Rational Design, Synthesis and Biological Evaluation of Heterocyclic Quinolones Targeting the respiratory chain of *Mycobacterium tuberculosis*

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ABSTRACT

A High-throughput screen (HTS) was undertaken against the respiratory chain dehydrogenase component, NADH:menaquinone oxidoreductase (Ndh) of *Mycobacterium tuberculosis* (Mtb). 11,000 compounds were selected for the HTS based on the known phenothiazine Ndh inhibitors, trifluoperazine and thioridazine. Combined HTS (11,000 compounds) and in-house screening of a limited number of quinolones (50 compounds) identified ~100 hits and four distinct chemotypes, the most promising of which contained the quinolone core. Subsequent Mtb screening of the complete in-house quinolone library (350 compounds) identified a further ~90 hits across three quinolone sub-templates. Quinolones containing the amine based side chain were selected as the pharmacophore for further modification, resulting in metabolically stable quinolones effective against multi drug resistant (MDR) Mtb. The lead compound, MTC420 displays acceptable anti-tuberculosis activity (Mtb IC50 =525 nM, Mtb Wayne IC50 = 76 nM and MDR Mtb patient isolates IC50 = 140 nM) and favourable pharmacokinetic and toxicological profiles.

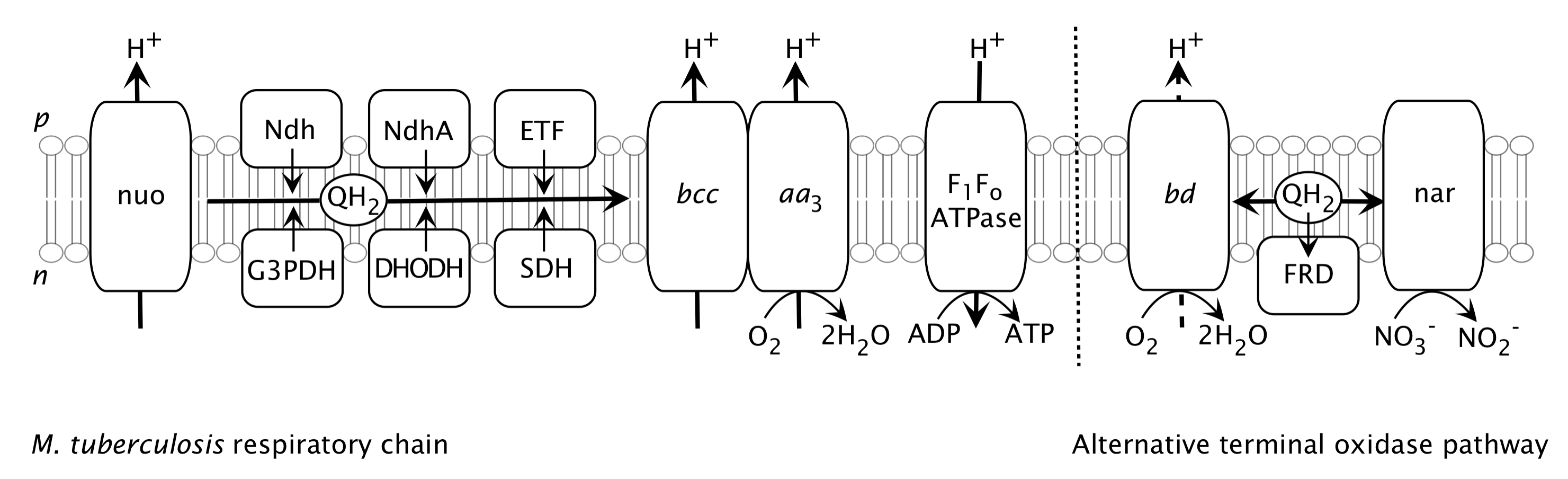
INTRODUCTION

In 2014 tuberculosis (TB) globally infected 9.6 million people resulting in an estimated 1.5 million deaths.1 With the emergence of multi-drug resistant (MDR) and extensively drug resistant (XDR) TB the need for new drug treatments targeting the disease has never been greater.2 Current first line drugs for TB were developed in 1952-1966 (Figure 1). Shortcomings of these drugs include; (i) long treatment regimens (6 to 9 months) leading to patient non-compliance, (ii) adverse drug-drug interactions with anti HIV drugs (HIV/AIDS is a common co-infection) and (iii) limited or no activity against MDR and XDR *Mycobacterium tuberculosis* (Mtb).3 Bedaquiline 4, 5 and delamanid 6, 7 are the only recently FDA approved drugs for the treatment of TB and their approval is currently only for MDR in cases where established treatments have failed (Figure 1).8 In order to find an effective treatment for MDR and XDR it is believed that a drug with a novel mode of action is required in order to circumvent resistance.



**Figure 1.** Current first line drugs used to treat tuberculosis and recently approved drugs for the treatment of MDR TB.

Targeting components of the Mtb respiratory chain (Figure 2) has been shown by us and other laboratories, to be effective in sterilizing both replicating and dormant Mtb.9-18 The initial target within this programme, Ndh (Rv1854c) is a single subunit 50 KDa enzyme involved in the redox reaction of NADH oxidation with subsequent menaquinol production. Ndh has been biochemically identified as a “choke point” and as such is essential for cell function and viability. 19 Essentiality of *ndh* has been shown by the inability of Mtb to tolerate insertion mutations in this gene20 and more recently in a study involving *ndh* knockout with subsequent confirmation by complementation.21 The other NADH-dependent electron donating dehydrogenases identified in the genome (Complex I and ndhA) have been shown not to be lethal.18, 22 These data are consistent with biochemical evidence that Ndh is a major source of electrons for the ETC.

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**Figure 2.** Schematic representation of the respiratory chain of *M. tuberculosis*. The chain components are Ndh/NdhA – type II NADH:(mena)quinone oxidoreductase (two isoforms), ETF – electron transferring flavoprotein (transfer of reducing equivalents from fatty acid β-oxidation into the Q-pool), nuo – protonmotive NADH dehydrogenase (Complex I), *bcc* – cytochrome *bcc* complex (note that there is no evidence for soluble cytochrome c in this organism), *aa*3 – cytochrome *bcc* oxidase, postulated to form a supercomplex with *bcc*. An alternative terminal oxidase pathway is utilised in *M. tuberculosis* under conditions of low oxygen tension, containing quinol oxidase (cytochrome *bd*), fumarate reductase (FRD) and nitrate reductase (nar) components. *P* and *n* correspond to the positive and negative sides of the respiratory membrane with respect to proton translocation. Proton movements are indicative only, and do not represent H+/e- ratios for the respective complexes.

Respiratory-chain inhibition-induced death represents a fundamental shift from traditional anti-tubercular drug design that have until recently relied on drugs that selectively target the replication machinery of Mtb. 9, 23-28 Anti-tubercular drugs developed to target the respiratory pathways should therefore have the potential to have sterilizing activity against current MDR and XDR Mtb strains.

Identification of hit compounds was achieved through a HTS screen of approximately 11,000 compounds that were predicted to possess activity against the Ndh enzyme. Ndh was chosen for the HTS due to the critical role as an important dehydrogenase during growth and pathogenicity9, 17 and due to its tractability for heterologous expression in *E.coli* and HTS29. The enzyme has been observed to be sensitive to phenothiazine-based inhibitors such as trifluoperazine and thioridazine9. These inhibitors have been shown by a number of different laboratories to have sterilizing activity against replicating and slow growing MDR Mtb (grown anaerobically) in both *in vitro* and *in vivo* models.14, 30, 31 These two compounds were used as the basis to employ a range of ligand-based chemoinformatics methods32-35 in the rational selection of the ~11,000 compounds for the HTS campaign (selected from a commercial library of ~750,000 compounds (Biofocus DPI)).36-40 Selected compounds were subject to a sequential high throughput screening campaign using an *in vitro* assay against recombinant Ndh as described previously.29

In addition to the HTS screen a limited selection of 50 quinolones were also screened in-house against Mtb Ndh. These compounds were selected for their structural diversity from a library of quinolones designed to target the NADH:ubiquinone oxidoreductase within the malaria parasite *Plasmodium falciparum* (*Pf*NDH2) as described previously.41-44 The HTS screen and in-house screen in combination generated ~100 hits across 4 distinct templates, the most potent of which were also tested for whole cell replicating Mtb activity. Following analysis of the *in vitro* biological data, predicted DMPK properties and investigations into chemical tractability the quinolone template was selected as the most promising for further development.

1. In previous antimalarial discovery projects41-48, the inhibitors based on the quinolone core displayed pharmacodynamics consistent of a privileged pharmacophore, with the ability to act on multiple electron transport chain (ETC) components. For example, quinolones with a dual mechanism of action against two respiratory enzymes, *Pf*NDH2 and cytochrome *b*c1 have recently been reported. 43 To exploit this phenomenon in this antitubercular discovery project, further screening and SAR investigations was switched to whole cell replicating TB activity. In order to fully establish the structure activity relationship (SAR) within the existing quinolone library with respect to whole cell Mtb activity a further library of ~350 compounds were screened against replicating Mtb. ~90 compounds were found to inhibit Mtb growth by >50% at 5 µM. Four sub-templates were then identified as having moderate *in vitro* Mtb potency. The most promising of which only had a very limited number of examples (see Table S1 – Supporting Information) within the existing library but demonstrated significantly more potency, as such the template based on compounds **1** and **2** was chosen for lead optimisation (Figure 3).



