The Association between Malaria and Iron Status or Supplementation in Pregnancy: A Systematic Review and Meta-Analysis

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Abstract

Introduction: Malaria prevention and iron supplementation are associated with improved maternal and infant outcomes. However, evidence from studies in children suggests iron may adversely modify the risk of malaria. We reviewed the evidence in pregnancy of the association between malaria and markers of iron status, iron supplementation or parenteral treatment.

Methods and Findings: We searched MEDLINE, EMBASE, the Cochrane Central Register of Controlled Trials, the Global Health Library, and the Malaria in Pregnancy library to identify studies that investigated the association between iron status, iron treatment or supplementation during pregnancy and malaria. Thirty one studies contributed to the analysis; 3 experimental and 28 observational studies. Iron supplementation was not associated with an increased risk of P. falciparum malaria during pregnancy or delivery in Africa (summary Relative Risk = 0.89, 95% Confidence Interval (CI) 0.66–1.20, I² = 78.8%, 5 studies). One study in Asia reported an increased risk of P. vivax within 30 days of iron supplementation (e.g. adjusted Hazard Ratio = 1.75, 95% CI 1.14–2.70 for 1–15 days), but not after 60 days. Iron deficiency (based on ferritin and C-reactive protein) was associated with lower odds for malaria infection (summary Odds Ratio = 0.35, 0.24–0.51, I² = 59.2%, 5 studies). With the exception of the acute phase protein ferritin, biomarkers of iron deficiency were generally not associated with malaria infection.

Conclusions: Iron supplementation was associated with a temporal increase in P. vivax, but not with an increased risk of P. falciparum; however, data are insufficient to rule out the potential for an increased risk of P. falciparum. Iron deficiency was associated with a decreased malaria risk in pregnancy only when measured with ferritin. Until there is more evidence, it is prudent to provide iron in combination with malaria prevention during pregnancy.

Introduction

Anemia affects the lives of more than 500 million women in developing countries. The consequences of anemia during pregnancy include maternal mortality and stillbirth [1–5]. Iron deficiency is generally regarded as the most common cause of anemia accounting for an estimated 50% of all anemia worldwide. Iron deficiency anemia (IDA) is thought to cause an estimated 600,000 perinatal and 100,000 maternal deaths per year globally [6]. In areas where anemia is highly prevalent, international guidelines recommend universal iron and folic acid supplementation throughout pregnancy [7].

A recently updated Cochrane review showed evidence that pregnant women taking iron supplements reduced their risk on anemia by 70% (risk ratio [RR] 0.3, 95% confidence interval [CI] 0.19–0.46) and iron-deficiency at term by 57% (RR 0.43, 95% CI 0.27–0.66) [8]. In addition, they were less likely to have low birth weight newborns (<2500 grams) compared with controls (RR 0.81, 95% CI 0.68–0.97). The mean birth weight was 30.8 grams greater among infants whose mothers received iron during pregnancy (95% CI 5.9–55.7 grams), whereas the relationship between iron supplementation and premature birth (<37 weeks of gestation; RR 0.88, 95% 0.77–1.01) and neonatal death was not significant (RR 0.90, 95% CI 0.68–1.19) [8]. It is not clear yet if supplementation translates into clinical improvements such as reducing the incidence of puerperal infection or postpartum hemorrhage. Other benefits of maternal iron supplementation include improved newborn iron stores which are determined...
through in utero acquisition of iron [9], and these iron stores remain associated with the infants’ iron status at 9 and 24 months of age [10,11]. This has important implications given that IDA has been associated with impaired cognitive and motor development in children [12,13].

The benefits of maternal iron supplementation in malaria-endemic areas are less clear. An analysis of 101,636 singleton live-born infants conducted using data from Demographic and Health Surveys of 19 malaria-endemic countries in sub-Saharan Africa found that infants whose mothers received any iron/folic acid supplements and intermittent preventive treatment with sulfadoxine-pyrimethamine (IPTp-SP) for malaria during pregnancy were less likely to have a neonatal death compared to women who did not receive either iron/folic acid or malaria prophylaxis (hazard ratio (HR): 0.76; 95% CI 0.58–0.99) [14]. However, this effect was not seen among mothers who received only iron/folic acid supplements or only IPTp-SP. A recent meta-analysis of 32 nationally representative health surveys in sub-Saharan Africa, confirmed the protective association of malaria prevention (IPTp-SP and ITNs) with a protective efficacy of 18% on neonatal mortality (95% CI 4–30%) and of 21% for low birth weight (95% CI 14–27%) [15].

In summary, malaria prevention and iron supplementation are each associated with improved maternal and infant outcomes. However, the benefits of iron supplementation in pregnancy must be carefully weighed against the possibility of adverse consequences caused by this intervention in certain settings. Evidence from several studies among children suggests iron supplementation and iron status may adversely modify the risk of malaria, complicating a universal policy of routine iron supplementation in children in malaria endemic areas [16–19]. However, a Cochrane review of this topic concluded in the presence of regular malaria surveillance and appropriate treatment there is no increase in malaria risk among children [20]. A technical working group on iron and malaria established by the U.S. National Institute of Child Health and Human Development recently reviewed the evidence and concluded, “The balance of evidence indicates that the administration of iron supplements, usually in combination with folic acid, increases the risk of malarial morbidity when given without malarial prophylaxis, and in the absence of universal access to treatment” [21].

Despite the universal recommendation for iron supplementation in many malaria endemic countries, this question has yet to be examined in pregnancy, thereby creating uncertainty among public health programs regarding the use of iron supplementation during pregnancy in malaria endemic areas. To address this gap, we review the evidence in pregnancy of the association between the risk of malaria and iron status, iron treatment or iron supplementation.

Methods

Search Strategy

Studies investigating the association between iron status, iron treatment or iron supplementation during pregnancy and malaria risk were identified by searching MEDLINE, EMBASE, the Cochrane Central Register of Controlled Trials, the Global Health Library from the World Health Organization (WHO) and the Malaria in Pregnancy library from their inception to January 2013 inclusive without language restrictions [22]. The search was limited to human studies and used the following search terms: (pregnant OR pregnancy OR placental OR placenta OR fetus OR fetal OR foetus OR foetal) AND (iron OR ferrous OR ferric) AND (malaria OR parasitemia OR parasitaemia OR plasmodium OR plasmodium OR falciparum or vivax). The references of all identified articles, as well as additional review articles, were examined to locate additional studies not identified during the computerized search. Authors of potentially relevant articles were contacted if their methodology suggested relevant data were available but not presented in the published manuscript. The study adhered to the PRISMA statement [23].

Selection

Using the search criteria defined above, 299 publications were identified. Each manuscript was reviewed by two authors (LS and AVE) and included if it met the following criteria: (1) study population included pregnant women; (2) data were available to assess the association between iron status, iron treatment, or iron supplementation and malaria; (3) for longitudinal studies and trials: an appropriate control group was included that did not receive iron; and (4) iron biomarkers were limited to serum ferritin, serum iron, serum transferrin, transferrin saturation (TS%), soluble transferrin receptor (sTfR), total iron binding capacity (TIBC), and erythrocyte protoporphyrin (EP). Discrepancies between reviewers were resolved through discussions until consensus was reached. For articles where it could be assumed that information was available but not presented in the format needed (e.g. the study reported on malaria and iron deficiency, but no data on iron deficiency by malaria status was presented), the authors were contacted for additional information.

Validity Assessment

Details of the methods used to assess validity are included in the supplemental appendix. In summary, the Cochrane Collaboration’s tool for assessing the risk of bias among randomized trials was used to determine the quality of included trials as ‘low’ (high risk of bias), ‘high’ (low risk of bias), or ‘unclear’ [24]. The assessment of non-randomized study designs were based on source population, participant selection, completeness of exposure and outcome data, appropriate tests, sample size, and measures to control confounding [25,26]. Quality was classified as low-to-moderate or good (Supplement 1, Figure S1.1, Figure S1.2 in File S1).

Data Abstraction

A standardized data abstraction form was used to collect the following data elements: year of publication, geographic location, population, design, number of subjects enrolled, malaria endemicity, duration, frequency and dose of iron, concurrent malaria treatment or prevention, incidence or prevalence of malaria parasitemia or placental malaria infection and biomarkers for iron status.

