64,65]. Although LAL is a common enzyme in many lineages, this is the first time it has been identified from a venomous animal. We interrogated the vgDbESTs for other transcripts with annotations related to lysosomal processes and singleton transcripts were identified in multiple species (data not shown). However, their quantities were considerably lower than LAL suggesting to us that an association between venom gland LAL and intracellular processes was unlikely. Furthermore, the identification of a signal peptide using SignalP v3.0 [66] and the comparable representation of this

enzyme (2%) with other venom toxin encoding transcripts (e.g. SPs, LAOs, growth factors), strongly implies these transcripts are a novel group of secreted venom components. Their biological contribution to the activity of *E. coloratus* venom and the venom gland and expression in other venomous snake genera is the subject of current research in our laboratories.

In addition to the discovery of LAL, two singleton transcripts were identified (additional file 2) from the Echis vgDbESTs as novel Serpentes zinc-dependent metallopeptidases [67]. A transcript exhibiting 67% identity to human dipeptidyl peptidase III (DPPIII) [Uni-Prot:Q53GT4] was identified in E. coloratus and a related EST exhibiting 84% similarity to Neprilysin from Gallus gallus [Uniprot:Q67BJ2] was identified in the E. p. leakeyi vgDbEST. While signal peptides were absent from these ESTs due to EST N-terminal truncation, the constitutive physiological targets of their mammalian analogues indicate that these metallopeptidases may contribute to pathology. Mammalian DPPIII exhibits particular affinity for the degradation of hypertension-inducing peptides via the inactivation and degradation of angiotensin II to angiotensin III; the consequential reduction in vasoconstrictor activity likely induces hypotension alongside thrombolysis, by reducing the activity of plasminogen activator inhibitors that constrain fibrinolysis [68-70]. We previously reported that the E. ocellatus vgDbEST contained a substantial number of novel, potentially hypotensive, venom toxins termed the renin-like aspartic proteases [26]. Neprilysin demonstrates affinity for a broader range of physiological targets, including natriuretic, vasodilatory and neuro peptides [71]. Specific functional interactions include the termination of brain neuropeptides, such as enkephalins and substance P, at peptidergic synapses [72], and the degradation of the hypotension-inducing atrial natriuretic peptide (ANP) [71]. It is notable that Neprilysin has been implicated in the inactivation of peptide transmitters and their modulators in vertebrates and invertebrates [71,73], suggesting the potential for conserved neurotoxic activity across a range of prey species.

## Discussion

The most numerically abundant venom toxin families in the four *Echis* species were the SVMPs, CTLs, PLA<sub>2</sub>s, and SPs. This is broadly consistent with previous viperid venom gland analyses, although considerable intergeneric variations in the EST-inferred expression levels of these toxin families have been observed [26,38,52,53,56-59]. The correlation of toxin families identified from the genus *Echis* and other viperid species support current theories of early venom toxin recruitment prior to the radiation of the Viperidae [74]. The absence of three finger toxins from the *Echis* vgDbESTs is particularly notable as their recent identification in other viper species [53,58]

Identification	No. of ESTs	Species present	Activty	Possible venom function
Aminopeptidase	8	E. c. sochureki	Hydrolysis of the N-terminal region of peptides [82].	Potential interference with angiogenesis and blood pressure control [83,84].
	I	E. coloratus		
Ectonucletotide pyrophosphatase/ phosphodiesterase	2	E. coloratus	Hydrolysis of nucleotides and nucleic acids [85].	Interaction with platelet function [85]. Activity previously described in <i>Echis</i> <i>carinatus</i> [86].
	3	E. c. sochureki		
5'-nucleotidase	3	E. coloratus	Cleavage of a wide variety of ribose and deoxyribose nucleotides [1].	Potential inhibitor of platelet aggregation [1]. Activity identified in a number of different lineages including <i>Echis carinatus</i> [86].
	2	E. p. leakeyi		
	I	E. c. sochureki		
Ectonucleoside triphosphate diphosphohydrolase 2 (E-NTPase 2)	2	E. coloratus	Hydrolysis of nucleoside-5'- triphosphates and diphosphates [87].	Potential inhibitor of platelet aggregation [87,88].

Table I: Under-represented toxin encoding transcripts from the Echis vgDbESTs potentially associated with venom function.

implies the venom gland recruitment of these toxins occurred prior to the divergence of the Viperidae; presumably these toxins have subsequently been lost in an ancestor of Echis. Consistent with the early, PCR-driven, reports of accelerated evolution of venom serine proteases [75], CTLs [76] and PLA<sub>2</sub>s [77], it is apparent from the Echis genus vgDbESTs and those of other vipers that the evolutionary forces driving venom toxin recruitment in the genus Echis have served to promote diversification in some toxin lineages (PII and PIII SVMPs, CTLs) while in comparison relatively low diversification exists in others (PI and PIV SVMPs, PLA<sub>2</sub>s, LAOs, the growth factors, and remaining minor venom components). Prey capture is considered a major biological imperative driving the venom toxin selection process. This project was undertaken to identify correlations between intrageneric dietary preferences and transcript expression in order to elucidate the influence dietary selection pressures may have on the toxin composition of snake venoms.