**Figure 3.** Identification of the quinolone template for lead optimisation.

A comprehensive medicinal chemistry SAR study around this series was then undertaken to establish optimised leads for further development. Screening data analysis (see Table S1 – Supplementary Information) shows NH2 and OAc at the 4-position are inactive for this particular sub-template (Table S1 - entries 20, 23 and 24) and show reduced activity for other quinolone sub-templates. Replacement of the phenyl ring with a pyridyl ring also rendered the sub-template inactive (Table S1 - entry 20). Modification of ring C resulting in loss of *in vitro* Mtb potency is a general trend that was seen across most quinolone sub-templates screened. Modifications of particular interest were therefore optimization of the side chain to optimize potency and DMPK, the nature of the group at 3-position and the electronic/steric effect of substituents placed at the 5, 6 and 7 positions (Figure 4).



**Figure 4.** Known SAR and SAR to be investigated.

CHEMISTRY

Following identification of quinolones **1** and **2** as the initial hits against Mtb, our initial efforts were focused on exploring the SAR of substituents placed in the A ring. The synthesis of these compounds was achieved in 3 – 5 steps from commercially available starting materials (Scheme 1). Oxazoline **4** was prepared from the corresponding isatoic anhydride **3** in yields of 34 – 75%. Where the isatoic anhydride was not commercially available, the oxazolines were synthesized in-house (see Supporting information). 4’-fluoropropiophenone **5** was reacted with piperidine to give ketone **6** in 32 – 97% yields. Reaction of oxazoline **4** with ketone **6** in the presence of triflic acid gave the desired quinolones **1**, **2**, **7a**-**k** in 23 – 45% yields.

**Scheme 1.** Synthesis of Quinolones **1, 2**, **7a-k**.*a*



*a* Conditions and reagents: (a) 2-amino-2-methyl-propanol, ZnCl2, PhCl, 135 oC, 24 h; (b) corresponding amine, K2CO3, DMF, 120oC to reflux, overnight ; (c) CF3SO3H, *n*-BuOH, N2, 130 oC, 24 h.

**Table 1.** Yields for the Synthesis of Compounds **1, 2, 7a-k**.



|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Compound | X | R | % yield **4** | % yield **6** | % yield **7** |
| **1** | H | H | - | 62 | 23 |
| **2** | 7-OMe | H | 75 | 62 | 36 |
| **7a** | 6-F | H | 60 | 62 | 26 |
| **7b** | 6-OMe, 7-OMe | H | 52 | 62 | 28 |
| **7c** | 6-Cl, 7-OMe | H | 45 | 62 | 35 |
| **7d** | 6-F, 7-OMe | H | 52 | 62 | 41 |
| **7e** | 5-OMe, 7-OMe | H | 58 | 62 | 32 |
| **7f** | 5-F,7-F | H | 68 | 62 | 45 |
| **7g** | 7-F | H | - | 62 | 35 |
| **7h** | 7-Cl | H | 64 | 62 | 37 |
| **7i** | H | F | - | 55 | 36 |
| **7j** | 7-OMe | F | 75 | 55 | 43 |
| **7k** | 5-F,7-F | F | 68 | 55 | 29 |

The nature of the group at 3-position of the quinolone was also studied. A small set of analogues with a hydrogen at 3-position were synthesized (Scheme 2). Substituted 2-aminoacetophenone **9** was converted from the respective aminobenzoic acid **8** using methyl lithium in 36% yield. 4-fluorobenzoate **10** was reacted with piperidine in the presence of potassium carbonate to give the piperidinyl benzoate **11** in 37% yield. Benzoate **11** was hydrolysed to benzoic acid which was then converted to acid chloride **12** by oxalyl chloride. Acylation of 2-aminoacetophenone **9** with acid chloride **12** provided the intermediate **13** in 30 – 51% yields. Cyclisation of the intermediate **13** in the presence of NaOH or KO*t*Bu gave the 3-H quinolones **14a-c** in 41 – 91% yields (Table 2).

Literature precedent from the development of ETC inhibitors in the antimalarial field lead us then to look at the presence of a halide at the 3-position. GSK’s pyridone series 49 demonstrated tolerance of the presence of a chlorine at 3-position and within our own group we have shown the combination of 3-chloro-7-methoxy enhances biological activity of the quinolone core.50 In order to achieve this the 3-H compounds were treated with sodium dichloroisocyanurate and sodium hydroxide to give 3-Cl quinolones **15a-d** in 40 – 61% yields, or NBS to give 3-Br quinolones **15e-f** in 55 – 63% yields.

**Scheme 2.** Synthesis of quinolones **14 a-c** and **15 a-f**.*a*



*a* Conditions and reagents: (a) MeLi, DME, 0oC, 2 h; (b) (i) K2CO3, DMF, reflux, overnight, (ii) NaOH (aq), MeOH, reflux, overnight; (c) oxalyl chloride, DCM, DMF (cat.), r.t., 2 h; (d) NEt3, THF, r.t., overnight; (e) NaOH (s), 1,4-dioxane, 110oC, 5 h or KO*t*Bu, *t*BuOH, 75oC, 16 h; (f) sodium dichloroisocyanurate, 1M NaOH (aq), MeOH, r.t., overnight (**15a-d**) or NBS, DCM, DMF, r.t., overnight (**15e-f**).

**Table 2.** Yields for the Synthesis of Compounds **14a-c** and **15a-f**.



|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Compound | X | R | Y | % yield **9** | % yield **11** | % yield **13** | % yield **14** | % yield **15** |
| **14a** | H | H | H | - | - | 51 | 70 | - |
| **14b** | OMe | H | H | 36 | - | 50 | 41 | - |
| **14c** | OMe | F | H | 36 | 37 | 30 | 68 | - |
| **15a** | H | H | Cl | - | - | 51 | 70 | 40 |
| **15b** | OMe | H | Cl | 36 | - | 50 | 41 | 61 |
| **15c** | OMe | F | Cl | 36 | 37 | 30 | 68 | 52 |
| **15d** | H | F | Cl | - | 37 | 60 | 91 | 58 |
| **15e** | H | H | Br | - | - | 51 | 70 | 63 |
| **15f** | H | F | Br | - | 37 | 60 | 91 | 55 |

Having identified 3-methyl and 5, 7-difluoro quinolone (followed by 6-fluoro-7-methoxy and 7-methoxy quinolone) to be optimal for Mtb activity (see Table 8), the focus of SAR explorations moved to the terminal ring of the side chain to further improve Mtb activity and optimise DMPK. Additional small groups, such as Me, F and CF3 attached at different positions on the terminal piperidine ring were investigated. In addition the effect of chirality was explored.51 Synthesis of compounds **17a-k** was achieved using chemistry described in Scheme 3.

**Scheme 3.** Synthesis of Quinolones **17a-k**.*a*



*a* Conditions and reagents: (a) corresponding amine, K2CO3, DMF, 120oC to reflux, overnight ; (b) CF3SO3H, *n*-BuOH, N2, 130 oC, 24 h.

**Table 3.** Yields for the Synthesis of Compounds **17a-k**.



|  |  |  |  |
| --- | --- | --- | --- |
| Compound | R | % Yield **16** | % Yield **17** |
| **17a** |  | 48 | 32 |
| **17b** |  | 73 | 54 |
| **17c** |  | 48 | 34 |
| **17d** |  | 40 | 57 |
| **17e** |  | 64 | 27 |
| **17f** |  | 84 | 45 |
| **17g** |  | 74 | 43 |
| **17h** |  | 75 | 40 |
| **17i** |  | 72 | 39 |
| **17j** |  | 69 | 41 |
| **17k** | -NHCH2Ph | 28 | 51 |

Incorporation of different amino groups into the side chain as an alternative to the potentially metabolically labile piperidine ring was also investigated. To incorporate a diethylamine group an alternative methodology was used to synthesise the side chain, commercial available 4-bromo-*N*,*N*-dimethylaniline **18** was treated with butyllithium for a lithium-halogen exchange and the intermediate was quenched with *N*,*N*-dimethylpropionamide to form the side chain **19** in 78% yield, reaction with oxazoline **4h** was then carried out to give quinolone **17l** in 46% yield (Scheme 4).

**Scheme 4.** Synthesis of quinolone **17l**.*a*



*a* Conditions and reagents: (a) (i) *n*BuLi, Et2O, -78oC, 30 min; (ii) *N*,*N*-dimethylpropionamide, -78oC to r.t., 2 h; (b) CF3SO3H, *n*-BuOH, N2, 130 oC, 24 h.

Extension of the side chain with a phenyl or benzyl group at the 2-position was also investigated using the synthetic methodologies shown in Scheme 5. In addition, replacement of piperidine by piperazine was investigated. This was to further explore the length of side chain that could be tolerated and to improve the solubility.

**Scheme 5.** Synthesis of Quinolones **21a-g**.*a*



*a* Conditions and reagents: (a) corresponding amine, K2CO3, DMF, 120oC to reflux, overnight; (b) CF3SO3H, *n*-BuOH, N2, 130 oC, 24 h.

**Table 4.** Yields for the Synthesis of Compounds **21a-g**.



|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Compound | X | A | B | % Yield **20** | % Yield **21** |
| **21a** | H | CH | CH2Ph | 64 | 33 |
| **21b** | 6-F | CH | CH2Ph | 64 | 40 |
| **21c** | 7-OMe | CH | CH2Ph | 64 | 42 |
| **21d** | H | N | CH2Ph | 58 | 30 |
| **21e** | 6-F | N | CH2Ph | 58 | 28 |
| **21f** | 7-OMe | N | Ph | 64 | 30 |
| **21g** | 7-OMe | N | CH2Ph | 58 | 38 |

In addition, the quinolone with a piperidine ring at the meta-position **24** was also synthesised by reacting the 3-bromopropiophenone **22** with piperidine using Buchwald coupling to yield the ketone intermediate **23**, which was coupled with oxazoline **4h** to give the quinolone in 45% yield (Scheme 6).

**Scheme 6.** Synthesis of Quinolone **24**.*a*



*a* Conditions and reagents: (a) Piperidine, Pd(OAc)2, XPhos, NaO*t*Bu, Toluene, 110 oC, 24 h; (b) CF3SO3H, *n*-BuOH, N2, 130 oC, 24 h.