Classification of Studies

Studies focusing on iron status, iron treatment, and iron supplementation were considered separately. Iron status studies were further stratified into iron deficiency and iron biomarkers studies. Study design was classified as randomized-controlled trial (RCT), prospective cohort study, retrospective cohort study, case-control study, cross-sectional study, or before-after study.

Outcomes

The primary outcomes of interest were peripheral malaria parasitemia during pregnancy or peripheral or placental malaria infection at the time of delivery by blood smear. Information from additional malaria tests were included where available (e.g. placental histology, polymerase chain reaction).
Meta-analysis

Data from studies of iron deficiency status and individual iron biomarkers were summarized using forest plots and meta-analysis was conducted where possible. If malaria was assessed by more than one method, the most common test was utilized for the pooled analysis to decrease heterogeneity. Iron supplementation data were transformed into summary risk ratios (RR) and 95% confidence intervals (CI) estimated as the ratio of the proportion of women with malaria infection among those receiving and not receiving iron supplementation during pregnancy. Available information in included studies allowed three subgroup analyses: by HIV status, by duration of iron supplementation, and by sickle cell genotype. Causes of anemia among HIV-infected pregnant women, iron status and effect of iron supplementation differ from HIV-negative pregnant women [27,28].

Iron deficiency data was described as ratio of the odds of malaria among iron deficient vs. non-deficient pregnant women in cross-sectional studies and case-control studies. Pooled analyses of iron deficiency and malaria were stratified by timing of malaria test (during pregnancy or at delivery) and the definition of iron deficiency. We used three groups: 1) an iron-deficiency definition based on ferritin and C-reactive protein (CRP, e.g. ferritin < 30 ng/mL with CRP < 8.2 ng/mL or ferritin < 70 ng/mL with CRP > 2.2 ng/mL, the most common definition), 2) a definition based on ferritin alone or in another combination (e.g. sTfR/log ferritin ratio), and 3) all other definitions that did not use ferritin (e.g. erythrocyte protoporphyrin). The summary odds ratio (OR) and 95% CI were computed as the pooled ratio of the odds of malaria among iron deficient and iron replete women; where available, adjusted odds ratios were used.

Mean differences (MD) were computed to compare the overall effect for all individual biomarkers by transforming biomarkers to the same unit (serum iron, TIBC, sTfR, and TS). Because of skewed distributions for most of the ferritin data, we used geometric means for ferritin; studies which did not present geometric means were transformed and geometric mean difference and 95% confidence interval calculated using the Taylor series approximation [29,30]. In addition, the ratios of geometric means were calculated. Individual biomarkers were not stratified by type of malaria test due to the similarity in results between peripheral and placental malaria. All meta-analyses were done using a DerSimonian and Laird random effects model [31]. The Cochrane’s chi-squared test for heterogeneity set at a significance of \( p < 0.10 \) was evaluated. The extent of heterogeneity was measured using I², a measure of the proportion of total variability explained by heterogeneity; this is expressed as a percentage, with 0–25% indicating no or little heterogeneity [32]. Data were analyzed using Stata 12.1 (Stata Corporation, College Station, Texas, USA).

Figure 1. Flow diagram of study selection. Footnote: *1 study (Kapito-Tembo 2010) appears in categories of iron supplementation and iron status. **Iron status is further stratified into iron deficiency vs. malaria risk and iron biomarkers vs. malaria risk. doi:10.1371/journal.pone.0087743.g001
<table>
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<th>Author, Year</th>
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<th>Malaria endemicity (Study prevalence)</th>
<th>Iron Dose (and folic acid if available)</th>
<th>Concurrent malaria control or treatment</th>
<th>Iron supplementation</th>
<th>Comparison group (no iron supplementation)</th>
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<td>Kapito-Tembo 2010 [34]</td>
<td>Malawi (Dec 2005–July 2009)</td>
<td>Cross-sectional</td>
<td>HIV (+) pregnant women with ≥34 gestational wks attending routine ANC. Women &lt;15 yrs and with immediate life-threatening medical and obstetric conditions were excluded</td>
<td>Endemicity not stated (10%: PCR; 5.5%: Microscopy)</td>
<td>Formulation not stated</td>
<td>49.7% IPTp-SP; 20.4% CTX; 15.4% IPTp+CTX; 59.6% bed net</td>
<td>1) Any iron use 2) duration iron use &gt;30 days</td>
<td>1) No iron use 2) duration iron use ≤30 days</td>
<td>Peripheral parasitemia at enrollment</td>
</tr>
<tr>
<td>Menendez 1994 [37]</td>
<td>The Gambia (1980s)</td>
<td>RCT</td>
<td>Multigravida pregnant women excluding those with a packed cell volume &lt;35% at either of the first 2 visits</td>
<td>Seasonal with high transmission (36 wks = 27% Placental malaria = 57%)</td>
<td>200 mg ferrous sulphate daily (=60 mg elemental iron) daily distributed by TBAs to study participants on a weekly basis and 5 mg Folic acid weekly</td>
<td>Women with parasitemia were treated with CQ 25 mg/base/kg for 3 days</td>
<td>Iron Placebo</td>
<td>1) Peripheral parasitemia; 2) Placental malaria</td>
<td></td>
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<tr>
<td>Menendez 1995 [33]</td>
<td>The Gambia (1980s)</td>
<td>Subgroup analysis from RCT (Menendez, 1994)</td>
<td>Multigravida pregnant women excluding those with a packed cell volume &lt;25% at either of the first 2 visits</td>
<td>Seasonal with high transmission (36 wks = 27% Placental malaria = 57%)</td>
<td>200 mg ferrous sulphate daily (=60 mg elemental iron) daily distributed by TBAs to study participants on a weekly basis and 5 mg Folic acid weekly</td>
<td>Women with parasitemia were treated with CQ 25 mg/base/kg for 3 days</td>
<td>Iron Placebo</td>
<td>1) Peripheral parasitemia at 36 wks; 2) Postnatal parasitemia; 3) Placental malaria</td>
<td></td>
</tr>
<tr>
<td>Mwapasa 2004 [35]</td>
<td>Malawi, Dec 2000, June 2002</td>
<td>Cross-sectional</td>
<td>Pregnant women attending the labor ward and were excluded if they were less than 15 years of age or had hypertension, multiple gestations or altered consciousness.</td>
<td>Perennial with peaks December to April (10–46)</td>
<td>400mg iron +5 mg Folic acid daily</td>
<td>95% of women received 1+ dose of IPTp-SP, 23.2% used bed net</td>
<td>1) Iron+FA use 2) duration of iron use &gt;30 days</td>
<td>1) No Iron+FA use 2) duration of iron use ≤30 days</td>
<td>Peripheral parasitemia at delivery</td>
</tr>
<tr>
<td>Nacher 2003 [38]</td>
<td>Thailand (1993–1997)</td>
<td>Prospective Cohort</td>
<td>Pregnant women in camps for displaced persons on the Thai-Burmese border excluding those with malaria during the index pregnancy before enrollment and women with malaria at enrollment or within 8 weeks of follow-up</td>
<td>Seasonal with EIR&lt;1 (PV = 15%; PF = 5.7%)</td>
<td>If HCT&lt;30%: 5 mg FA+600 mg Ferrous Sulfate daily until delivery</td>
<td>PV-CQ; PF-Quinine, MQ or Artesunate</td>
<td>Duration of FA+Iron: 1–15d 16–30d 31–60d &gt;60d</td>
<td>No FA+ Iron use</td>
<td>First malaria episode during follow-up</td>
</tr>
<tr>
<td>Nyomugeny 2000 [36]</td>
<td>Uganda (Feb 1996–Feb 1998)</td>
<td>RCT</td>
<td>Primigravida in their first or second trimester attending ANC for the first time without severe anemia</td>
<td>Hyper-endemic (38.2% at enrollment; 39% at delivery)</td>
<td>120 mg elemental iron daily +5 mg folic acid weekly</td>
<td>Active case management (group A also received 300 mg CQ weekly)</td>
<td>Active case management+Iron/FA</td>
<td>Active case management+Placebo</td>
<td>1) Peripheral parasitemia; 2) Placental malaria</td>
</tr>
</tbody>
</table>
Table 1. Cont.