(i) Recruitment of novel venom toxins and diet. The *Echis* vgDbESTs reveal the recruitment of novel renin-like aspartic proteases in *E. ocellatus* [26], LAL and DPPIII in *E. coloratus* and Neprilysin in *E. p. leakeyi*. The potential hypotensive role of venom aspartic proteases has been discussed previously [26]. Whilst expression in the venom proteome requires experimental verification, the presence of a signal peptide suggests that LAL is more likely to be secreted in the venom gland rather than acting as an intra-

cellular protein. LAL has been implicated in severe alveolar destruction following over-expression of these enzymes in the lungs of mice [64]. Lipases such as LAL and lipoprotein lipase may also contribute to an influx of fatty acids into the brain by hydrolysing lipoproteins in the microvascular system of the cerebral cortex [78]. The suggestion that these fatty acids are then intra-cellularly internalised within lysosomes [78] correlates with intriguing observations from E. coloratus induced pathology, where increases in the size and numbers of lysosomes within the neuronal tissue of guinea pigs were implicated in neuron lysis and cerebral damage [79]. We infer from the predominately vertebrate-only diet of E. coloratus and the exclusive, yet substantial, representation of LAL in this species (2% - equivalent to the SPs, LAOs and growth factors) that LALs may play a contributory, albeit not yet understood, role in prey envenoming. As singletons, it is more difficult to argue that the novel recruitments of DPPIII and Neprilysin represent additional adaptations to prey preference; as they are found in such low numbers it is impossible to determine whether they are indeed novel species-specific venom gland recruitments or are rare transcripts that remain undetected in other snake species. We previously reported that invertebrate feeding likely evolved as a basal trait in the genus Echis [10]. The absence of genus-wide transcripts encoding novel putative venom toxin families implies that the adaptation to invertebrate feeding in Echis did not evolve as a consequence of recruiting novel invertebrate-specific venom toxins. However,

we cannot exclude the possibility that the novel recruitment of LAL into the *E. coloratus* venom gland transcriptome may result from the subsequent reversion to vertebrate feeding observed in this species [10], particularly given the absence of these well represented putative toxin transcripts in other members of the genus.

(ii) Changes in toxin family expression and diet. All the major *Echis* venom toxin families (SVMP, CTL, PLA<sub>2</sub>, SP) exhibited considerable intrageneric variation in transcriptional representation. Thus, the E. p. leakeyi vgDbEST was notable for its absence of PI and PIV SVMPs, short coding disintegrins and CRISPs and atypically abundant representation of PII SVMPs, CTLs and PLA2s. The CRISPs were only represented by clusters in E. c. sochureki and E. coloratus, species whose vgDbESTs draw similarities, particularly in their high comparative expression of PIII SVMPs and short coding disintegrins. The only distinguishing feature (in terms of transcript abundance) in the E. ocellatus vgDbEST was the atypically high number of PIV SVMPs. However, none of these toxin encoding expression profiles showed a clear association with diet. Most notably, E. p. leakeyi and E. c. sochureki exhibit distinct toxin encoding profiles (Figure 2), despite both species feeding predominately on invertebrates and exhibiting highly invertebratelethal venom [10].

(iii) Diversification of venom toxins and diet. The above observations imply adaptations to diet are occurring within venom toxin families rather than resulting from changes in expression levels of entire toxin families. Evidence supporting this hypothesis is provided by substantial increases in representation of echicetin-like CTLs (relative to other CTLs) in both E. p. leakeyi and E. c. sochureki, implying perhaps a significant role for these platelet aggregation inhibitors in invertebrate prey capture. The absence of PI SVMPs in these species perhaps suggests that this SVMP isoform is more associated with a vertebrate diet. Furthermore, a number of atypical observations identified from the E. coloratus vgDbEST may be associated with a reversion to vertebrate feeding [10], including; (i) increases in the representation of CRISPs, (ii) increases in cluster diversity of the SPs and (iii) the identification of putative novel venom toxins (LAL and DPPIII). However, the general similarity between the toxin encoding expression profiles of E. c. sochureki and E. coloratus (Figure 2), despite E. coloratus exhibiting a significant reduction in venom toxicity to invertebrates [10], indicates that more analytical molecular tools are required to determine whether snake prey specificity is achieved through subtle alterations in isoform expression levels within the major venom toxin families. We are subjecting the Echis genus vgDbEST data generated here to a phylogenetic analysis on each toxin class to determine species-specific trends in diversification, which will inform us whether multiple levels of gene control in the *Echis* genus venom gland (switching of transcriptional expression, gene duplication conferring functional diversification and novel gene expression) maybe responsible for evolutionary responses to dietary pressures.