A series of analogues with a pyrrole heterocycle in the side chain were also synthesized to further explore the side chain SAR and enhance the metabolic stability. The synthetic route to these compounds is illustrated in Scheme 7. Utilizing Copper and trans-*N*,*N*’-Dimethyl-1,2cyclohexanediamine catalyzed *N*-arylation with 4-bromopropiophenone the side chain ketone intermediate **31** was formed in 30 – 62 % yields.52, 53 Final cyclisation with oxazoline gave quinolones **32a-g** in 35 – 57% yields.

**Scheme 7.** Synthesis of quinolones **32a-g**.*a*



*a* Conditions and reagents: (a) EtMgBr, THF, 0 oC, 1h; (b) PCC, DCM, r.t., 2h; (c) 5mol% CuI, 20mol% trans-*N*,*N*’-Dimethyl-1,2cyclohexanediamine, K3PO4, Toluene, 110oC, 24 h; (d) CF3SO3H, *n*-BuOH, N2, 130 oC, 24 h.

**Table 5.** Yields for the Synthesis of Compounds **32a-g**.



|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Compound | X | Y | R | % yield **31** | % yield **32** |
| **32a** | 5-F,7-F | - |  | 38 | 55 |
| **32b** | 5-F,7-F | - |  | 30 | 57 |
| **32c** | 5-F,7-F | - |  | 46 | 35 |
| **32d** | 5-F | *-* |  | 62 | 32 |
| **32e** | 5-F,7-F | - |  | 62 | 30 |
| **32f** | 5-F,7-F | *m*-Cl |  | 49 | 39 |
| **32g** | 5-F,7-F | *o*-F |  | 52 | 39 |

Using fluorine to block metabolism and improve oral absorptions was further explored. Research by Smith has shown that gem-difluorinated piperidine compounds exhibited a significant improvement in metabolic stability.54 This led to the design and synthesis of fluorinated quinolones **38a-f** as well as the alcohol side chain quinolones **38g-i**. The chemistry used in the synthesis of these compounds is shown in Scheme 8.

**Scheme 8.** Synthesis of Quinolones **38a-l**.*a*



*a* Conditions and reagents: (a) corresponding amine, K2CO3, DMF, 120oC to reflux, overnight; (b) CF3SO3H, *n*-BuOH, N2, 130 oC, 24 h.

**Table 6.** Yields for the Synthesis of Compounds **38a-l**.



|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Compound | X | R | % Yield **37** | % Yield **38** |
| **38a** | 5-F,7-F |  | 38 | 45 |
| **38b** | 5-F,7-F |  | 37 | 47 |
| **38c** | 5-F,7-F |  | 25 | 33 |
| **38d** | 5-F,7-F |  | 32 | 30 |
| **38e** | 7-OMe |  | 32 | 32 |
| **38f** | 6-Cl,7-OMe |  | 32 | 30 |
| **38g** | 5-F,7-F |  | 69 | 48 |
| **38h** | 5-F,7-F |  | 54 | 50 |
| **38i** | 5-F,7-F |  | 32 | 43 |
| **38j** | 5-F,7-F |  | 41 | 20 |
| **38k** | 5-F,7-F |  | 43 | 25 |
| **38l** | 5-F,7-F |  | 41 | 37 |

Removal of the benzyl group from the chiral proline derivatives **38j-l** was achieved using hydrogenation (Scheme 9) in good yields.

**Scheme 9.** Synthesis of compounds **39a-c.**



For the gem-difluoro analogues (**42a** (MTC420) and **42b**), 4-bromopropiophenone was first converted to a more reactive 4-iodopropiophenone by an aromatic Finkelstein reaction catalysed by copper(I) iodide in combination with *N*,*N*-dimethyl-1,2-diaminoethane.55 A subsequent Buchwald-Hartwig amination using Pd2(dba)3 and Xantphos with the gem-fluorinated amine gave the ketone side chain **41a-b** in 12 – 28% yields.56 Reaction with oxazoline gave quinolones **42a-b** in 47 – 56% yields (Scheme 10).

**Scheme 10.** Synthesis of quinolones **42a-b**.*a*



*a* Conditions and reagents: (a) CuI, *N*,*N*-dimethyl-1,2-diaminoethane, NaI, 1,4-dioxane, 110oC, 24 h; (b) Pd2(dba)3, Xantphos, NaO*t*Bu, 1,4-dioxane, 110oC, 24 h; (c) CF3SO3H, *n*-BuOH, N2, 130 oC, 24 h.

**42a** was identified as the lead compound in the series as it exhibited good potency and metabolic stability (See Table 11 and Table 12), further investigation of the pyrrolidine side chain was undertaken to improve solubility and potency. Further modifications have included adding chirality and introducing amide functionality to rapidly ascertain if it is tolerated within the template. Quinolones **45a-h** were therefore synthesised using chemistry described in Scheme 11. To incorporate the amide group, Ullmann coupling of 4-bromopropiophenone with *D*-proline gave the carboxylic acid intermediate **43a-b**. Crosslinking the carboxylic acid by EDC/NHS to respective amine provided the ketone side chain **44** in 52 – 90% yields. This was subsequently coupled with oxazoline in 12 – 34% yields to afford quinolones **45a-g.**

**Scheme 11.** Synthesis of Quinolones **45a-g**.*a*



*a* Conditions and reagents: (a) D-proline, CuI, K2CO3, DMF, 140oC, 24 h; (b) (i) EDC, N-hydroxysuccinamide, CHCl3, NEt3, amine, r.t., 6 h; (ii) amine, NEt3, r.t., 2h; (c) CF3SO3H, *n*-BuOH, N2, 130 oC, 24 h.

**Table 7.** Yields for the Synthesis of Compounds **45a-g**.



|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Compound | R | n | % yield **44** | % yield **45** |
| **45a** | -NHtBu | 2 | 52 | 20 |
| **45b** | -NEt2 | 2 | 80 | 34 |
| **45c** |  | 2 | 90 | 25 |
| **45d** |  | 2 | 70 | 18 |
| **45e** |  | 2 | 65 | 24 |
| **45f** | -NMe2 | 1 | 45*a* | 15 |
| **45g** | -NHtBu | 1 | -*b* | 12 |

*a*Alternative methodology used please see supporting information. *b* Used crude.

Incorporation of an amide moiety largely resulted in reduced anti-tuberculosis activity (Table 7). As such our attention returned to **42a** and improving its pharmacokinetic profile. Use of a pro-drug strategy, previously used successfully within other quinolone development programs57 was investigated leading to the synthesis of compound **46**. Compound **46** was synthesized by reacting **42a** with potassium *tert*-butoxide and acetyl chloride to give the acetate pro-drug in high yield.

**Scheme 12.** Synthesis of pro-drug **46**. *a*



*a* (a) i. *t*BuOK, THF, r.t., 1h. ii. Acetyl chloride, r.t., 3h.

RESULTS AND DISCUSSION

***Structure Activity Relationships (SAR)*** *-*Initial SAR investigations around the hit compounds **1** and **2** focused on establishing the optimal A-ring substituents (X). Compounds **1**, **2** and **7a-7h** demonstrate the most favorable X groups are 5-F, 7-F closely followed by 6-F, 7-OMe and 7-OMe. Compounds **7i-7k** were synthesised with a view to reducing the potential metabolism of the piperidine ring. Pleasingly a good level of potency was maintained. Concomitantly the potential for replacing the methyl group at Y was also investigated. When Y=H activity is lost as demonstrated by compounds **14a-c**. Halogenation was also investigated; again this largely resulted in reduced anti-tuberculosis activity (**15a-f**). The one exception to this being **15e** possessing a Br at Y. This affect appeared to be compound specific rather than a general trend across all brominated analogues and as such it was decided that the methyl group was the optimal group at this position.

**Table 8.** Mtb IC50 values for compounds **1, 2, 7a-k**, **14a-c** and **15a-f**.



|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Compound | X | Y | R | Mtb IC50 (µM) |
| **1** | H | Me | H | 1.50 ± 0.19 |
| **2** | 7-OMe | Me | H | 0.73 ± 0.01 |
| **7a** | 6-F | Me | H | 1.83 ± 0.22 |
| **7b** | 6-OMe, 7-OMe | Me | H | >10 |
| **7c** | 6-Cl, 7-OMe | Me | H | >10 |
| **7d** | 6-F, 7-OMe | Me | H | 0.52 ± 0.06 |
| **7e** | 5-OMe, 7-OMe | Me | H | >10 |
| **7f** | 5-F, 7-F | Me | H | 0.27 ± 0.08 |
| **7g** | 7-F | Me | H | >10 |
| **7h** | 7-Cl | Me | H | >10 |
| **7i** | H | Me | F | >10 |
| **7j** | 7-OMe | Me | F | 1.32 ± 0.10 |
| **7k** | 5-F, 7-F | Me | F | 0.94 ± 0.12 |
| **14a** | H | H | H | >10 |
| **14b** | 7-OMe | H | H | >10 |
| **14c** | 7-OMe | H | F | >10 |
| **15a** | H | Cl | H | 1.56 ± 0.22 |
| **15b** | 7-OMe | Cl | H | 2.82 ± 0.21 |
| **15c** | 7-OMe | Cl | F | >10 |
| **15d** | H | Cl | F | >10 |
| **15e** | H | Br | H | 0.60 ± 0.09 |
| **15f** | H | Br | F | >10 |

With 5-F, 7-F and 3-methyl confirmed as optimal for anti-tuberculosis activity optimising the side chain then became the focus of the SAR studies (Table 9). Initial investigations into piperidine ring substituents at the 4-position revealed that in addition to 4-F **7k**, a methyl group is also tolerated as demonstrated with compound **17b**. It rapidly became apparent that there was a size limitation to the group tolerated at the 4-position with larger groups such as CF3, cyclopropyl and gem-difluoro resulting in loss of potency. Movement of the F and Me groups to the 3-position resulted in improvements in anti-tuberculosis activity as demonstrated by compounds **17e-h**. Interestingly racemic and enatiomerically pure analogues of the 3-methyl derivative **17f** showed little variation in potency, which is in direct contrast to the pyrrolidine analogues discussed later. Replacement of the piperidine ring with a number of alternative amines was also investigated. Increasing ring size (**17j**) and use of dimethyl amine (**17l**) retained good potency. Incorporation of secondary amines (**17k**) and more polar groups such as *N*-methyl piperazine (**17i**) reduced anti-tuberculosis activity. Moving the piperidine group from the *para* to the *meta*-position (**24**) also resulted in loss of activity.