<table>
<thead>
<tr>
<th>Author, Year</th>
<th>Country</th>
<th>Iron Dose (and folate if available)</th>
<th>Concurrent malaria endemicity</th>
<th>Study Design</th>
<th>Population</th>
<th>Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>van Eijk</td>
<td>Kenya</td>
<td>200 mg ferrous sulphate 3 times per day</td>
<td>16.7% Placental parasitemia in 3rd trimester</td>
<td>Before-After study</td>
<td>Pregnant women with uncomplicated singleton pregnancies &gt; 32 weeks gestation excluding those with underlying chronic illness</td>
<td>Peripheral parasitemia in 3rd trimester</td>
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Results

Of 299 published studies identified, 31 met the inclusion criteria (figure 1). Included studies were from a broad range of geographic locations and published between 1970 and 2012. Studies were categorized as those assessing the association between oral iron supplementation and malaria risk in pregnancy [33–39], iron deficiency and malaria risk in pregnancy [34,40–50], iron biomarkers and malaria risk in pregnancy [40–43,46,47,50–61], or iron treatment and malaria risk in pregnancy [62,63]. These included two randomized controlled trials [36,37] and one sub-group analysis [33] from the main trial [37], two prospective cohort studies [38,62], one before-after study [39], one retrospective cohort study [63], six case-control studies [40,47,49,52–54], and 18 cross-sectional studies (table 1–4) [34,35,41–46,48,50,51,55–61]. One cross-sectional study was described in two articles, the original analyzed serum ferritin relative to malaria infection [57] and the supplemental article considered iron deficiency status controlling for inflammation relative to malaria infection [45].

Quality of the Included Studies

The two trials, both published before the more widespread adoption of the CONSORT guidelines, were assessed as low quality, mainly because allocation concealment was not described. Overall, there were ten observational studies among the 25 evaluated which we marked as good quality. None of the observational studies reported a justification for the sample size of the study. For 16 studies, the association between iron and malaria was reported as one of the primary purposes of the study. For 16 studies, the association between iron and malaria was reported as one of the primary purposes of the study. For 16 studies, the association between iron and malaria was reported as one of the primary purposes of the study. For 16 studies, the association between iron and malaria was reported as one of the primary purposes of the study.

Oral Iron Supplementation and Malaria Risk

P. falciparum malaria. The effect of oral iron supplementation on malaria risk in pregnancy was assessed in seven studies which included the two trials [36,37], the one randomized trial sub-group analysis [33], one prospective cohort study [38], one before-after study [39], and two cross-sectional studies [34,35]. The dose and duration of iron supplementation varied in the studies, as did the level of malaria endemicity and use of malaria prevention interventions (table 1).

A meta-analysis was performed among the iron supplementation studies stratified by timing of malaria test. This analysis excluded the sub-group analysis from the RCT because these women were represented in the parent study [33], and the study using hazard ratios because these data were only presented by duration of iron supplementation and therefore not comparable to the other studies [38]. The overall pooled result of iron supplementation during pregnancy or delivery was not associated with an increased risk of malaria (RR pooled 0.89, 95% CI 0.66–1.12; I² 38.4%, figure 2). Stratum specific results also showed no increase in risk of malaria associated with iron supplementation during pregnancy (RR pooled 0.58, 95% CI 0.20–1.73; I² 89.9%) or at the time of delivery (RR pooled 1.02, 95% CI 0.75–1.39; I² 73.0%, figure 2). Baseline iron status was not assessed in any of the studies, so a subgroup analysis among iron-replete and iron deficient women was not possible.

The results of further sub-group analysis can be seen in the supplement (File S1). No significant difference was seen by HIV TX, and Comprehensive Meta analysis (version 2.2.055) was used for subgroup analysis.
Table 2. Summary of included studies for iron deficiency and malaria risk in pregnancy.

<table>
<thead>
<tr>
<th>Author, Year</th>
<th>Country (Time period)</th>
<th>Study Design</th>
<th>Population</th>
<th>Malaria endemcity (Study prevalence)</th>
<th>Iron Dose (and folic acid if available)</th>
<th>Concurrent malaria control or treatment</th>
<th>Exposure: Malaria</th>
<th>Comparison: no malaria</th>
<th>Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>Abrams 2005 [40] a</td>
<td>Malawi (Feb-Oct 2002)</td>
<td>Case-control</td>
<td>Pregnant women attending the labor ward who did not have HIV, preeclampsia or multiple gestations</td>
<td>Perennial with peaks December to April (12.7%)</td>
<td>Not stated</td>
<td>91% took antimalarial tablets; 26% slept under mosquito nets</td>
<td>Peripheral parasitemia at delivery</td>
<td>No peripheral parasitemia at delivery</td>
<td>Iron deficiency b at delivery</td>
</tr>
<tr>
<td>Engmann 2008 [41] d</td>
<td>Ghana (May-Aug, 2003)</td>
<td>Cross-sectional</td>
<td>Pregnant women 18–40 yrs with singleton pregnancies receiving ANC. Women with sickle cell, major congenital or current illnesses were excluded.</td>
<td>Endemicity not stated (7%)</td>
<td>All patients attending ANC receive free iron supplementation (formulation not stated)</td>
<td>All patients attending ANC receive IPTp</td>
<td>Peripheral parasitemia at enrollment</td>
<td>No peripheral parasitemia at enrollment</td>
<td>Iron deficiency e at enrollment</td>
</tr>
<tr>
<td>Hindaraker 2002 [47] a</td>
<td>Tanzania (Feb 1995–Mar 1996)</td>
<td>Case-Control f</td>
<td>Pregnant women at their first ANC visit</td>
<td>Endemicity not stated (18.1%)</td>
<td>NA</td>
<td>NA</td>
<td>Peripheral parasitemia at enrollment</td>
<td>No peripheral parasitemia at enrollment</td>
<td>Iron deficiency b at enrollment</td>
</tr>
<tr>
<td>Matteelli 1994 [42]</td>
<td>Zanzibar (Dec 1989–Apr 1990)</td>
<td>Cross-sectional</td>
<td>Pregnant women admitted for uncomplicated delivery</td>
<td>Highly endemic (21.5%)</td>
<td>Not stated</td>
<td>Not stated</td>
<td>Peripheral parasitemia at delivery</td>
<td>No peripheral parasitemia at enrollment</td>
<td>Iron deficiency d at enrollment</td>
</tr>
<tr>
<td>Ouedraogo 2012 [43] a</td>
<td>Benin (Jan 2010–May 2011)</td>
<td>Cross-sectional</td>
<td>HIV (-) pregnant women with &lt;28 gestational wks attending routine ANC who had not yet taken IPTp, iron, folic acid, vitamin B12 or anti-helminthics</td>
<td>Perennial with two high seasonal peaks (15.1%)</td>
<td>NA</td>
<td>NA</td>
<td>Peripheral parasitemia at enrollment</td>
<td>No peripheral parasitemia at enrollment</td>
<td>Iron deficiency e at enrollment</td>
</tr>
<tr>
<td>Senga 2011 [49] a</td>
<td>Malawi (2004–2005)</td>
<td>Case-Control</td>
<td>All pregnant women who attended the hospital for delivery excluding those with emergency obstetric conditions</td>
<td>Highly endemic with year-round transmission and seasonal peaks</td>
<td>60 mg iron i.v. suppositories at delivery</td>
<td>95% of women received 1+ doses of IPT-SP</td>
<td>Placental malaria</td>
<td>No placental malaria</td>
<td>Iron deficiency b at delivery</td>
</tr>
<tr>
<td>Van Santen 2011 [50] a</td>
<td>Gabon (2000–2004)</td>
<td>Cross-sectional</td>
<td>Primigravida without peripheral parasitemia at enrollment with singleton pregnancy with no indication of systemic infection</td>
<td>Stable meso-hyperendemic</td>
<td>60 mg iron daily provided through ANC</td>
<td>Not stated</td>
<td>Placental Malaria</td>
<td>No Placental Malaria</td>
<td>Iron deficiency b at delivery</td>
</tr>
<tr>
<td>Kabuye-Mena 2008 [48]</td>
<td>Tanzania (2002–2005)</td>
<td>Cross-sectional</td>
<td>Participants of the Mother-Offspring Malaria Studies Project in Tanzania excluding those with evidence of chronic or debilitating illnesses</td>
<td>Intense malaria transmission BIR 7400 (12.4%)</td>
<td>Not stated</td>
<td>56-8% of women used IPT; ITNs were used by 14.7% of women</td>
<td>Iron deficient at delivery d</td>
<td>Iron eplate at delivery</td>
<td>Placental malaria</td>
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### Table 2. Cont.