Correlations between variation in venom gland toxin encoding profiles and snakebite symptomatologies from the genus *Echis* are unclear, particularly given the similar, predominately incoagulable and haemorrhagic, clinical outcomes observed throughout the genus [19-21] and the presence of multiple isoforms of toxin families implicated in haemorrhage and coagulopathy. However, some observations of atypical symptoms can be tentatively explained; substantial increases in PLA<sub>2</sub> representation and the unique presence of Neprilysin may correlate with the rare manifestation of neurotoxicity observed in an *E. pyramidum* envenomation [22], whilst the putative function of DPPIII may imply a contributory role in cases of hypotension observed following *E. coloratus* snakebite [20].

Venom gland transcriptome surveys provide valuable new data that we are correlating with a proteomic analysis of the venom from each *Echis* species. With this comprehensive description of the venom composition of each major *Echis* lineage, we will identify, using proteomic (antivenomic) techniques [3], the extent to which the intrageneric variation in venom composition impacts on the preclinical efficacy of commercially available antivenoms. We hope that such analyses will (i) explain past antivenom failures described following snakebite by members of this medically important genus [18,22-24] and (ii) identify the venom toxin mix required to generate an antivenom with continent-wide clinical effectiveness against *Echis* envenoming.

## Conclusion

The first comprehensive comparison of intrageneric venom gland transcriptomes reveals substantial venom variation in the genus *Echis*. The observed variations in venom toxin encoding profiles reveal little association with venom adaptations to diet previously described from this genus. We hypothesise that relatively subtle alterations in toxin expression levels within the major venom toxin families are likely to be predominately responsible for prey specificity, although we cannot rule out a contributory role for novel putative venom toxins, such as lysosomal acid lipase. The observation of substantial venom variation within the medically important genus *Echis* strongly advocates further investigations into the medical significance of venom variation and its potential impact upon antivenom therapy.

## Methods

Venom gland cDNA libraries were constructed from ten wild-caught specimens of *Echis coloratus* (Egypt), *E. p. leakeyi* (Kenya) and *E. c. sochureki* (Sharjah, UAE), maintained in the herpetarium of the Liverpool School of Tropical Medicine, using identical protocols described for the construction of the venom gland cDNA library from *E. ocellatus* [26]. Clones from the cDNA libraries were picked randomly and sequenced (NERC Molecular Genetics Facility, UK) using M13 forward primers.

Bioinformatic processing was carried out using the Parti-Gene pipeline [80] with the same protocols used previously [26]. Briefly, sequences were processed (to exclude low quality, contaminating vector sequences and poly A+ tracts) using Trace2dbEST [81]. Subsequently, assembly was undertaken in PartiGene version 3.0, using high stringency clustering parameters [26,81]. A total of 1070 (*E. coloratus*), 1078 (*E. p. leakeyi*) and 1156 (*E. c. sochureki*) processed ESTs were entered into respective species databases alongside the 883 ESTs generated from the *E. ocellatus* vgDbEST [26]. Assembled ESTs were BLAST annotated against UniProt (v56.2), TrEMBL (v39.2) and separate databases containing only Serpentes nucleotide and protein sequences derived from the same Uniprot/TrEMBL release versions.

Clustering was performed incrementally (96 sequences per round) to determine the number of sequences required to construct a representative transcriptome (i.e. the point where further sequencing only adds to existing clusters). We estimate that a minimum of 800 EST sequences were required to provide an accurate representation of the three vgDbESTs (additional file 3). For longer clones (i.e. SVMPs), representatives of each cluster were subject to primer walking to acquire sufficient sequence data for isoform classification. SVMPs were characterised based upon the presence or absence of additional domains extending from the metalloproteinase domain [30]. PIVs were distinguished from PIIIs by the presence of an additional cysteine residue in the cysteine-rich region at positions 397 or 400 [[28,30] (numbering from 30)].

Additional file 2 displays the catalogue of venom toxin transcripts present in each of the four *Echis* vgDbESTs based upon significant (>1e-05) BLAST annotation. Presentation of the fully assembled and annotated vgDbESTs can be viewed at <u>http://venoms.liv.ac.uk</u>. The sequences reported in this paper have also been submitted into dbEST division of the public database GenBank: *E. coloratus* [GenBank: <u>GR947900-GR948969</u>], *E. c. sochureki* [GenBank: <u>GR948970-GR950126</u>] and *E. p. leakeyi* [GenBank: <u>GR950127-GR951204</u>].

All animal experimentation was conducted using standard protocols approved by the University of Liverpool Animal Welfare Committee and performed with the approval of the UK Home Office (40/3216) under project licence # 40/3216.