**Table 9.** Mtb IC50 values for compounds 1**7a-l** and **24**.



|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Compound | R | Mtb IC50 (µM) | Compound | R | Mtb IC50 (µM) |
| **17a** |  | >10 | **17h** |  | 0.47 ± 0.02 |
| **17b** |  | 0.61 ± 0.05 | **17i** |  | >10 |
| **17c** |  | >10 | **17j** |  | 0.49 ± 0-07 |
| **17d** |  | >10 | **17k** | -NHCH2Ph | >10 |
| **17e** |  | 0.31 ± 0.03 | **17l** |  | 0.41 ± 0.002 |
| **17f** |  | 0.37 ± 0.04 | **24** | *meta* | >10 |
| **17g** |  | 0.47 ± 0.03 |  |  |  |

The size limitation and unfavorable incorporation of piperazine was further confirmed by our concomitant investigation in to extended side chain analogues (Table 10). The aim of this series was to explore the space available and to improve solubility with the incorporation of piperazine to facilitate salt based formulation.

**Table 10.** Mtb IC50 values for compounds **21a-g**.



|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Compound | X | A | B | Mtb IC50 (µM) |
| **21a** | H | CH | CH2Ph | >10 |
| **21b** | 6-F | CH | CH2Ph | >10 |
| **21c** | 7-OMe | CH | CH2Ph | >10 |
| **21d** | H | N | CH2Ph | 5.74 ± 0.66 |
| **21e** | 6-F | N | CH2Ph | >10 |
| **21f** | 7-OMe | N | Ph | >10 |
| **21g** | 7-OMe | N | CH2Ph | >10 |

With this information in hand several small heterocyclic, fluorinated, chiral and amide analogues were synthesized to investigate SAR and improve DMPK (Table 11). Compounds **32a-g** are pyrrole derivatives. An unsubstituted pyrrole moiety is well tolerated in the 5-F (**32d**) and 5-F, 7-F (**32e**) analogues, however increasing the size of the pyrrole group by addition of a fused benzene ring (**32b**) again results in loss of potency. Incorporation of a halogen on the aromatic ring was also investigated but reduced potency.

**Table 11.** Mtb IC50 values for compounds **32a-g**, **38a-j**, **39a-c**, **42a-b** and **45a-g**.



|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| Compound | X | R | Mtb IC50 (µM) | Compound | X | R | Mtb IC50 (µM) |
| **32a** | 5-F,7-F |  | >10 | **38i** | 5-F,7-F |  | >10 |
| **32b** | 5-F,7-F |  | >10 | **38j** | 5-F,7-F |  | 0.96 ± 0.06 |
| **32c** | 5-F,7-F |  | >10 | **39a** | 5-F,7-F |  | 1.59 ± 0.05 |
| **32d** | 5-F |  | 0.71 ± 0.05 | **39b** | 5-F,7-F |  | 0.32 ± 0.04 |
| **32e** | 5-F,7-F |  | 0.44 ± 0.02 | **39c** | 5-F,7-F |  | >10 |
| **32f** | 5-F,7-F  Y= *m*-Cl |  | >10 | **42a** | 5-F,7-F |  | 0.53 ± 0.08 |
| **32g** | 5-F,7-F  Y= *o*-F |  | >10 | **42b** | 5-F,7-F |  | 0.36 ± 0.04 |
| **38a** | 5-F,7-F |  | 0.23 ± 0.003 | **45a** | 5-F,7-F |  | 0.96 ± 0.05 |
| **38b** | 5-F,7-F |  | 1.80 ± 0.09 | **45b** | 5-F,7-F |  | >10 |
| **38c** | 5-F,7-F |  | 1.53 ± 0.04 | **45c** | 5-F,7-F |  | >10 |
| **38d** | 5-F,7-F |  | >10 | **45d** | 5-F,7-F |  | >10 |
| **38e** | 7-OMe |  | >10 | **45e** | 5-F,7-F |  | >10 |
| **38f** | 6-Cl,7-OMe |  | >10 | **45f** | 5-F,7-F |  | >10 |
| **38g** | 5-F,7-F |  | 5.01 ± 0.03 | **45g** | 5-F,7-F |  | >10 |
| **38h** | 5-F,7-F |  | >10 |  |  |  |  |

Fluorinated analogues were synthesized in order to improve metabolic stability (see Table 11). Both mono (**38a** and **38b**) and gem-difluoro (**42a**) substituted pyrrolidine derivatives exhibited good to excellent potency. The gem-difluoro azetidine (**38c**) and 3-substituted piperidine (**42b**) also demonstrated good potency. Incorporation of an alcohol group in the side chain to reduce lipophilicity and potentially facilitate pro-drug approaches provided mixed results. Gem-methyl, OH analogues **38g-i** were not tolerated whereas inclusion of prolinol (**39a-b**) gave good anti-tuberculosis activity. Benzylated analogue **38j** and amide analogues **45a-g** largely resulted in loss of potency. For the pyrrolidine analogues the effect of chirality on activity was marked with the (*R*)-3-fluoro analogue **38a** (Mtb IC50 = 0.23 µM) demonstrating significantly superior potency over the (*S*)-3-fluoro analogue **38b** (Mtb IC50 = 1.80 µM). The effect of chirality was also observed with the prolinol analogues, (*S*)-prolinol analogue **39b** (Mtb IC50 = 0.32 µM) being more active than (*R*)-prolinol analogue **39a** (Mtb IC50 = 1.52 µM). The overall SAR trends for the series can be seen in Figure 5.



**Figure 5.** Overall SAR trends for the heterocyclic quinolone series.

***In vitro DMPK and toxicity*** - Analogues demonstrating good potency were then moved through our screening cascade and evaluated for microsomal turnover and HEPG2 cytotoxicity. None of the compounds were found to be cytotoxic and all had good therapeutic indexes. From the earlier analogues tested (entries 1-6 in Table 12) it was apparent that the compounds were being metabolised quickly by liver microsomes. Resolving this issue was therefore the driving force for a large proportion of the medicinal chemistry manipulations described in Table 11 above.

**Table 12.** HEPG2 and microsomal turnover t1/2 for selected analogues.

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Compound | Mtb  IC50  (µM) | Mtb  IC90  (µM) | HEPG2 GLU  (µM) | Therapeutic Index | Microsomal Turnover (h, m, r) t1/2 (min) |
| **7f** | 0.270 ± 0.080 | 0.78 | >100 | >370 | h-7.31  m-8.27  r-8.30 |
| **7k** | 0.950 ± 0.120 | 1.83 | 102.2 | 108 | h-5.7  m-4.4  r-8.4 |
| **17b** | 0.611 ± 0.048 | 1.93 | >100 | >164 | h-<10  m-<10  r-<10 |
| **17e** | 0.300 ± 0.025 | 0.56 | 188.1 | 627 | h-7.8  m-6.8  r-10.1 |
| **17f** | 0.367 ± 0.040 | 0.63 | >100 | >272 | h-7.9  m-22.3  r-10.8 |
| **17h** | 0.400 ± 0.023 | 0.66 | 85.54 | 223 | h-8.54  m-7.65  r- 5.72 |
| **32e** | 0.432 ± 0.020 | 0.69 | 141 | 342 | h-10.2  m-20.7  r-30.1 |
| **38a** | 0.231 ± 0.036 | 0.50 | 150.6 | 649 | h-10.2  m-4.4  r-10.6 |
| **38d** | >10 | >10 | ND | ND | h-60  m-60  r-60 |
| **42a** | 0.525 ± 0.080 | 1.10 | >100 | >190 | h-72.8  m-114.9  r-61.6 |
| **42b** | 0.361 ± 0.041 | 0.83 | ND | ND | h-17.4  m-16.2  r-13.7 |

Two strategies were employed to address the metabolic stability issues (Figure 6). The first was to replace the piperidine ring with an alternative heterocycle. Amongst those selected pyrrole (**32e**) provided the most active compound with a modest improvement in metabolic stability. Fluorination of the pyrrole (**38d**) at the 3 and 4 positions resulted in complete resolution of metabolic instability; however anti-tuberculosis activity was also lost. From earlier SAR studies we knew that replacing the piperidine ring (**7f**) with a pyrrolidine ring (**17l**) was tolerated in terms of activity and may provide us with more opportunity to modify the ring in what we believe to be a limited space. Mono-fluorination (**38a**) provided a very modest improvement in stability. Subsequent synthesis of the gem-difluoro analogue (**42a**) however provided us with a compound with both good anti-tuberculosis activity and excellent metabolic stability. The equivalent six membered ring analogue **42b** had good potency but comparatively decreased metabolic stability as expected (Table 12).

**Figure 6.** Resolution of metabolic stability problems.

Selected analogues were also measured for Caco-2 permeability, stability in plasma, % plasma protein binding (PPB) and solubility (Table 13). All compounds performed well in these assays with the exception of solubility which is a common issue for the quinolone chemotype.