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<td>Endemicity not stated (10%: PCR; 5.9%: Microscopy)</td>
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<td>Senga 2012 [44]</td>
<td>Malawi (1992–1995)</td>
<td>Cross-sectional</td>
<td>Pregnant women at first ANC visit and at delivery participating in a cohort study to assess effect MiP on fetal hemoglobin</td>
<td>Highly endemic with year-round transmission and seasonal peaks</td>
<td>60 mg iron and 250 µg folic acid daily through ANC</td>
<td>69.1% at delivery at least one dose of SP</td>
<td>Iron deficiency</td>
<td>No iron deficiency</td>
<td>Peripheral parasitemia at enrollment Peripheral and placental malaria at delivery</td>
</tr>
</tbody>
</table>


1. Author contacted and additional information was obtained.
2. Ferritin < 30 ng/mL with CRP ≤ 8.2 ng/mL or ferritin < 70 ng/mL with CRP > 8.2 ng/mL.
3. Serum ferritin < 10 (µg/L) or Erthyrocyte protoporphyrin > 70 (µmol/mol heme).
4. Author contacted and author responded, but no additional information was available.
5. Serum ferritin = 16 ng/mL.
6. Serum ferritin ≤ 15 ng/mL.
7. Selected based on hemoglobin status (<70 g/L, 70–90 g/L, 90–110 g/L, 110–150, and >150).
8. Serum ferritin ≤ 15 ng/mL.
9. sTfR/log ferritin ratio > 1.6.
10. Red cell zinc protoporphyrin/heme > 2.7 µg/g hemoglobin.

DOI: 10.1371/journal.pone.0087743.t002
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<td>Case-control</td>
<td>Pregnant women attending the labor ward who did not have HIV, preeclampsia or multiple gestations</td>
<td>Perennial with peaks December to April (12.7%)</td>
<td>Not stated</td>
<td>91% took antimalarial tablets; 26% slept under mosquito nets</td>
<td>Peripheral parasitemia at delivery</td>
<td>No peripheral parasitemia</td>
<td>Iron biomarkers* at delivery</td>
</tr>
<tr>
<td>Asaolu 2009 [53]</td>
<td>Nigeria (Not stated)</td>
<td>Case-control</td>
<td>Pregnant women attending ANC</td>
<td>Endemicity not stated (Unclear if selected on malaria status)</td>
<td>Not stated</td>
<td>Not stated</td>
<td>Peripheral parasitemia at enrolment</td>
<td>No peripheral parasitemia at enrolment</td>
<td>Serum iron (μmol/L)</td>
</tr>
<tr>
<td>Ayoya 2006 [51]</td>
<td>Mali (June-Aug 2002)</td>
<td>Cross-sectional</td>
<td>Pregnant women (18-45 yrs) attending community health clinic during study period excluding those who used oral iron or anthelminthics since the start of pregnancy or those with a blood transfusion in the 3 months before study entry</td>
<td>Highly endemic (11%)</td>
<td>NA</td>
<td>NA</td>
<td>Peripheral PF parasitemia at enrolment</td>
<td>No peripheral PF parasitemia at enrolment</td>
<td>Iron biomarkers* at enrollment</td>
</tr>
<tr>
<td>Engmann 2008 [41] d</td>
<td>Ghana (May-Aug, 2003)</td>
<td>Cross-sectional</td>
<td>Pregnant women 18-40 yrs with singleton pregnancies receiving ANC. Women with sickle cell, major congenital or current illnesses were excluded.</td>
<td>Endemicity not stated (7%)</td>
<td>All patients attending ANC receive free iron supplementation (formulation not stated)</td>
<td>All patients attending ANC receive IPTp</td>
<td>Peripheral parasitemia at enrollment</td>
<td>No peripheral parasitemia at enrollment</td>
<td>Serum ferritin (μg/L) at enrollment</td>
</tr>
<tr>
<td>Eteng 2010 [54]</td>
<td>Nigeria (before 2010)</td>
<td>Case-control</td>
<td>Pregnant women attending ANC with symptomatic malaria and healthy controls. Women were included if they were not on iron therapy or hematinic drugs</td>
<td>Malaria endemic (NA-selected on malaria status)</td>
<td>None used</td>
<td>Not stated</td>
<td>Peripheral parasitemia at enrollment</td>
<td>No peripheral parasitemia at enrollment</td>
<td>Iron biomarkers* at enrollment</td>
</tr>
<tr>
<td>Hinderaker 2002 [47] a</td>
<td>Tanzania (Feb 1995-Mar 1996)</td>
<td>Case-Control</td>
<td>Pregnant women at their first ANC visit</td>
<td>Endemicity not stated (22.8%)</td>
<td>NA</td>
<td>NA</td>
<td>Peripheral parasitemia at enrolment</td>
<td>No peripheral parasitemia at enrolment</td>
<td>Iron biomarkers* at enrollment</td>
</tr>
<tr>
<td>Huddle 1999 [55]</td>
<td>Malawi (Nov 1993-Feb 1994)</td>
<td>Cross-sectional</td>
<td>Pregnant women attending ANC aged 14-45 y with no history of c-section and hb&gt;80 g/l</td>
<td>Endemicity not stated (31%)</td>
<td>Not stated</td>
<td>Not stated</td>
<td>Peripheral parasitemia</td>
<td>No peripheral parasitemia</td>
<td>Iron biomarkers*</td>
</tr>
<tr>
<td>Massawe 2002 [56] d</td>
<td>Tanzania (Aug-Sept 1998)</td>
<td>Cross-sectional</td>
<td>Consecutive primigravida women ≥20 yrs attending their first ANC visit</td>
<td>Not stated (43.4%)</td>
<td>NA</td>
<td>NA</td>
<td>Peripheral parasitemia at enrollment</td>
<td>No peripheral parasitemia at enrollment</td>
<td>Iron biomarkers* at enrollment</td>
</tr>
<tr>
<td>Matteelli 1994 [42]</td>
<td>Zanzibar (Dec 1989-Apr 1990)</td>
<td>Cross-sectional</td>
<td>Pregnant women admitted for uncomplicated delivery</td>
<td>Highly endemic (21.5%)</td>
<td>Not stated</td>
<td>Not stated</td>
<td>Peripheral parasitemia at delivery</td>
<td>No peripheral parasitemia at delivery</td>
<td>Serum ferritin (ng/mL)</td>
</tr>
<tr>
<td>Mockenhaupt 2000 [57]</td>
<td>Ghana (Nov-Dec 1998)</td>
<td>Cross-sectional</td>
<td>Pregnant women attend ANC</td>
<td>Endemicity not stated (32%)</td>
<td>Not stated</td>
<td>Not stated</td>
<td>Peripheral parasitemia at enrollment</td>
<td>No peripheral parasitemia at enrollment</td>
<td>Serum ferritin (ng/mL)</td>
</tr>
</tbody>
</table>
### Table 3. Cont.