## **Authors' contributions**

NRC participated in the experiments, the comparative analysis and drafted the manuscript. RAH participated in the experiments, the design of the study and reviewed the manuscript. WW participated in the design of the study and reviewed the manuscript. SCW participated in the experiments, the design of the study, the comparative analysis and reviewed the manuscript. All authors have read and approved the paper.

## Additional material

#### Additional file 1

Summary statistics following clustering and assembling of ESTs for E. coloratus, E. p. leakeyi and E. c. sochureki. Click here for file [http://www.biomedcentral.com/content/supplementary/1471-

2164-10-564-\$1.doc]

## Additional file 2

Catalogue of venom toxin encoding ESTs determined from the Echis vgDbESTs. Putative novel venom toxins are in bold and underlined. Key - SVMP: snake venom metalloproteinases; PI, PII, PIII, PIV: respective sub-group of SVMPs; ND: sub-class not determined; DIS: short coding disintegrins; CTL: C-type lectins; PLA<sub>2</sub>: group II phospholipases A<sub>2</sub>; SP: serine proteases; LAO: L-amino oxidases; CRISP: cysteine-rich secretory proteins; VEGF: vascular endothelial growth factors; NGF: nerve growth factors; PEPT: peptidases; AP: aminopeptidase; DPP: dipeptidyl peptidase III; NEP: neprilysin; PE: Purine liberators; PHOS: phosphdiesterase; 5'-NUC: 5'-nucleotidase; E-NTPase: ectonucleoside triphosphate diphosphohydrolase; LAL: lysosomal acid lipases; RLAP: renin-like aspartic proteases; HYAL: hyaluronidases; KTZ: kunitz-type protease inhibitors. Click here for file

[http://www.biomedcentral.com/content/supplementary/1471-2164-10-564-S2.doc]

# Additional file 3

An overview of clustering processes for three species of the genus Echis. The graph demonstrates the percentage of ESTs that are added to clusters (ESTs >1) as the cumulative number of ESTs entering the database increase. In all species the number of ESTs affecting the proportion of EST clusters and singletons reaches a plateau after 800 sequences. Click here for file

[http://www.biomedcentral.com/content/supplementary/1471-2164-10-564-S3.jpeg]

#### Acknowledgements

The authors wish to thank Paul Rowley for expert herpetological assistance, Damien Egan and Paul Vercammen (Breeding Centre for Endangered Arabian Wildlife, United Arab Emirates) for providing specimens of *E. c. sochureki*, Ann Hedley and Mark Blaxter (NERC Molecular Genetics Facility, University of Edinburgh) for providing sequencing and bioinformatic advice regarding the PartiGene pipeline and Tim Booth, Bela Tiwari and Jorge Soares (NERC Environmental Bioinformatics Centre) for bioinformatic advice. This work was funded by Research Studentship NER/S/A/2006/ 14086 from the Natural Environmental Research Council (NERC) to NRC, access to the NERC Molecular Genetics Facility at the University of Edinburgh (ref MGF 150) to WW, the Leverhulme Trust (Grant F/00 174/I) to WW and RH and the Biotechnology and Biological Sciences Research Council (BBSRC) to RH and SCW (BB/F012675/I).