**Table 13.** Caco-2 permeability, stability in plasma, % PPB and solubility values for selected analogues.

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| Compound | Caco-2 permeability  (cm-1/s) | Stability in plasma (r,h) T1/2 (min) | Human PPB  (%) | Solubility (µg/mL)    pH1 pH7.4 CMa | | |
| **5k** | ND | r->180  h->180 | 95.82 | >150 | <1 | 12 |
| **17e** | 22.86 x 10-6 | r->180  h->180 | 98.45 | >150 | <1 | 10 |
| **32e** | 30.97 x 10-6 | r->180  h->180 | 96.1 | 5.1 | 3.6 | 61 |
| **38b** | 15.51 x 10-6 | r->180  h->180 | 98.97 | <1 | <1 | 2.5 |
| **42a** | 10.00 x 10-6 | r->180  h->180 | 97.30 | < 1 | < 1 | 55 |

a CM – culture media - Middlebrook 7H9 broth with addition of 10% albumin–dextrose–catalase solution (Becton Dickinson), 0.2% [vol/vol] glycerol and 0.05% [vol/vol] Tween 80.

A number of analogues also underwent additional *in vitro* DMPK (Table 14) experiments further confirming the metabolism issues detailed above.

**Table 14.** *In vitro* DMPK measurements for selected analogues.

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Compound | Aqueous Solubility (µM) | Human % PPB | LogD7.4 | Human Microsomes CLint (µL/min/mg) | Rat Hepatocytes CLint (µL/min/106cells) |
| **7d** | 2 | 98.8 | 3.9 | > 300.0 | 231.3 |
| **15b** | 0.5 | 98.8 | 3.6 | > 300.0 | 48.9 |
| **17g** | < 0.5 | 99.5 | 4.7 | > 300.0 | 183.4 |
| **17h** | < 0.3 | 99.3 | > 3.2 | > 300.0 | 91.2 |
| **17j** | < 0.1 | 99.4 | 4.8 | > 300.0 | 243.4 |
| **38a** | 0.9 | 99.1 | 3.8 | 174.9 | 117.6 |
| **38b** | 1 | 98.6 | 3.6 | 197.4 | 150.1 |
| **39a** | 4 | 95.5 | > 3.4 | > 300.0 | 36.5 |
| **39b** | 4 | 95.5 | > 3.4 | > 300.0 | 36.5 |
| **42a** | 0.2 | 99.7 | 4 | 89.7 | 52.2 |

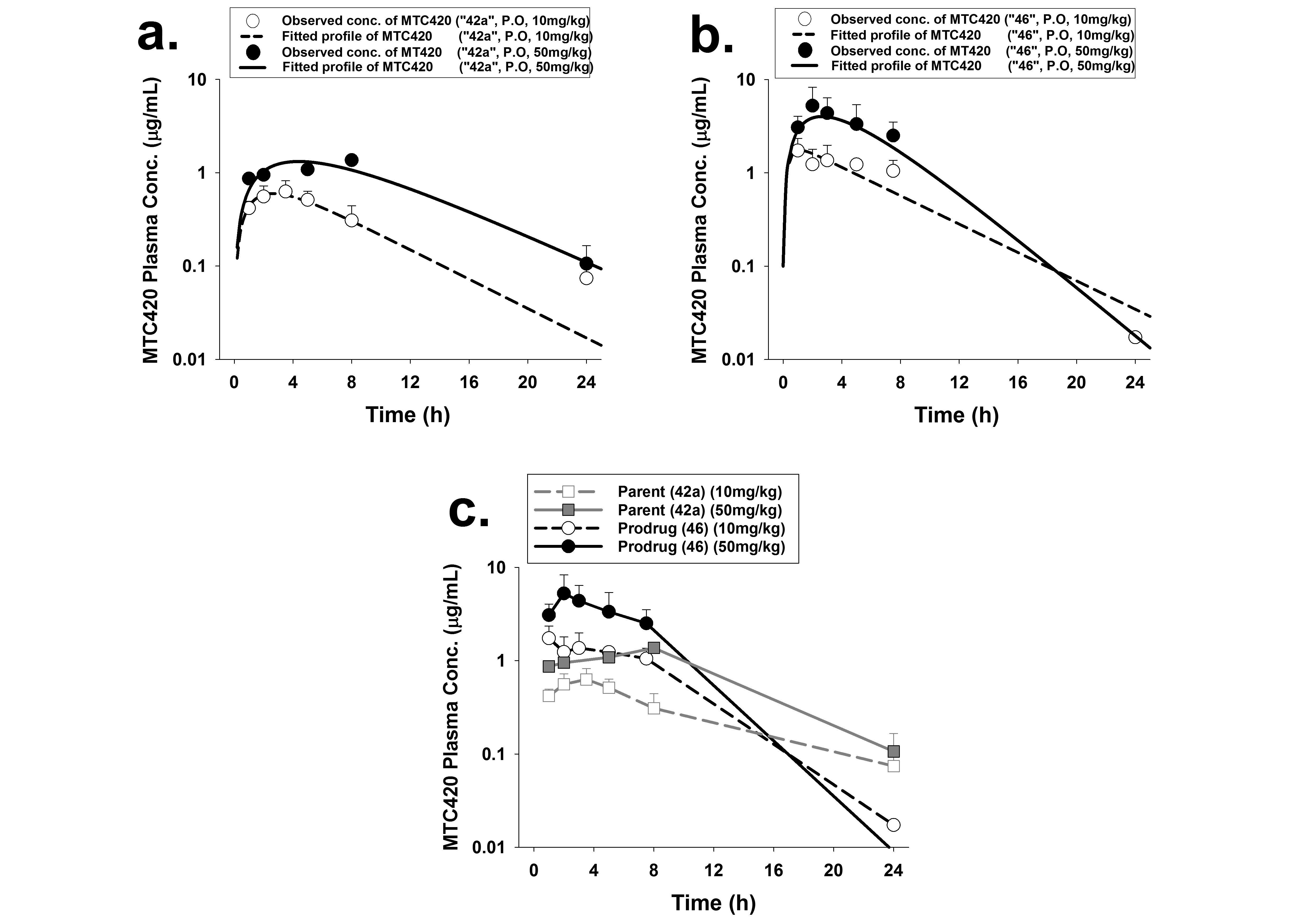
***Biological profile*** - Having selected **42a** as the lead compound, full biological profiling was undertaken to establish its pharmacokinetic and toxicological profile in addition to its activity against slow-growing (Wayne assay) and MDR-resistant Mtb (Table 15).

**Table 15.** Biological profile of **42a**.

|  |  |
| --- | --- |
|  | |
| ***In vitro* anti-tuberculosis activity** |  |
| Replicating sensitive Mtb IC50 (µM) | 0.525 |
| Replicating sensitive Mtb IC90 (µM) | 1.10 |
| Dormant (Wayne Model) Mtb IC90 (µM) | 0.076 |
| MDR Mtb (05TB42059) IC50 (µM) | 0.140 |
| MDR Mtb (DQ707(S315N kat G)) IC50 (µM) | 0.548 |
| ***In vitro* DMPK** |  |
| Microsomal Turnover (h, m, r) T1/2 (min) | h-72.8, m-114.9, r-61.6 |
| Microsomal Clint (h, m, r) (µL/min/mg) | h-9.52, m-6.03, r-11.25 |
| Caco-2 permeability (cm-1/s) A to B | 10.00 x 10-6 |
| Caco-2 permeability (cm-1/s) B to A | 9.8 x 10-6 |
| Stability in plasma (r,h) T1/2 (min) | r->180, h->180 |
| Human % PPB | 97.30 |
| Solubility (µg/mL) pH1, pH7.4, CM | <1, <1, 55 |
| CYP2C8 Inhibition (% at 10 µM) | 38 |
| CYP2C9 Inhibition (% at 10 µM) | 0 |
| CYP2D6 Inhibition (% at 10 µM) | 0 |
| CYP3A4 Inhibition (% at 10 µM) | 0 |
| CYP3A5 Inhibition (% at 10 µM) | 0 |
| ***In vitro* toxicity** |  |
| HEPG2 IC50 GLU (µM) | >100 |
| TI | >190 |
| hERG IC50 (µM) | >25 |
| Ames | -ve |

**42a** demonstrated comparable activity against all tested strains of sensitive and MDR Mtb as well as having good potency against dormant, non-replicating TB. It demonstrated a suitable *in vitro* DMPK and toxicity profile to undergo *in vivo* pharmacokinetic analysis.

***Pharmacokinetics*** - the pharmacokinetic profile of **42a** can be seen in Figure 7 and Table 16. Analysis of data from the parent compound indicated solubility limited absorption as the PK did not increase linearly with dose from 10 mg/kg to 50 mg/kg. At this point the acetate pro-drug strategy was deployed in an attempt to improve exposure.



**Figure 7.** Pharmacokinetics after oral dosing of **42a (a.)**, **46 (b.)** and an overlay of both **(c.)**

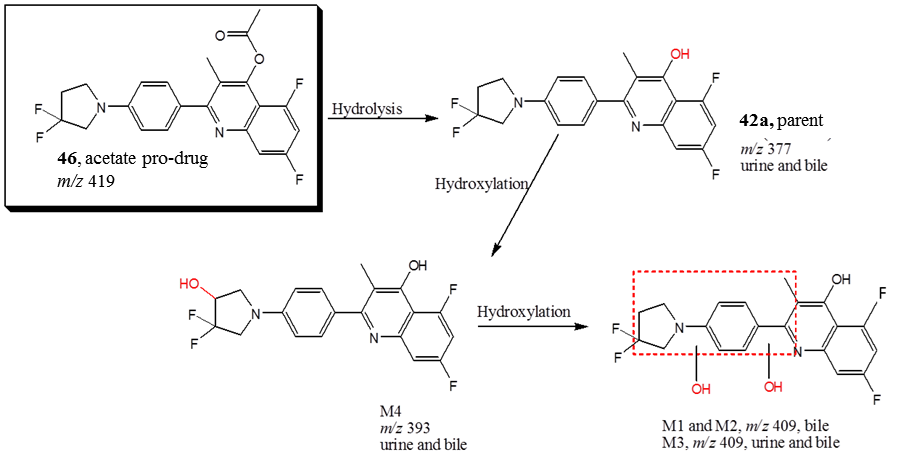
**Table 16.** Pharmacokinetic parameters for **42a** and **46**.