<table>
<thead>
<tr>
<th>Author, Year</th>
<th>Country (Time period)</th>
<th>Study Design</th>
<th>Population</th>
<th>Malaria endemicity (Study prevalence)</th>
<th>Iron Dose (and folic acid if available)</th>
<th>Concurrent malaria control or treatment</th>
<th>Malaria Infection</th>
<th>Comparisons group: No malaria infection</th>
<th>Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ndyomugenyi 2008 [61]</td>
<td>Uganda (2003–2004)</td>
<td>Cross-sectional</td>
<td>Pregnant women &gt;16 weeks at first ANC</td>
<td>Hyperendemic (35%)</td>
<td>NA</td>
<td>NA</td>
<td>Parasitemia at enrollment</td>
<td>No parasitemia at enrollment</td>
<td>Serum ferritin (μg/L) at enrollment</td>
</tr>
<tr>
<td>Ouédraogo 2012 [43]</td>
<td>Benin (Jan 2010–May 2011)</td>
<td>Cross-sectional</td>
<td>HIV (-) pregnant women with &lt;28 gestational wks attending routine ANC who had not yet taken IPTp, iron, folic acid, vitamin B12 or anti-helminthics</td>
<td>Perennial with two high seasonal peaks (15.1%)</td>
<td>NA</td>
<td>NA</td>
<td>Peripheral parasitemia at enrollment</td>
<td>No peripheral parasitemia at enrollment</td>
<td>Serum ferritin (μg/L) at enrollment</td>
</tr>
<tr>
<td>Reinhardt 1978 [58]</td>
<td>Ivory Coast (Not stated)</td>
<td>Cross-sectional</td>
<td>Women with singleton deliveries</td>
<td>Endemicity not stated (39.4%; Peripheral parasitemia: 32.8%)</td>
<td>Not stated</td>
<td>Not stated</td>
<td>Peripheral parasitemia or placental malaria</td>
<td>Neither peripherallIron biomarkers</td>
<td></td>
</tr>
<tr>
<td>Saad 2012 [52]</td>
<td>Sudan (Aug-Dec 2010)</td>
<td>Case-control</td>
<td>Control group consisted of pregnant women with uncomplicated malaria and healthy pregnant women</td>
<td>Malaria endemic (NA-selected on severe malaria status)</td>
<td>NA</td>
<td>NA</td>
<td>Peripheral parasitemia at enrollment</td>
<td>No peripheral parasitemia at enrollment</td>
<td>Serum ferritin (μg/L) at enrollment</td>
</tr>
<tr>
<td>Shulman 1996 [59]</td>
<td>Kenya (Nov 1993)</td>
<td>Cross-sectional</td>
<td>Pregnant women attending ANC</td>
<td>Perennial transmission EIR 10 (23.6%)</td>
<td>Not stated</td>
<td>Not stated</td>
<td>Peripheral parasitemia at enrollment</td>
<td>No peripheral parasitemia at enrollment</td>
<td>Serum ferritin (ng/mL)</td>
</tr>
<tr>
<td>VanderJagt 2007 [60]</td>
<td>Nigeria (June-Aug 2003)</td>
<td>Cross-sectional</td>
<td>Healthy normotensive pregnant women with no history of hypertension, proteinuria or other complications of pregnancy</td>
<td>Endemicity not stated (9.4%)</td>
<td>Iron and folate supplements are provided to women at ANC whose hematocrit is indicative of anemia</td>
<td>25% of women took malaria prophylaxis</td>
<td>Peripheral parasitemia at enrollment</td>
<td>No peripheral parasitemia at enrollment</td>
<td>Iron biomarkers</td>
</tr>
</tbody>
</table>


*Author contacted and additional information was obtained.

1Serum iron <12 μmol/L; Mean Serum Iron; Mean TIBC.

2Author contacted and author responded, but no additional information was available.

3Selected based on hemoglobin status (<70 g/L, 70-90 g/L, 90-110 g/L, 110-150, and >150).

4Serum iron (μmol/L); Serum ferritin (μg/L); TIBC (μmol/L) Transferrin saturation (%).

5Serum iron (μmol/L); Serum ferritin (μg/L); TIBC (μmol/L) Transferrin saturation (%).

6Serum iron (μmol/L); TS (%); Serum ferritin (μg/L); sTfR (mg/L).

7Serum ferritin (μg/L) sTfR (mg/L).

8Serum iron (μg/100 mL); Transferrin (mg/100 mL).

9Serum iron (μg/dL); Serum ferritin (ng/mL); TIBC (pg/mL).

10Iron (μmol/L); TIBC (μmol/L); TS (%); Ferritin (μg/L); sTfR (mg/L).

doi:10.1371/journal.pone.0087743.t003
### Table 4: Summary of included studies for parenteral iron treatment and malaria risk in pregnancy.

<table>
<thead>
<tr>
<th>Study Design</th>
<th>Population</th>
</tr>
</thead>
<tbody>
<tr>
<td>Intervention</td>
<td>Malaria control or treatment</td>
</tr>
<tr>
<td>Comparison group A</td>
<td></td>
</tr>
<tr>
<td>Byles</td>
<td>Tanzania</td>
</tr>
<tr>
<td>Retrospective</td>
<td>Pregnant women delivering in Malang provincial hospital for delivery, with at least one parasitemia associated with iron supplementation, but only during the first 30 days (aHR 1.75, 95% CI 1.14–2.70, p = 0.009) and 16–30 days (aHR 1.70, 95% CI 1.10–2.60, p = 0.01), after which the effect declined, and reversed for women who had taken iron supplementation for &gt;60 days (aHR 0.67, 95% CI 0.40–1.02, p = 0.06, table 5) [30].</td>
</tr>
<tr>
<td>Byles and Foly</td>
<td>Tanzania</td>
</tr>
<tr>
<td>Cohort</td>
<td>Women used at least one dose of SP, and 23% used nets [35]. By contrast, the cohort study conducted on the Thai-Burmese border suggested no increase in risk of first episode of <em>P. falciparum</em> associated with iron supplementation up to 60 days (e.g. 1–15 days iron supplementation: Adjusted Hazard Ratio (aHR) 1.30, 95% CI 0.67–2.50, p = 0.4, table 5). Longer supplementation was even associated with a reduced risk (aHR iron supplementation &gt;60 days: 0.37, 95% CI 0.17–0.77, p = 0.009, table 5). No antimalarial prophylaxis was used in this cohort and there was no information on net use but women were screened for malaria weekly and treated promptly [30].</td>
</tr>
<tr>
<td>Retrospective</td>
<td>Pregnant women delivering in Madang provincial hospital, with at least one parasitemia associated with iron supplementation, but only during the first 30 days (aHR 1.75, 95% CI 1.14–2.70, p = 0.009) and 16–30 days (aHR 1.70, 95% CI 1.10–2.60, p = 0.01), after which the effect declined, and reversed for women who had taken iron supplementation for &gt;60 days (aHR 0.67, 95% CI 0.40–1.02, p = 0.06, table 5) [30].</td>
</tr>
</tbody>
</table>

**P. falciparum malaria.** In the single study from Asia that assessed the association between iron supplementation and *P. falciparum* malaria, there was an increase in risk of first episode of *P. falciparum* parasitemia associated with iron supplementation, but only during the first 30 days (aHR 1.58, 95% CI 1.19–2.11, p = 0.001) and 16–30 days (aHR 1.65, 95% CI 1.30–2.07, p = 0.001), after which the effect declined, and reversed for women who had taken iron supplementation for >60 days (aHR 0.60, 95% CI 0.40–1.00, p = 0.06, table 5) [30].

### Parenteral Iron Treatment and Malaria Risk

Associations between parenteral iron treatment among severely anemic pregnant women and malaria risk were evaluated in one retrospective and one prospective study (table 6) [62,63]. The retrospective study compared malaria during the post-natal period between severely anemic pregnant women treated during pregnancy with a single intravenous dose of 1750 mg of iron to women without anemia. A marked increase in the odds of postnatal malaria was observed among primiparous women (OR 5.5, 95% CI 2.2–13.5) whereas this association was not seen among multiparous women (OR 1.1, 95% CI 0.73–1.70) [63]. The prospective cohort study assessed the incidence of local and general reactions to total dose infusion of iron dextran among four groups of pregnant women: A: iron only; B: iron+anthistamine; C: iron+chloroquine; D: iron+anthistamine+chloroquine. However, they only assessed the presence of malaria following infusion among those women with general reactions. Among women with general reactions, those not concurrently receiving chloroquine with their iron infusion were more likely to have malaria parasitemia after the infusion compared to women who did receive concurrent chloroquine (28.2% vs. 0%, respectively; p = 0.17) [62]. Indications, timing of iron treatment and malaria assessment between the studies were too dissimilar to perform a pooled analysis.
doi:10.1371/journal.pone.0087743.g002

Table 5. Summary of study outcomes – Iron Supplementation vs. Malaria Risk.