#### References

- 1. Aird SD: **Ophidian envenomation strategies and the role of purines.** *Toxicon* 2002, **40:**335-393.
- Chippaux JP, Williams V, White J: Snake venom variability: methods of study, results and interpretation. Toxicon 1991, 29:1279-1303.
- Gutiérrez JM, Lomonte B, Leün G, Alape-Girün A, Flores-Díaz M, Sanz L, Angulo Y, Calvete JJ: Snake venomics and antivenomics: proteomic tools in the design and control of antivenoms for the treatment of snakebite envenoming. J Proteomics 2009, 72:165-182.
- Sasa M: Diet and snake venom evolution: can local selection alone explain intraspecific venom variation? *Toxicon* 1999, 37:249-252.
- 5. Sasa M: Reply. Toxicon 1999, 37:259-260.
- 6. Mebs D: Toxicity in animals. Trends in evolution? *Toxicon* 2001, **39:**87-96.
- Daltry JC, Wüster W, Thorpe RS: Diet and snake venom evolution. Nature 1996, 379:537-540.
- 8. Kordiš D, Gubenšek F: Adaptive evolution of animal toxin multigene families. *Gene* 2000, **261**:43-52.
- Jorge da Silva N Jr, Aird SD: Prey specificity, comparative lethality and compositional differences of coral snake venoms. Comp Biochem Physiol 2001, 128C:425-456.
- Barlow A, Pook CE, Harrison RA, Wüster W: Co-evolution of diet and prey-specific venom activity supports the role of selection in snake venom evolution. Proc R Soc B 2009, 276:2443-2449.
- Creer S, Malhotra A, Thorpe RS, Stöcklin R, Favreau P, Chou WH: Genetic and ecological correlates of intraspecific variation in pitviper venom composition detected using matrix-assisted laser desorption time-of-flight mass spectrometry (MALDI-TOF-MS) and isoelectric focusing. *J Mol Evol* 2003, 56:317-329.
- Sanz L, Gibbs HL, Mackessy SP, Calvete JJ: Venom proteomes of closely related Sistrurus rattlesnakes with divergent diets. J Proteome Res 2006, 5:2098-2112.
- Warrell DA: Snake venoms in science and clinical medicine. I. Russell's viper: biology, venom and treatment of bites. Trans R Soc Trop Med Hyg 1989, 83:732-740.
- Prasad NB, Uma B, Bhat SKG, Gowda TV: Comparative characterization of Russell's viper (Daboia/Vipera russelli) venoms from different regions of Indian peninsula. Biochim Biophys Acta 1999, 1428:121-136.
- Shashidharamurthy R, Jagadeesha DK, Girish KS, Kemparaju K: Variation in biochemical and pharmacological properties of Indian cobra (Naja naja) venom due to geographical distribution. Mol Cell Biochem 229:93-101.
- Theakston RDG, Phillips RE, Warrell DA, Galigedera Y, Abeysekera DT, Dissanayake P, Hutton RA, Aloysius DJ: Failure of India (Haffkine) antivenom in treatment of Vipera russelli pulchella (Russell's viper) envenoming in Sri Lanka. Toxicon 1989, 27:82.
- Galán JA, Sánchez EE, Rodríguez-Acosta A, Pérez JC: Neutralization of venoms from two Southern Pacific rattlesnakes (Crotalus helleri) with commercial antivenoms and endothermic animal sera. Toxicon 2004, 43:791-799.
- Visser LE, Kyei-Faried S, Belcher DW, Geelhoed DW, Schagen van Leeuwen J, van Roosmalen J: Failure of a new antivenom to treat Echis ocellatus snake bite in rural Ghana: the importance of quality surveillance. Trans R Soc Trop Med Hyg 2008, 102:445-450.
- Warrell DA, Davidson NM, Greenwood BM, Ormerod LD, Pope HM, Watkins BJ, Prentice CRM: Poisoning by bites of the sawscaled or carpet viper (Echis carinatus) in Nigeria. QJM 1977, 46:33-62.

- Warrell DA: Clinical toxicology of snakebite in Africa, the Middle East/Arabian Peninsula and Asia. In Handbook of clinical toxicology of animal venoms and poisons Edited by: Meier J, White J. Boca Raton, Florida: CRC Press; 1995:433-595.
- Habib AG, Gebi UI, Onyemelukwe GC: Snake bite in Nigeria. Afr J Med Med Sci 2001, 30:171-178.
- 22. Gillissen A, Theakston RDG, Barth J, May B, Krieg M, Warrell DA: Neurotoxicity, haemostatic disturbances and haemolytic anaemia after a bite by a Tunisian saw-scaled or carpet viper (Echis 'pyramidum'-complex): Failure of antivenom treatment. Toxicon 1994, 32:937-944.