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
|  | Parent 42a | | | Prodrug 46\* | |
| **Dose (mg/kg)** | **0.5 (iv)** | **10 (po)** | **50 (po)** | **10 (po)** | **50 (po)** |
| T1/2 (h) | 1.48 | 3.8 | 4.2 | 3.9 | 2.3 |
| CL (L/h/kg) | 0.524 | - | - | - | - |
| Vss (L/kg) | 0.291 | - | - | - | - |
| Cmax (µg/mL) | - | 0.61 | 1.4 | 1.7 | 4.0 |
| AUC (mg.h/L) | 0.964 | 5.4 | 16.5 | 12.3 | 29.6 |
| Oral Bioavailability (% F) | N/A | 28.0 | 17.1 | 63.8 | 30.7 |

\*These two studies were dosed with prodrug **46** orally, and measured for the parent **42a** in plasma.

Initial findings with both the 10 mg/kg and 50 mg/kg dose of pro-drug demonstrated a significant increase in overall exposure as indicated by a significantly increased AUC, Cmax accompanied with increased bioavailability.

Metabolite ID work was undertaken to establish the metabolic activity exerted upon **46** (Figure 8 and Table 17).



**Figure 8.** Metabolic pathways of pro-drug **46** in SD rat urine and bile.

### Table 17. Identified metabolites of pro-drug 46 in SD rat urine and bile (MS)

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| Peak ID | Mass Shift | Found *m/z* | Biotransformation | R.T(min) | Relative MS Abundance | |
| Bile | Urine |
| **46** | 0 | 419 | Parent | 14.3 | ND | 1.85E+07 |
| **M1** | -10 | 409 | Hydrolysis/ Hydroxylation | 8.6 | 5.89E+06 | ND |
| **M2** | -10 | 409 | Hydrolysis/ Hydroxylation | 9.2 | 4.21E+06 | ND |
| **M3** | -10 | 409 | Hydrolysis/ Hydroxylation | 9.9 | 5.92E+07 | 2.61E+07 |
| **M4** | -26 | 393 | Hydrolysis/ Hydroxylation | 10.1 | 2.12E+07 | 3.80E+06 |
| **M5 -42a** | -42 | 377 | Hydrolysis | 11.4 | 5.36E+06 | 6.12E+06 |

In the study, five metabolites were detected in the urine and bile of SD rats dosed with **46**. These metabolites were named as M1 through to M5 based on their eluting time under HPLC conditions. Among the five metabolites, M1, M2 and M3 were identified as di-hydroxy **42a**; M4 was identified as hydroxylated **42a**; M5 was identified as active drug **42a**. Location of the hydroxyl groups was established through mass spectrometry fragmentation patterns (see supporting information). M3 to M5 were detected both in urine and bile samples, M1 and M2 were only detected in the bile sample.

The presence of the pro-drug in the rat urine indicates that that the pro-drug does not completely break down to its active metabolites as predicted. As the plasma levels obtained are a measure of parent drug only, they are not a true representation of the drug levels present. Studies are currently underway to establish if a more suitable pro-drug can be synthesised that will resolve the issue and provide a compound suitable for *in vivo* efficacy testing.

CONCLUSIONS

To conclude, a 3-6 step synthesis of a range of 2-mono aryl amine 3-methyl quinolones with potent anti-tuberculosis activity has been reported. Compounds have been developed that are metabolically stable and have a good pharmacokinetic and toxicological profile. Importantly, the lead compound **42a** demonstrates equipotent activity against all drug sensitive and multi-drug resistant strains of Mtb tested. Work continues to develop a suitable pro-drug to embark on *in vivo* efficacy studies.

EXPERIMENTAL SECTION

***Chemistry***

All reactions that employed moisture sensitive reagents were performed in dry solvent under an atmosphere of nitrogen in oven dried glassware. All reagents were purchased from Sigma Aldrich or Alfa Aesar chemical companies, and were used without purification.Thin layer chromatography (TLC) was carried out on Merck silica gel 60 F-254 plates and U.V. inactive compounds were visualised using iodine or anisaldehyde solution. Flash column chromatography was performed on ICN Ecochrom 60 (32-63 mesh) silica gel eluting with various solvent mixtures and using an air line to apply pressure. NMR spectra were recorded on a Brucker AMX 400 (1 H, 400 MHz; 13C, 100 MHz) spectrometer. Chemical shifts are described on parts per million (δ) downfield from an internal standard of trimethylsilane. Mass spectra were recorded on a VG analytical 7070E machine and Fisons TRIO spectrometers using electron ionisation (EI) and chemical ionisation (CI). The optical rotation of the products were determined on Perkin Elmer Polarimeter (Model: 343Plus), and data was collected and processed by Expert Read 1.00.02 software. All compounds were found to be >95% pure by HPLC unless specified below. See supporting information for experimental methods and data relating to all intermediates.

Purity determination was performed by HPLC analysis using Agilent 1200 solvent delivery system. The HPLC methods used the following conditions: Knauer Eurospher 100-5 C18(250 mm X 4.6 mm) at 25oC with 1.5 mL/min flow rate; Method A: 90% acetonitrile containing 0.05% trifluoroacetic acid and 10% water containing 0.05% trifluoroacetic acid; Method B: 80% methanol and 20% acetonitrile.

**General procedure for the preparation quinolones 1, 2, 7a-k, 17a-l, 21a-g, 24, 32a-g, 38a-j, 42a-b and 45a-g.** Trifluoromethanesulfonic acid (26 µL, 0.31 mmol, 0.2 eq) was added to oxazoline **4** (1.54 mmol) and the respective ketone (1.54 mmol, 1eq) in anhydrous n-butanol (10 mL). The mixture was heated to 130oC for 24 h (followed by tlc). The reaction was cooled and the solvent removed under reduced pressure. Sat. NaHCO3 (aq) was added and the resulting aqueous solution was extracted with ethyl acetate (x 3), the combined organic layers were washed with water and brine, dried over MgSO4, filtered and concentrated to a yellow solid. The crude product was triturated with diethyl ether to give the desired quinolone. In cases where trituration was not possible compounds were purified by flash column chromatography.

*Preparation of 3-methyl-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***1.*** Light yellow powder (Yield 23%); m.p 290-292 oC; 1H NMR (400MHz, CDCl3), δH 8.46 (s, 1H, NH), 8.35 (d, 1H, J = 8.1 Hz, Ar), 7.59-7.52 (m, 1H, Ar), 7.36 (d, 2H, J = 8.7 Hz, Ar), 7.30 (dd, 2H, J = 15.1 H, 7.2 Hz, Ar), 6.96 (d, 2H, J = 8.7 Hz, Ar), 2.10 (3H.CH3), 1.78-1.61 (m, 10H, CH2); 13C NMR (100MHz, CDCl3), δC 179.1, 152.9, 148.0, 139.4, 131.8, 129.9, 126.7, 125.5, 124.0, 123.5, 117.4, 116.5, 115.6, 50.0, 26.0, 13.0; MS (ES+), [M + H] + (100), 319.2, HRMS calculated for 319.1810 C21H23N2O, found 319.1808; Anal. C21H22N2O requires C 79.21%, H 6.96%, N 8.80%, found C 78.83%, H 6.85%, N 8.42%.

*Preparation of 7-methoxy-3-methyl-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***2****.* Orange powder (Yield 36%); m.p 278-280 oC; 1H NMR (400MHz, CDCl3), δH 10.09 (s, 1H, NH), 8.16 (d, 1H, J = 8.5 Hz, Ar), 7.39 (d, 2H, J 0 8.9 Hz, Ar), 7.10 (d, 2H, J = 8.9 Hz, Ar), 6.92 (dd, 2H, J = 8.5 Hz, 2.6 Hz, Ar), 3.89 (s, 3H, OCH3), 3.33-3.28 (m, 2H, CH2), 2.06 (s, 3H, CH3), 1.80-1.61 (m, 6H, CH2); 13C NMR (100MHz, CDCl3), δC 176.4, 161.8, 152.8, 129.5, 126.5, 124.7, 115.3, 114.7, 114.3, 97.7, 54.7, 25.3, 24.1, 11.4; MS (ES+), [M + H] + (100), 348.2, HRMS calculated for 348.1916 C22H25N3O, found 348.2002; Anal. C22H24N2O2 requires C 75.83%, H 6.94%, N 8.04%, found C 75.47%, H 6.83%, N 7.61%.

*Preparation of 6-fluoro-3-methyl-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***7a****.* Orange powder (Yield 26%); m.p 328-330 oC 1H NMR (400MHz, DMSO), δH 11.53 (s, 1H, NH), 7.71 (ddd, 1H, J = 13.9 Hz, 9.3 Hz, 3.9 Hz, Ar), 7.51 (ddd, 1H, J 9.1 Hz, 8.4 Hz, 3.0 Hz, Ar), 7.38 (d, 2H, J = 8.9 Hz, Ar), 7.07 (d, 2H, J = 8.9 Hz, Ar), 3.30-3.26 (m, 4H, CH2), 1.95 (s, 3H, CH3), 1.66-1.55 (m, 6H, CH2); 13C NMR (100MHz, DMSO), δC 176.2, 157.1, 152.2, 148.6, 136.6, 130.2, 124.3, 121.2, 120.4, 115.0, 113.9,109.1, 49.1, 25.3, 24.3, 12.8; MS (ES+), [M + H] + (100), 337.2, HRMS calculated for 337.1716 C21H22N2OF, found 337.1728; Anal. C21H21N2OF requires C 74.98%, H 6.29%, N 8.33%, found C 74.51%, H 6.07%, N 8.04%.