<table>
<thead>
<tr>
<th>Author, Year</th>
<th>Study Design</th>
<th>HIV Status</th>
<th>Type &amp; Timing of malaria test</th>
<th>Exposure</th>
<th>Comparison to exposure</th>
<th>Risk estimate (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Outcome: First malaria episode during follow-up – <em>Plasmodium vivax</em></strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Nacher 2003 [38]</td>
<td>Prospective Cohort</td>
<td>All</td>
<td>Peripheral BS during pregnancy</td>
<td>Iron+FA 1–15 d</td>
<td>No FA+ Iron use</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Iron+FA 16–30d</td>
<td>No FA+ Iron use</td>
<td>aHR(^1) 1.70 (1.10, 2.60)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Iron+FA 31–60d</td>
<td>No FA+ Iron use</td>
<td>aHR(^1) 1.30 (0.94, 1.80)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Iron+FA &gt;60d</td>
<td>No FA+ Iron use</td>
<td>aHR(^1) 0.67 (0.40, 1.02)</td>
</tr>
<tr>
<td><strong>Outcome: First malaria episode during follow-up – <em>Plasmodium falciparum</em></strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Nacher 2003 [38]</td>
<td>Prospective Cohort</td>
<td>All</td>
<td>Peripheral BS during pregnancy</td>
<td>Iron+FA 1–15 d</td>
<td>No FA+ Iron use</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Iron+FA 16–30d</td>
<td>No FA+ Iron use</td>
<td>aHR(^1) 0.90 (0.40, 2.10)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Iron+FA 31–60d</td>
<td>No FA+ Iron use</td>
<td>aHR(^1) 1.40 (0.80, 2.40)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Iron+FA &gt;60d</td>
<td>No FA+ Iron use</td>
<td>aHR(^1) 0.37 (0.17, 0.77)</td>
</tr>
</tbody>
</table>

aHR: adjusted Hazard Ratio. BS: Blood slide. FA: Folic Acid.
\(^1\)Adjusted for *Plasmodium falciparum* or *Plasmodium vivax*, gravidity, age, estimated gestational age, past mean hematocrit.
doi:10.1371/journal.pone.0087743.g002
Table 6. Summary of study outcomes – Iron Treatment vs. Malaria Risk.

<table>
<thead>
<tr>
<th>Author, Year</th>
<th>Study Design</th>
<th>Outcome</th>
<th>Exposure</th>
<th>Comparison</th>
<th>Risk estimate</th>
</tr>
</thead>
<tbody>
<tr>
<td>Oppenheimer, 1986 [63]</td>
<td>Retrospective Cohort</td>
<td>Postnatal malaria- primigravida</td>
<td>Total Dose Iron Infusion 20-4% (11/54)</td>
<td>No Total Dose Iron Infusion 4-5% (3/67)</td>
<td>RR 4.55 (1.34–15.49) OR 3.46 (1.44–20.7)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Postnatal malarial- multigravida</td>
<td>Total Dose Iron Infusion 8-4% (8/95)</td>
<td>No Total Dose Iron Infusion 7.6% (14/184)</td>
</tr>
<tr>
<td>Byles, 1970 [62]</td>
<td>Prospective Cohort</td>
<td>Generalized reactions after total dose iron infusion</td>
<td>Total dose iron+ Concurrent CQ 1.5% (7/462)</td>
<td>Total dose iron+ No concurrent CQ 8.6% (39/455)</td>
<td>OR 0.16 (0.07–0.37) RR 0.18 (0.08–0.39)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>PF parasitemia present among participants with generalized reactions after total dose iron infusion</td>
<td>Total dose iron+ Concurrent CQ 0.0% (0/7)</td>
<td>Total dose iron+ No concurrent CQ 28.2% (11/39)</td>
</tr>
</tbody>
</table>

doi:10.1371/journal.pone.0087743.t006

Figure 3. Malaria parasitemia by blood smear among iron deficient and non-deficient participants, by definition of iron deficiency during pregnancy. Footnote: CI: confidence interval. CRP: C-Reactive Protein. EP: Erythrocyte protoporphyrin. Hb: hemoglobin. *Use of adjusted odds ratios: Kapito-Tembo 2010: odds ratio adjusted for CD4 count, gravidity, and intestinal infections, Dreyfuss: odds ratio adjusted for hookworm infection, serum retinol and trimester of pregnancy. **Iron deficiency definition: Ferritin <30 ng/mL & CRP < = 8.2 ng/mL or ferritin <70 ng/mL & CRP > 8.2 ng/mL. The weight for each study is indicated as a grey block around the risk estimate. For Dreyfuss 2000, malaria parasitemia was limited to P. vivax in Asia. All other studies were conducted in Africa where P. falciparum is the predominant species.
doi:10.1371/journal.pone.0087743.g003
Iron Deficiency and Malaria Risk

The effect of iron deficiency on malaria risk in pregnancy was assessed in 12 studies including three case-control studies [40,47,49], and nine cross-sectional studies (table 2) [34,41–46,48,50]. Among these studies, five considered evidence of malaria at the time of delivery [40,44,48–50], six considered malaria at the time of enrollment [34,41–43,45,47], and one did not specify when iron deficiency was assessed [46]. Seven of these 12 studies used the same definitions of iron deficiency (serum ferritin <30 ng/mL with CRP < 8.2 ng/mL or serum ferritin < 70 ng/mL with CRP > 8.2 ng/mL), and pooled analysis showed an overall decreased odds for peripheral parasitemia in pregnancy (ORpooled 0.33, 95% CI 0.24–0.51; I^2 59.2%, five studies, figure 3) associated with iron deficiency, but this was not significant for placental malaria (ORpooled 0.34, 95% CI 0.11–1.10; I^2 75.8%, two studies, figure 4). Other definitions using ferritin tended to show similar results with decreased odds of malaria among iron-deficient women (figure 3 and 4); however, where definitions were used which did not include ferritin, no overall effect was seen (ORpooled for malaria in pregnancy: 1.65, 95% CI 0.88–3.09, I^2 59.6%, three studies, figure 3). Additional iron deficiency definitions and malaria outcomes which could not be included because of overlap of studies are listed in the supplement File S1. A subgroup analysis was conducted by gravidity which indicated that the association between malaria parasitaemia and markers of iron status were similar among the different gravidae groups (Figure S3.2 in File S1).

Iron Biomarkers and Malaria Risk

Iron biomarkers evaluated included serum ferritin, serum iron, total iron binding capacity, soluble transferrin receptor, transferrin saturation (as %) and serum transferrin (figure 4). Studies which could not be included in forest plots because of insufficient information are presented in Table 7. The pooled analysis of serum ferritin was conducted among ten studies for which geometric mean values were provided or could be calculated and showed that the concentration of serum ferritin was higher among malaria infected pregnant women compared to those who were uninfected (geometric mean differencepooled 0.53, 95% CI 0.44–0.62, I^2 90.8%, figure 5), which corresponds to ferritin levels.

<table>
<thead>
<tr>
<th>Study</th>
<th>Test</th>
<th>Definition Iron Deficiency</th>
<th>Malaria, Odds Ratio (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1) Definition using ferritin &amp; CRP</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Kabyemela 2008*† Placental BS</td>
<td>Ferritin 30/CRP 8.2**</td>
<td>J. J.</td>
<td>0.20 (0.11, 0.36)</td>
</tr>
<tr>
<td>van Santen 2011 † Placental BS</td>
<td>Ferritin 30/CRP 8.2**</td>
<td>18/45 12/24</td>
<td>0.67 (0.25, 1.81)</td>
</tr>
<tr>
<td>Subtotal (I^2-squared = 75.8%, p = 0.042)</td>
<td></td>
<td></td>
<td>0.34 (0.11, 1.10)</td>
</tr>
<tr>
<td>2) Other definitions using ferritin</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Senga 2011* Placental histology sTfR/log ferritin ratio &gt;1.6</td>
<td>J.</td>
<td>0.36 (0.17, 0.76)</td>
<td></td>
</tr>
<tr>
<td>Matteelli 1994 Peripheral BS</td>
<td>Ferritin &lt;16 ng/mL</td>
<td>12/27 94/177</td>
<td>0.71 (0.31, 1.60)</td>
</tr>
<tr>
<td>Subtotal (I^2-squared = 29.9%, p = 0.232)</td>
<td></td>
<td></td>
<td>0.49 (0.26, 0.96)</td>
</tr>
<tr>
<td>3) Other definitions not using ferritin</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Senga 2012 Placental BS</td>
<td>EP &gt;2.7 μg/g hb</td>
<td>178/935 61/390</td>
<td>1.27 (0.92, 1.74)</td>
</tr>
<tr>
<td>Subtotal (I^2-squared = .%, p = .)</td>
<td></td>
<td></td>
<td>1.27 (0.92, 1.74)</td>
</tr>
<tr>
<td>Overall (I^2-squared = 88.0%, p = 0.000)</td>
<td></td>
<td></td>
<td>0.54 (0.24, 1.21)</td>
</tr>
</tbody>
</table>