- Kochar DK, Tanwar PD, Norris RL, Sabir M, Nayak KC, Agrawal TD, Purohit VP, Kochar A, Simpson ID: Rediscovery of severe sawscaled viper (Echis sochureki) envenoming in the Thar Desert region of Rajasthan, India. Wilderness Enviro Med 2007, 18:75-85.
- 24. Warrell DA: Unscrupulous marketing of snake bite antivenoms in Africa and Papua New Guinea: choosing the right product-'What's in a name?'. Trans Royal Soc Trop Med Hygiene 2008, 102(5):397-399.
- Pook CE, Joger U, Stümpel N, Wüster W: When continents collide: phylogeny, historical biogeography and systematics of the medically important viper genus Echis (Squamata: Serpentes: Viperidae). Mol Phylogenet Evol 2009 in press. doi:10.1016/ j.ympev.2009.08.002
- j.ympev.2009.08.002
  Wagstaff SC, Harrison RA: Venom gland EST analysis of the saw-scaled viper, Echis ocellatus, reveals novel α<sub>9</sub>β<sub>1</sub> integrinbinding motifs in venom metalloproteinases and a new group of putative toxins, renin-like aspartic proteases. Gene 2006, 377:21-32.
- Paine MJ, Desmond HP, Theakston RDG, Crampton JM: Gene expression in *Echis carinatus* (carpet viper) venom glands following milking. *Toxicon* 1992, 30:379-386.
- Wagstaff SC, Sanz L, Juárez P, Harrison RA, Calvete JJ: Combined snake venomics and venom gland transcriptomic analysis of the ocellated carpet viper, Echis ocellatus. J Proteomics 2009, 71(6):609-623.
- Harrison RA, Ibison F, Wilbraham D, Wagstaff SC: Identification of cDNAs encoding viper venom hyaluronidases: cross-generic sequence conservation of full-length and unusually short variant transcripts. Gene 2007, 392:22-33.
- Fox JW, Serrano SMT: Structural considerations of the snake venom metalloproteinases, key members of the M12 reprolysin family of metalloproteinases. *Toxicon* 2005, 45:969-985.
- Warrell DA, Pope HM, Prentice CRM: Disseminated intravascular coagulation caused by the carpet viper (Echis carinatus): Trial of Heparin. Brit J Haemat 1976, 33:335-342.
- Fox JW, Serrano SMT: Insights into and speculations about snake venom metalloproteinase (SVMP) synthesis, folding and disulfide bond formation and their contribution to venom complexity. FEBS J 2008, 275:3016-3030.
- Huang TF, Holt JC, Lukasiewicz H, Niewiarowski S: Trigamin. A low molecular weight peptide inhibiting fibrinogen with platelet receptors expressed on glycoprotein IIb-IIIa complex. J Biol Chem 1987, 262:16157-16163.
   Calvete JJ, Marcinkiewicz C, Monleün D, Esteve V, Celda B, Juárez P,
- Calvete JJ, Marcinkiewicz C, Monleün D, Esteve V, Celda B, Juárez P, Sanz L: Snake venom disintegrins: evolution of structure and function. *Toxicon* 2005, 45:1063-1074.
- Wagstaff SC, Favreau P, Cheneval O, Laing GD, Wilkinson MC, Miller RL, Stöcklin R, Harrison RA: Molecular characterisation of endogenous snake venom metalloproteinase inhibitors. Biochem Biophys Res Commun 2008, 365(4):650-656.
- Shimokawa K, Jai LG, Wang XM, Fox JW: Expression, activation and processing of the recombinant snake venom metalloproteinase, pro-atrolysin E. Arch Biochem Biophys 1996, 335:283-294.
- 37. Okuda D, Koike H, Morita T: A new gene structure of the disintegrin family: a subunit of dimeric disintegrin has a short coding region. *Biochemistry* 2002, 41:14248-14254.
- Francischetti IMB, My-Pharm V, Harrison J, Garfield MK, Ribeiro JMC: Bitis gabonica (Gaboon viper) snake venom gland: toward a catalog for the full-length transcripts (cDNA) and proteins. Gene 2004, 357:55-69.
- Juárez P, Wagstaff SC, Sanz L, Harrison RA, Calvete JJ: Molecular cloning of Echis ocellatus disintegrins reveals non-venom secreted proteins and a pathway for the evolution of ocellatusin. J Mol Evol 2006, 63:183-193.