*Preparation of 6,7-dimethoxy-3-methyl-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***7b.*** Very pale yellow solid (Yield 28%); 1H NMR (400 MHz, DMSO) δH 11.24 (s, 1H, NH), 7.45 (s, 1H, Ar), 7.36 (d, *J* = 8.8 Hz, 2H, Ar), 7.16 – 6.98 (m, 3H, Ar), 3.83 (s, 3H, OCH3), 3.82 (s, 3H, OCH3), 3.29 – 3.25 (m, 4H, CH2), 1.93 (s, 3H, CH3), 1.69 – 1.53 (m, 6H, CH2); 13C NMR (101 MHz, DMSO) δC 175.90 (C=O), 152.89, 152.05, 146.82, 146.54, 135.51, 130.19, 124.73, 117.34, 114.98, 113.15, 104.50, 99.38, 55.86 (OCH3), 55.79 (OCH3), 49.20, 25.35, 24.32, 12.86 (CH3); HRMS (ESI) C23H27N2O3 [M+H]+ requires 379.2022, found 379.2012 (100%); Anal. C23H26N2O3 requires C 72.99%, H 6.92%, N 7.40%, found C 71.98%, H 6.96%, N 6.96%.

*Preparation of 6-chloro-7-methoxy-3-methyl-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***7c.***White solid (Yield 35%); m.p. >300oC. 1H NMR (400 MHz, DMSO) δH 11.42 (s, 1H, NH), 8.02 (s, 1H, Ar), 7.38 (d, *J* = 8.8 Hz, 2H, Ar), 7.21 (s, 1H, Ar), 7.07 (d, *J* = 8.9 Hz, 2H, Ar), 3.91 (s, 3H, OCH3), 3.31 – 3.22 (m, 4H, CH2), 1.93 (s, 3H, CH3), 1.71 – 1.52 (m, 6H, CH2); 13C NMR (101 MHz, DMSO) δC 175.63 (C=O), 156.74, 152.17, 148.16, 140.13, 130.21, 126.09, 124.18, 118.08, 117.91, 114.89, 114.25, 100.13, 56.59 (OCH3), 49.10, 25.32, 24.32, 12.70 (CH3); HRMS (ESI) C22H24N2O235Cl[M+H]+ requires 383.1526, found 383.1513 (100%), C22H24N2O237Cl[M+H]+ requires 385.1497, found 385.1501 (34%). Anal. C22H23N2O2Cl requires C 69.01%, H 6.05%, N 7.32%, found C 68.98%, H 6.04%, N 7.23%.

*Preparation of 6-fluoro-7-methoxy-3-methyl-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***7d****.* White solid (Yield 41%) 1H NMR (400 MHz, DMSO) δH 11.39 (s, 1H, NH), 7.71 (d, *J* = 11.9 Hz, 1H, Ar), 7.37 (d, *J* = 8.7 Hz, 2H, Ar), 7.24 (d, *J* = 7.5 Hz, 1H, Ar), 7.07 (d, *J* = 8.8 Hz, 2H, Ar), 3.90 (s, 3H, OCH3), 3.30 – 3.19 (m, 4H, CH2), 1.92 (s, 3H, CH3), 1.74 – 1.48 (m, 6H, CH2); 13C NMR (101 MHz, DMSO) δC 175.94 (C=O), 152.15, 151.00, 150.87, 150.35, 147.88, 137.55, 130.20, 124.30, 114.93, 113.57, 110.03, 101.12, 56.36 (OCH3), 49.13, 25.33, 24.32, 12.70 (CH3); HRMS (ESI) C22H24N2O2F [M+H]+ requires 367.1822, found 367.1818. Anal. C22H23N2O2F requires C 72.11%, H 6.33%, N 7.64%, found C 71.95%, H 6.45%, N 7.37%.

*Preparation of 5,7-dimethoxy-3-methyl-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***7e****.* White solid (Yield 32%); m.p. 264 – 265oC. 1H NMR (400 MHz, DMSO) δH 10.93 (s, 1H, NH), 7.33 (d, *J* = 8.7 Hz, 2H, Ar), 7.05 (d, *J* = 8.7 Hz, 2H, Ar), 6.64 (d, *J* = 2.2 Hz, 1H, Ar), 6.25 (d, *J* = 2.1 Hz, 1H, Ar), 3.78 (s, 3H, OCH3), 3.77 (s, 3H, OCH3), 3.32 – 3.11 (m, 4H, CH2), 1.82 (s, 3H, CH3), 1.70 – 1.48 (m, 6H, CH2); 13C NMR (101 MHz, DMSO) δC 176.49 (C=O), 161.75, 161.03, 152.02, 145.53, 143.94, 130.15, 124.47, 115.49, 114.98, 109.24, 94.23, 91.57, 55.97 (OCH3), 55.48 (OCH3), 49.22, 25.35, 24.32, 12.82 (CH3); HRMS (ESI) C23H27N2O2 [M+H]+ requires 379.2022, found 379.2007. Anal. C23H26N2O2 requires C 72.99%, H 6.92%, N 7.40%, found C 72.13%, H 6.88%, N 7.03%.

*Preparation of 5,7-difluoro-3-methyl-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***7f.*** Off white solid (0.25 g, 35 %); mp 305-306 °C;1H NMR (400 MHz, DMSO) δ 11.50 (bs, 1H), 7.37 (d, *J* = 8.8 Hz, 2H), 7.15 (d, *J* = 9.2 Hz, 1H), 7.08 (d, *J* = 8.9 Hz, 2H), 6.98 (t, *J* = 9.6 Hz, 1H), 3.30 (m, 4H), 1.88 (s, 3H), 1.61 (m, 6H); 13C NMR (100 MHz, CDCl3) δC 175.2, 152.1, 148.6, 130.2, 116.1, 114.9, 100.2, 49.2, 25.3, 24.3, 12.6; MS (ES+) *m/z* 355 (M + H)+ HRMS calculated for 355.1622 C21H21N2OF2, found 355.1625; Purity HPLC 95% (method A) Rt = 2.34 min.

*Preparation of 7-fluoro-3-methyl-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***7g.*** Off white solid (0.15 g, 35 % ); mp 343-345 °C;1H NMR (400 MHz, DMSO) dd, *J* = 9.0, 6.6 Hz, 1H), 7.38 (d, *J* = 8.8 Hz, 2H), 7.30 (dd, *J* = 10.5, 2.3 Hz, 1H), 7.10 (m, 1H), 7.05 (d, *J* = 8.8 Hz, 2H), 3.28 (m, 4H), 1.94 (s, 3H), 1.62 (m, 6H); 13C NMR (100 MHz, DMSO) C not soluble in DMSO; MS (ES+) *m/z* 337 (M + H)+ HRMS calculated for 337.1716 C21H22N2OF, found 337.1722; Purity HPLC 97% (Method B) Rt = 2.44 min.

*Preparation of 7-chloro-3-methyl-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***7h.***Off white solid (0.17 g, 37 %); mp 342-343 °C;1H NMR (400 MHz, DMSO) δ 8.08 (d, *J* = 8.7 Hz, 1H), 7.59 (s, 1H), 7.40 ( d, *J* = 8.8 Hz, 2H), 7.18 (dd, *J =* 8.7, 2.0 Hz, 1H), 7.04 (d, *J*  = 8.8 Hz, 2H), 3.08 (m, 4H), 1.95 (s, 3H), 1.61 (m, 6H); 13C NMR (100 MHz, CDCl3) δC not soluble in DMSO; MS (ES+) *m/z* 353 (M + H)+ HRMS calculated for 353.1425 C21H22N2O35Cl, found 353.1421; Purity HPLC 97% (Method A) Rt = 2.07 min.

*Preparation of 2-(4-(4-fluoropiperidin-1-yl)phenyl)-3-methylquinolin-4(1H)-one* ***7i.*** White solid (0.18 g, 36 %). 1H NMR (400 MHz, DMSO) 8.10 (d, *J* = 8.8 Hz, 1H), 7.57 (m, 2H), 7.40 (d, *J* = 8.8 Hz, 2H), 7.24 (dd, *J* = 7.2, 6.8 Hz, 1H), 7.11 (d, *J* = 8.8 Hz, 2H), 4.88 (d, *J* = 48.8 Hz, 1H), 3.24 (m, 4H), 2.03 (m, 2H), 1.95 (s, 3H), 1.80 (m, 2H); 13C NMR (100 MHz, DMSO) C 176.4, 150.7, 130.6, 130.0, 124.9, 123.3, 122.1, 119.0, 114.8, 113.8, 89.4, 87.8, 44.6, 44.5, 30.5, 30.3, 12.6; MS (ES+) *m/z* 337 (M + H)+ HRMS calculated for 337.1716 C21H22N2OF, found 337.1720; Purity HPLC 96% (Method A) Rt = 2.21 min.