Figure 4. Malaria parasitemia among iron deficient and non-deficient participants, by definition of iron deficiency at the time of delivery. Footnote: BS: bloodsmear. CI: confidence interval. CRP: C-reactive protein. EP: Erthyrocyte protoporphyrin. Hb: hemoglobin. sTfR: Soluble transferrin receptor. *Adjusted odds ratios used: Kabyemela 2008: odds ratio adjusted for gravidity. Senga 2011: odds ratio adjusted for gravidity, age, and blood group. Placental histology in Senga 2011: active infection defined by acute or chronic infection (parasites alone, or parasites in the presence of haemozoin), No placental infection defined by the absence of parasites and haemozoin. ** Ferritin 30/CRP 8.3: Ferritin <30 ng/mL & CRP < 8.2 ng/mL or ferritin <70 ng/mL & CRP >8.2 ng/mL. The weight for each study is indicated as a grey block around the risk estimate.
Table 7. Summary of study outcomes – Iron biomarkers vs. malaria risk, and outcomes not presented in forest plot.

<table>
<thead>
<tr>
<th>Author, Year</th>
<th>Study Design</th>
<th>Biomarker (Outcome)</th>
<th>Exposure % (n or IQR)</th>
<th>Comparison to exposure % Outcome (n or IQR)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Engmann 2008 [41]</td>
<td>Cross-sectional</td>
<td>Serum ferritin (µg/L)</td>
<td>Peripheral parasitemia at enrollment Median 91 (IQR: 33, 157)</td>
<td>No peripheral parasitemia at enrollment Median 33 (IQR: 20, 50)</td>
<td>Not provided</td>
</tr>
<tr>
<td>Matteelli 1994 [42]</td>
<td>Cross-sectional</td>
<td>Serum ferritin (ng/mL)</td>
<td>Peripheral parasitemia at delivery Mean 65.7 (n = 106)</td>
<td>No peripheral parasitemia at delivery Mean 36.5 (n = 98)</td>
<td>p = 0.002</td>
</tr>
<tr>
<td>Ndyomugyenyi 2008 [61]</td>
<td>Cross-sectional</td>
<td>Serum ferritin (µg/L)</td>
<td>Peripheral parasitemia at enrollment (Means not available)</td>
<td>No peripheral parasitemia at enrollment (means not available)</td>
<td>p = 0.007</td>
</tr>
<tr>
<td>Saad 2012 [52]</td>
<td>Case-control</td>
<td>Serum ferritin (µg/L)</td>
<td>Peripheral parasitemia at enrollment (Uncomplicated malaria) Median 63.3 (IQR: 30.5, 113.2)</td>
<td>No peripheral parasitemia at enrollment Median 34.4 (IQR: 7.9, 60.3)</td>
<td>p = 0.041</td>
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<td></td>
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<td>Serum ferritin (µg/L)</td>
<td>Peripheral parasitemia at enrollment (Severe malaria) Median 78.6 (IQR: 44.1, 148.9)</td>
<td>No peripheral parasitemia at enrollment Median 34.4 (IQR: 7.9, 60.3)</td>
<td>p = 0.002</td>
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<tr>
<td>Dreyfuss 2000 [46]</td>
<td>Cross-sectional</td>
<td>Erythrocyte Protoporphyrin (µmol/mol heme)</td>
<td>Peripheral parasitemia at enrollment Geometric mean 90 (CI 54, 150) (n = 57)</td>
<td>No peripheral parasitemia at enrollment Geometric mean 84 (CI 52, 134) (n = 231)</td>
<td>P = 0.3</td>
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</table>

CI: confidence interval. IQR: Interquartile range.
doi:10.1371/journal.pone.0087743.t007

Figure 5. Geometric mean difference in ferritin among pregnant women infected with malaria compared to pregnant women without malaria. Footnote: CI: confidence interval. SD: standard deviation. *Van Santen 2011 and Massawe 2002: study population primigravidae. Abrams 2005 and Ouedraogo 2012: study population HIV(-) women. The weight for each study is indicated as a grey block around the risk estimate. For Dreyfuss 2000, the malaria parasitemia was limited to *P. Vivax* in Asia. All other studies were conducted in Africa where *P. falciparum* is the predominant species.
doi:10.1371/journal.pone.0087743.g005
being 70% (95% CI 55–86%) higher in women with malaria compared to those without.

The same pattern was seen in four studies which could not be included in the meta-analysis (table 7). No significant pooled differences were detected for serum iron (eight studies), total iron binding capacity (five studies), soluble transferrin receptor (four studies), serum transferrin (three studies), and transferrin saturation (four studies), whereby it should be noted that the heterogeneity was 0% for soluble transferrin receptor and transferrin saturation (figure 6 and 7).

**Discussion**

To our knowledge, this systematic review represents the first effort to assess the association of iron status and use on the risk of malaria in pregnancy. Pooled analyses of iron deficiency status, using a definition based on ferritin and adjusting for inflammation with CRP, indicated pregnant women with iron deficiency had a reduced risk of malaria infection during pregnancy. However, pooled analyses from the remaining biomarkers of iron deficiency were not associated with malaria. Data from randomized and observational studies did not show an increased risk of *P. falciparum* malaria among the participants who received oral iron supplementation, whereas one cohort study in Asia showed an increased risk of *P. vivax* in the first 30 days of oral iron (and folate) supplementation. None of the studies however, assessed the individual’s baseline iron status which may modify the effect of iron supplementation on malaria risk, complicating the interpretation of these data. Except for the studies using parenteral iron treatment, all included studies used oral iron supplementation for the prevention of iron deficiency anemia during pregnancy; oral iron was not given for the treatment of established anemia and/or iron-deficiency in these studies. This difference can be important because the treatment of anemia in pregnancy recommends using temporarily higher doses of iron, specific policies for iron dose and duration vary by country [7].

Only two studies evaluated parenteral treatment with iron dextran and the study qualities were insufficient to make conclusions. In addition, new parenteral iron treatment options using complex iron-polysaccharides are available which may not have a similar risk profile with regards to infections such as malaria [64].

Several hypotheses have been provided as to why malaria infection may be less prevalent among people with iron deficiency.

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**Figure 6. Mean difference in soluble transferring receptor and serum transferrin among pregnant women infected with malaria compared to pregnant women without malaria.** Footnote: CI: confidence interval. SD: standard deviation. *Van Santen 2011 and Massawe 2002: study population primigravidae. Abrams 2005: study population HIV(-) women. The weight for each study is indicated as a grey block around the risk estimate. Removal of the study of Eteng 2010, an outlier for Serum Transferrin, gives the following result: Pooled mean difference Serum Transferrin (g/L): −0.11, 95% CI −0.30 to 0.09, I² 14.8%.

doi:10.1371/journal.pone.0087743.g006
Iron is vital for the survival of the malaria parasite and the parasite is unable to thrive in iron-deficient environments [65]. Specifically, iron deficiency suppresses erythropoiesis, the production of red blood cells, thereby reducing the opportunity for the parasite to infect the host [66]. Additionally, some aspects of the host immunity against malaria may be boosted with iron deficiency. For example, the host macrophage defense utilizes nitric oxide in the control of P. falciparum and because iron can down-regulate the formation of nitric oxide within macrophages [67], iron deficiency may therefore enable a stronger host-macrophage response.