- 40. Markland FS: Snake venoms and the haemostatic system. Toxicon 1998, 36:1749-1800.
- 41. Kini RM: Anticoagulant proteins from snake venoms: structure, function and mechanism. *Biochem J* 2006, **397:**377-387.
- Peng M, Lu W, Beviglia V, Niewiarowski S, Kirby EP: Echicetin: a snake venom protein that inhibits binding of von Willebrand factor and alboaggregins to platelet glycoprotein Ib. Blood 1993, 81:2321-2328.
- Polgár J, Magnenat EM, Peitsch MC, Wells TN, Saqi MS, Clemetson KJ: Amino acid sequence of the alpha subunit and computer modelling of the alpha and beta subunits of echicetin from the venom of Echis carinatus (saw-scaled viper). Biochem J 1997, 323:533-537.
- Siigur E, Aaspõllu A, Trummal K, Tõnismägi K, Tammiste I, Kalkkinen N, Siigur J: Factor X activator from Vipera lebetina venom is synthesized from different genes. Biochim Biophys Acta Prot Proteomics 2004, 1702:41-51.
- Bharati K, Hasson SS, Oliver J, Laing GD, Theakston RDG, Harrison RA: Molecular cloning of phospholipases A<sub>2</sub> from venom glands of Echis carpet vipers. Toxicon 2003, 41:941-947.
- Kemparaju K, Prasad BN, Gowda VT: Purification of a basic phospholipase A<sub>2</sub> from Indian saw-scaled viper (Echis carinatus) venom: characterization of antigenic, catalytic and pharmacological properties. *Toxicon* 1994, 32:1187-1196.
- Kemparaju K, Krishnakanth TP, Gowda VT: Purification and characterization of a platelet aggregation inhibitor acidic phospholipase A<sub>2</sub> from Indian saw-scaled viper (Echis carinatus) venom. Toxicon 1999, 37:1659-1671.
- Jasti J, Paramasivam M, Srinivasan A, Singh TP: Structure of an acidic phospholipase A<sub>2</sub> from Indian saw-scaled viper (Echis carinatus) at 2.6 Å resolution reveals a novel intermolecular interaction. Acta Cryst 2004, D60:66-72.
- Zhou X, Tan TC, Valiyaveettil S, Go ML, Kini RM, Velazquez-Campoy A, Sivaraman J: Structural characterization of myotoxic Ecarpholin S from Echis carinatus venom. Biophysical J 2008, 95:3366-3380.
- Petan T, Križaj I, Punger.ar J: Restoration of enzymatic activity in a Ser-49 phospholipase A2 homologue decreases its Ca2+independant membrane-damaging activity and increases its toxicity. *Biochemistry* 2007, 46:12795-12809.
- Wüster W, Peppin L, Pook CE, Walker DE: A nesting of vipers: Phylogeny and historical biogeography of the Viperidae (Squamata: Serpentes). Mol Phylogenet Evol 2008, 49(2):445-459.
- Cidade DAP, Simão TÁ, Dávila ÁMR, Wagner G, Junqueira-de-Azevedo ILM, Ho PL, Bon C, Zingali RB, Albano RM: Bothrops jararaca venom gland transcriptome: Analysis of the gene expression pattern. Toxicon 2006, 48:437-461.
- Pahari S, Mackessy SP, Kini RM: The venom gland transcriptome of the Desert Massasauga rattlesnake (Sistrurus catenatus edwardsii): towards an understanding of venom composition among advanced snakes (Superfamily Colubroidea). BMC Mol Biol 2007, 8:115.
- Župunski V, Kordiš D, Gubenšek F: Adaptive evolution in the snake venom Kunitz/BPTI protein family. FEBS Letters 2003, 547:131-136.
- 55. Du XY, Clemetson KJ: Snake venom L-amino acid oxidases. Toxicon 2002, 40:659-665.
- 56. Junqueira-de-Azevedo ILM, Ho PL: A survey of gene expression and diversity in the venom glands of the pit viper snake Bothrops insularis through the generation of expressed sequence tags (ESTs). Gene 2002, 299:279-291.
- 57. Kashima S, Roberto PG, Soares AM, Astolfi-Filho S, Pereira JO, Giuliati S, Faria M, Xavier MAS, Fontes MRM, Giglio JR, Franca SC: Analysis of Bothrops jararacussu venomous gland transcriptome focusing on structural and functional aspects: I - gene expression profile of highly expressed phospholipases A<sub>2</sub>. Biochimie 2004, 86:211-219.
- Junqueira-de-Azevedo ILM, Ching ATC, Carvalho E, Faria F, Nishiyama ML, Ho PL, Diniz MRV: *Lachesis muta* (Viperidae) cDNAs reveal diverging pit viper molecules and scaffolds typical of Cobra (Elapidae) venoms: Implications for snake toxin repertoire evolution. *Genetics* 2006, 173:877-889.
- 59. Zhang B, Liu Q, Yin W, Zhang X, Huang Y, Luo Y, Qiu P, Su X, Yu J, Hu S, Yan G: Transcriptomic analysis of Deinagkistrodon acutus venomous gland focusing on cellular structure and functional