Evidence from iron chelation studies using desferrioxamine (DFO) provides support that iron-deficient environments are unfavorable to foster malaria infections. In one randomized double-blind placebo cross-over study, DFO among asymptomatic parasitemic adults was associated with enhanced parasite clearance compared to placebo [68]. Another double-blind randomized study among children with cerebral malaria showed the median recovery time from children in deep coma was reduced by half (and the rate of parasite clearance was improved) when DFO was added to treatment with quinine compared to the antimalarial treatment plus placebo [69].

Multiple iron biomarkers have been associated with malaria infection among various populations. The most widely reported is serum ferritin, a known acute-phase reactant protein that can increase in response to underlying infection or inflammation regardless of underlying iron stores [55,70–76]. Our data confirm serum ferritin is increased in the presence of malaria infection in pregnant women. The reported association between malaria and iron deficiency reported above adjusted for the effect of inflammation. Results from our pooled analysis did not find statistically significant associations between the remaining biomarkers and malaria. These results are in agreement with findings in the literature among non-pregnant populations where associations have been inconsistent between malaria and: sTfR [21,70,72,73,77,78], transferrin [72,73], and transferrin saturation [72].

Despite the evidence that serum ferritin increases as a result of pregnancy per se, infection or inflammation, it remains the international standard for defining iron deficiency [79]. This creates an inherent challenge to measuring iron deficiency using cutoffs of serum ferritin in pregnant populations living in malaria endemic settings (see also Figure S3.1 in File S1). Consensus exists...
that serum ferritin should be measured with inflammatory markers such as C-reactive protein, α1-antichymotrypsin, or α1-acid glycoprotein; however, agreement has yet to be made on the best inflammatory markers and their corresponding cutoff values for defining iron deficiency in the presence of inflammation [80–83]. Other iron biomarkers have been proposed to replace serum ferritin including sTfR. While it has been suggested sTfR is unaffected by pregnancy or inflammation [84], evidence of how pregnancy affects concentrations of sTfR is lacking [79]. The extent to which sTfR is affected by pregnancy will determine the appropriateness of using sTfR as a biomarker for detecting iron deficiency in pregnancy, especially in malaria endemic settings.

While the evidence of iron supplementation related to malaria risk in pregnancy is not of strong quality, the available data suggest iron supplementation does not increase the risk of malaria. Among the seven studies included in this review, only two of them represented primary data analyses from randomized trials, both of which failed to see an association between iron supplementation in pregnancy and malaria risk. No routine medical malaria prevention was used in these trials among the women included in this review, and they were conducted before the wide-scale introduction of insecticide treated nets [36,37]. The same applied to the study using a before-after design [39].

Prevalence of use of malaria prevention strategies varied greatly between the two surveys included for iron supplementation, and these showed the greatest variety in the association between iron supplementation and malaria [34,35]. One survey reported less malaria among iron supplemented (HIV-infected) women (figure 2) [34]; however, it should be noted, that we transformed their data from odds ratios to risk ratios to be able to combine the results of the different studies [34]. Kapito-Tembo (2010) reported an odds ratio of 0.4, 95% CI 0.14–1.12, in multivariable analysis, comparing iron supplemented vs. not supplemented when adjusting for age, gravidity, bed net use, socio-economic status, antenatal visits, IPTp use or cotrimoxazole prophylaxis [34].

The higher risk of malaria among women that used iron for >30 days in the subgroup analysis (Figure S2.3 in File S1) could potentially be explained by the association with IPTp [34,35]. Women who use iron for ≤30 days may be protected by the last dose of IPTp with SP if this was given at the visit when the iron supplementation commenced. SP is known to provide four to six weeks of post-treatment prophylaxis against malaria after which protective drug levels wane, placing women who continue using iron at risk until they receive the next dose of SP. In addition, some women may have initially been iron deficient, but with continued supplementation, have become iron replete, which may contribute to development of malaria [18,85]. Among children it has been suggested that an individual’s baseline iron status may modify the risk of malaria associated with iron supplementation; e.g. among iron-deficient persons, iron supplementation may decrease anemia, whereas among iron-replete persons, supplementation may have no benefits for anemia reduction and only result in an increased risk for malaria [21]. However, none of the iron supplementation studies included baseline iron status, although they all showed either a decrease in anemia (RRprimary 0.79, 95% CI 0.73–0.85, four studies, I2 31.7%, Figure S3.1 in File S1), or an improvement in hemoglobin (mean increase 0.79 g/dl, 95% CI 0.64–0.95, three studies, I2 15-6%, Figure S3.2 in File S1). While direct evidence of risk associated with iron supplementation is lacking, strong evidence exists that iron supplementation improves hemoglobin levels and decreases the risk of anemia and improves newborn iron stores [8,9,86].

This review is subject to several limitations, an examination of which may inform the design and conduct of future studies of iron supplementation during pregnancy in malaria endemic areas. First, the 31 studies identified were widely heterogeneous. Among the seven studies with data on iron supplementation, the cumulative dose of iron used and the timing and type of malaria testing varied as did the study design and methodology reducing comparability and preventing a pooled analysis across all available studies. Blood means used to be the standard for detecting malaria whereas now rapid diagnostic malaria tests, placental histology, and polymerase chain reactions are increasingly used, with the latter two tests having a higher sensitivity in detecting malaria [83]. Furthermore, the Entomological Inoculation Rate (a measure of malaria transmission in an area) and the prevalence of malaria varied greatly across these studies, as did the use of malaria prevention strategies, and the dose and duration of iron supplementation (table 1, 2, 3, 4). The distribution of biomarkers was not always normally distributed, and approximations of geometric means were made for ferritin. Consistency on these methodological aspects would allow for more accurate synthesis of the body of literature. The majority of these data are based on cross-sectional studies which provide evidence of an association between these factors, but cannot confirm a causal association between iron deficiency (measured by serum ferritin) and malaria risk. In addition, the direction of this association between malaria and iron status is unclear, as iron status may affect the malaria risk, but malaria infection itself causes an inflammatory response as well as hemolysis and will affect these measures of iron status. Lastly, with the exception of three studies, these data represent the risk associated with Plasmodium falciparum infections in Africa [38,46,63], and the effect of iron on malaria risk may differ by species and regions.

**Directions for Future Research**

While in children in malaria endemic areas there has been enough evidence of increased risk of morbidity associated with iron use to result in a more restricted policy in this population, surprisingly little is known of the risk-benefits in pregnancy and universal iron supplementation continues to be recommended during pregnancy. Because the relationship between malaria and iron status is difficult to disentangle, descriptive studies are not adequate, and randomized placebo controlled intervention trials may be the only way to answer the causal relationship between iron and malaria infection. However, these are costly and there may be ethical considerations due the need to withhold an intervention that is policy in most countries and the proven health benefit of iron supplementation in pregnant women. Using a clinical trial register, we identified five studies in various stages which will start to address some of these issues; baseline assessment of iron status will be important as well as the use of different definitions of iron deficiency, preferably not all ferritin-based [88–92].

These data suggest that iron supplementation may be safe in malaria endemic areas among women concurrently using malaria prevention strategies such as insecticide treated nets, monthly IPTp or daily cotrimoxazol (in HIV-infected women). Although IPTp and insecticide treated nets are currently the main policies to prevent malaria in pregnant women in sub-Saharan Africa, their coverage is less than optimal and it would be important to ensure that individual women receiving iron supplementation are indeed protected by an insecticide treated net and receive IPTp [93]. Because IPTp with SP is contraindicated in the first trimester, it is important to ensure that women using iron early in pregnancy are using an insecticide treated net and are screened for malaria at each scheduled visit until they can receive IPTp. With some countries moving towards iron fortification, it will also be prudent
Conclusion

Our review suggests iron supplementation may be safe during pregnancy in malaria endemic areas. However, the available data on iron supplementation in pregnancy and malaria risk are limited and insufficient to rule out any potential for an increased risk of malaria in malaria endemic settings. While iron deficiency (based on a definition with ferritin and CRP) is associated with a marked decreased risk of malaria in pregnancy, this association was not present among any of the other biomarkers of iron deficiency. Therefore, we find the results of iron deficiency and malaria risk to be inconclusive. Until more data are available from pregnant women, it would be prudent, based on the available evidence among children, to recommend that the provision of universal iron supplementation in pregnant women should always occur in conjunction with malaria prevention strategies during pregnancy in malaria endemic areas.

References

Association between Malaria and Iron in Pregnancy