aspects using expressed sequence tags. BMC Genomics 2006, 7:152.

- 60. Yamazaki Y, Morita T: Structure and function of snake venom cysteine-rich secretory proteins. *Toxicon* 2004, **44**:227-231.
- Gorbacheva EV, Starkov VG, Tsetlin VI, Utkin YN, Vulfius CA: Viperidae snake venoms block nicotinic acetylcholine receptors and voltage-gated Ca2+ channels in identified neurons of fresh-water snail Lymnaea stagnalis. Biochem (Moscow) A Membrane Cell Biol 2008, 2:14-18.
- 62. Kemparaju K, Girish KS: **Snake venom hyaluronidase:** a therapeutic target. *Cell Biochem Funct* 2006, **24:**7-12.
- 63. Doley R, Tram NNB, Reza MA, Kini RM: Unusual accelerated rate of deletions and insertions in toxin genes in the venom glands of the pygmy copperhead (Austrelaps labialis) from Kangaroo Island. BMC Evol Biol 2008, 8:70.
- Li Y, Qin Y, Li H, Wu R, Yan C, Du H: Lysosomal acid lipase overexpression disrupts lamellar body genesis and alveolar structure in the lung. Int J Exp Path 2007, 88:427-436.
- Qu P, Du H, Wilkes DS, Yan C: Critical roles of lysosomal acid lipase in T cell development and function. Am J Pathol 2009, 174:944-956.
- Bendtsen JD, Nielsen H, von Heijne G, Brunak S: Improved prediction of signal peptides: SignalP 3.0. J Mol Biol 2004, 340:783-795.
- Baral PK, Jajanin-Jozi( N, Deller S, Macheroux P, Abrami( M, Gruber K: The first structure of dipeptidyl-peptidase III provides insight into the catalytic mechanism and mode of substrate binding. J Biol Chem 2008, 283(32):22316-22324.
- Lee CM, Snyder SH: Dipeptidyl-aminopeptidase III of rat brain: selective affinity for enkephalin and angiotensin. J Biol Chem 1982, 257(20):12043-12050.
- Abrami( M, Zubanovi( M, Vitale L: Dipeptidyl peptidase III from human erythrocytes. Biol Chem Hoppe Seyler 1988, 369:29-38.
- Skurk T, Lee YM, Hauner H: Angiotensin II and its metabolites stimulate PAI-I protein release from human adipocytes in primary culture. Hypertension 2001, 37:1336-1340.
- Turner AJ, Isaac RE, Coates D: The neprilysin (NEP) family of zinc metalloendopeptidases: Genomics and function. Bioessays 2001, 23(3):261-269.
- Matsas R, Fulcher IS, Kenny AJ, Turner AJ: Substance P and (Leu)enkephalin are hydrolysed by an enzyme in pig caudate synaptic membranes that is identical with the endopeptidase of kidney microvilli. Proc Natl Acad Sci USA 1983, 80:3111-3115.
- Isaac RE: Neuropeptide-degrading endopeptidase activity of locust (Schistocerca gregaria) synaptic membranes. Biochem J 1988, 255:843-847.
- Fry BG, Scheib H, Weerd L van der, Young B, McNaughtan J, Ramjan SFR, Vidal N, Poelmann RE, Norman JA: Evolution of an arsenal. Mol Cell Prot 2008, 7:215-246.
- Deshimaru M, Ogawa T, Nakashima KI, Nobuhisa I, Chijiwa T, Shimohigashi Y, Fukumaki Y, Niwa M, Yamashina I, Hattori S, Ohno M: Accelerated evolution of crotalinae snake venom gland serine proteases. FEBS Letters 1996, 397:83-88.
   Ogawa T, Chijiwa T, Oda-Ueda N, Ohno M: Molecular diversity
- Ogawa T, Chijiwa T, Oda-Ueda N, Ohno M: Molecular diversity and accelerated evolution of C-type lectin-like proteins from snake venom. *Toxicon* 2005, 45:1-14.
- Nakashima K, Ogawa T, Oda N, Hattori M, Sakaki Y, Kihara H, Ohno M: Accelerated evolution of *Trimeresurus flavovirids* venom gland phosphlipase A<sub>2</sub> isoenzymes. Proc Natl Acad Sci USA 1993, 90:5964-5968.
- 78. Brecher P, Kuan HT: Lipoprotein lipase and acid lipase activity in rabbit brain microvessels. J Lipid Res 1979, 20:464-471.
- 79. Sandbank U, Djaldetti M: Effect of Echis colorata venom inoculation on the nervous system of the dog and guinea pig. Acta Neuropath 1966, 6:61-69.
- 80. The PartiGene EST-software pipeline at the nematode and neglected genomics database [http://www.nematodes.org/bio informatics/PartiGene/index.shtml]
- Parkinson J, Anthony A, Wasmuth J, Schmid R, Hedley A, Blaxter M: PartiGene - constructing partial genomes. *Bioinformatics* 2004, 20:1398-1404.
- 82. Glenner GG, Folk JE: **Glutamyl peptidases in rat and guinea pig** kidney slices. *Nature* 1961, **192:**338-340.
- Marchio S, Lahdenranta J, Schlingemann RO, Valdembri D, Wesseling P, Arap MA, Hajitou A, Ozawa MG, Trepel M, Giordano RJ, Nanus DM, Dijkman HB, Ooserwijk E, Sidman RL, Cooper MD, Bussolino F,

Pasqualini R, Arap W: Aminopeptidase A is a functional target

- in angiogenic blood vessels. *Cancer Cell* 2004, **5**:151-162. Fournie-Zaluski MC, Fassot C, Valentin B, Djordjijevic D, Reaux-Le Goazigo A, Corvol P, Roques BP, Llorens-Cortes C: **Brain renin**-84. angiotensin system blockade by systemically active aminopeptidase A inhibitors: a potential treatment of salt-dependent hypertension. Proc Natl Acad Sci USA 2004, dependent hypertension. 101:7775-7780.
- 85. Fürstenau CR, Trentin DDS, Barreto-Chaves MLM, Sarkis JJF: Ectonucleotide pyrophosphate/phosphodiesterase as part of a multiple system for nucleotide hydrolysis by platelets from rats: Kinetic characterization and biochemical properties. Platelets 2006, 17(2):84-91.
- 86. Taborska E: Intraspecies variability of the venom of Echis carinatus. Physiol Bohemoslov 1971, 20:307-318.
- 87. Sales PBV, Santoro ML: Nucleotide and DNase activities in Brazilian snake venoms. Comp Biochem Physiol C Toxicol Pharmacol 2008, 147(1):85-95.
- 88. Champagne DE: Antihemostatic molecules from saliva of blood-feeding arthropods. Pathophysiol Haemos Thromb 2005, 34:221-227.