*Preparation of 2-(4-(4-fluoropiperidin-1-yl)phenyl)-7-methoxy-3-methylquinolin-4(1H)-one* ***7j****.* Yellow solid (Yield 43%) 1H NMR (400 MHz, DMSO) δ 11.26 (s, 1H, NH), 8.00 (d, *J* = 8.9 Hz, 1H, Ar), 7.39 (d, *J* = 8.6 Hz, 2H, Ar), 7.12 (d, *J* = 8.6 Hz, 2H, Ar), 7.05 (d, *J* = 2.1 Hz, 1H, Ar), 6.88 (dd, *J* = 8.9, 2.2 Hz, 1H, Ar), 5.02 – 4.77 (m, 1H, CH), 3.82 (s, 3H, OCH3), 3.57 – 3.44 (m, 2H, CH2), 3.32 – 3.20 (m, 2H, CH2), 2.13 – 1.95 (m, 2H, CH­2), 1.91 (s, 3H, CH3), 1.86 – 1.71 (m, 2H, CH2); 13C NMR (101 MHz, DMSO) δ 176.78 (C=O), 161.89, 151.28, 147.80, 141.66, 130.39, 127.22, 125.12, 117.98, 115.22, 114.02, 113.14, 99.23, 89.01 (d, *J* = 169.4 Hz, C-F), 55.74, 44.87 (d, *J* = 6.8 Hz), 30.84 (d, *J* = 19.0 Hz), 12.78 (CH3); HRMS (ESI) C22H24N2O2F[M+H]+ requires 367.1822, found 367.1836. Anal. C22H23N2O2F requires C 72.11%, H 6.33%, N 7.64%, found C 71.32%, H 6.34%, N 7.46%.

*Preparation of 5,7-difluoro-2-(4-(4-fluoropiperidin-1-yl)phenyl)-3-methylquinolin-4(1H)-one* ***7k.*** White solid (29%); m.p > 320 °C. 1H NMR (400 MHz, DMSO) 11.51 (s, 1H), 7.40 (m, 2H), 7.15 (m, 3H), 7.00 (m, 1H), 3.49 (m, 2H), 3.24 (m, 2H), 2.0 (m, 2H), 1.89 (s, 3H), 1.75 (m, 2H); 13C NMR (100 MHz, DMSO) C not soluble in DMSO; MS (ES+) *m/z* 373 (M + H)+ HRMS calculated for 373.1519 C21H20N2OF3, found 373.7528; Purity HPLC 97% (Method A) Rt = 2.18 min.

**General procedure for the preparation of compounds 14a-c.** To a solution of ketone **13** (0.24 mmol) in anhydrous 1,4-dioxane (8 ml) was added ground sodium hydroxide (30 mg, 0.75 mmol, 3 equiv). The mixture was allowed to reflux at 110oC for 5 h. The solution was cooled to room temperature and acidified by addition of 2N hydrochloric acid. The solid was filtered and washed with water, followed by ethyl acetate and dried.

*Preparation of 2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***14a.*** White solid (0.25 g, 70 %). m.p. 350 °C; 1H NMR (400 MHz, DMSO) 11.42 (bs, 1H), 8.07 (d, *J* = 8.0 Hz, 1H), 7.76 (d, *J* = 8.3 Hz, 1H), 7.71 (d, *J* = 8.6 Hz, 2H), 7.64 (dd, *J* = 8.3, 7.0 Hz, 1H), 7.30 (dd, *J* = 8.3, 7.0 Hz, 1H), 7.07 (d, *J* = 8.8 Hz, 2H), 6.29 (s, 1H), 3.33 (m, 4H), 1.19 (m, 6H); 13C NMR (100 MHz, DMSO) C not soluble in DMSO; MS (ES+) *m/z* 305 (M + H)+ HRMS calculated for 305.1654 C20H21N2O, found 305.1662; Anal. C20H20N2O requires C 78.92%, H 6.62%, N 9.20%, found C 78.67%, H 6.55%, N 8.89%.

*Preparation of 7-methoxy-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***14b.*** White solid (0.065 g, 41 %). m.p. 350 °C; 1H NMR (400 MHz, DMSO) 11.30 (bs, 1H), 7.96 (d, *J* = 8.9 Hz, 1H), 7.69 (d, *J* = 8.7 Hz, 2H), 7.23 (d, *J* = 2.3 Hz, 1H), 7.07 (d, *J* = 8.9 Hz, 2H), 6.89 (dd, *J* = 8.0, 4.0 Hz, 1H), 6.21 (s, 1H), 3.86 (s, 3H), 3.34 (m, 4H), 1.60 (m, 6H); 13C NMR (100 MHz, DMSO) C not soluble in DMSO; MS (ES+) *m/z* 335 (M + H)+ HRMS calculated for 335.1760 C21H23N2O2, found 335.1761; Purity HPLC 96% (method A) Rt = 1.81 min.

*Preparation of 2-(4-(4-fluoropiperidin-1-yl)phenyl)-7-methoxyquinolin-4(1H)-one* ***14c****.* Yellow solid (Yield 68%). 1H NMR (400 MHz, DMSO) δH 13.70 (s, 1H, NH), 8.18 (d, J = 9.2 Hz, 1H, Ar), 7.88 (d, J = 9.0 Hz, 2H, Ar), 7.58 (d, J = 2.3 Hz, 1H, Ar), 7.34 (dd, J = 9.2, 2.4 Hz, 1H, Ar), 7.31 – 7.19 (m, 3H, Ar), 4.93 (dtt, J = 48.9, 7.0, 3.4 Hz, 1H, CH), 3.98 (s, 3H, OCH3), 3.71 – 3.58 (m, 2H, CH2), 3.51 – 3.37 (m, 2H, CH2), 2.10 – 1.88 (m, 2H, CH2), 1.87 – 1.68 (m, 2H, CH2); HRMS (ESI) C21H22N2O2F[M+H]+ requires 353.1665, found 353.1667; Anal. C21H21N2O2F requires C 71.57%, H 6.01%, N 7.95%, found C 71.12%, H 5.93%, N 7.71%.

**General procedure for the preparation of compounds 15a-d.** Quinolone **14** (0.33 mmol) was added to MeOH (20 mL), 2M NaOH (4 mL) and water (4 mL). Sodium dichloroisocyanurate (36 mgs, 0.17 mmol, 0.5 eq) was added at room temperature and the resultant light orange solution was allowed to stir overnight. The solvent was removed *in vacuo* and the residue was dissolved in EtOAc (100 mL), followed by washing with water (50 mL) and brine (50 mL). The crude product was purified by column chromatography (eluting with 100 % EtOAc) to afford the desired product.

*Preparation of 3-chloro-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***15a***. White solid (40 mgs, 40 % ); 1H NMR (400 MHz, DMSO)  12.01 (bs, 1H), 8.15 (d, *J* = 7.9 Hz, 1H), 7.69 (m 2H), 7.52 (d, *J* = 8.7 Hz, 2H), 7.38 (m, 1H), 7.09 (d, *J* = 8.8 Hz, 2H), 3.33 (m, 4H), 1.61 (m, 6H); 13C NMR (100 MHz, DMSO) C 171.7, 152.5, 148.7, 139.3, 132.2, 130.7, 125.4, 124.0, 123.8, 122.1, 118.9, 114.5, 113.2, 48.9, 25.3, 24.3; MS (ES+) *m/z* 339 (M + H)+ HRMS calculated for 339.1264 C20H20N2O35Cl, found 339.1252; Purity HPLC 98% (method A) Rt = 2.13 min.

*Preparation of 3-chloro-7-methoxy-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***15b.*** White solid (27 mgs, 61 % ); 1H NMR (400 MHz, DMSO)  11.82 (bs, 1H), 8.04 (d, *J* = 9.0 Hz, 1H), 7.52 (d, *J* = 8.8 Hz, 2H), 7.11 (m, 3H), 6.99 (dd, *J* = 9.2, 2.4 Hz, 1H), 3.85 (s, 3H), 3.33 (m, 4H), 1.61 (m, 6H); 13C NMR (100 MHz, DMSO) C 162.3, 148.1, 141.1, 130.6, 127.3, 118.2, 114.7, 114.2, 112.9, 99.7, 55.8, 49.1, 25.2, 24.2; MS (ES+) *m/z* 369 (M + H)+ HRMS calculated for 369.1370 C21H22N2O235Cl, found 369.1375; Purity HPLC 99% (method A) Rt = 1.83 min.

*Preparation of 3-chloro-2-(4-(4-fluoropiperidin-1-yl)phenyl)-7-methoxyquinolin-4(1H)-one* ***15c***. Yellow solid (Yield 52%). MP 304 – 306oC. 1H NMR (400 MHz, DMSO) δH 11.86 (s, 1H, NH), 8.03 (d, *J* = 9.0 Hz, 1H, Ar), 7.52 (d, *J* = 8.8 Hz, 2H, Ar), 7.14 (d, *J* = 8.9 Hz, 2H, Ar), 7.10 (d, *J* = 2.3 Hz, 1H, Ar), 6.98 (dd, *J* = 9.0, 2.4 Hz, 1H, Ar), 4.90 (dtt, *J* = 21.4, 7.3, 3.6 Hz, 1H, CH), 3.84 (s, 3H, OCH3), 3.63 – 3.46 (m, 2H), 3.34 – 3.19 (m, 2H), 2.13 – 1.90 (m, 2H), 1.85 – 1.58 (m, 2H); 13C NMR (101 MHz, DMSO) δC 171.31 (C=O), 162.33 (C-O), 151.61, 148.08, 141.06, 130.67, 127.30, 122.71, 118.20, 114.66, 114.20, 112.89, 99.63, 88.89 (d, *J* = 169.5 Hz, C-F), 55.81, 44.53 (d, *J* = 6.8 Hz), 30.69 (d, *J* = 19.1 Hz); HRMS (ESI) C21H21N2O2F35Cl[M+H]+ requires 387.1276, found 387.1287. Anal. C21H20N2O2FCl requires C 65.20%, H 5.21%, N 7.24%, found C 64.90%, H 5.35%, N 6.95%.

*Preparation of 3-chloro-2-(4-(4-fluoropiperidin-1-yl)phenyl)quinolin-4(1H)-one* ***15d***. Light yellow solid (0.19 g, 58 %). 1H NMR (400 MHz, DMSO) 12.05 (bs, 1H), 8.15 (d, *J* = 8.0 Hz, 1H), 7.70 (d, *J* = 4.0 Hz, 2H), 7.54 (d, *J* = 8.8 Hz, 2H), 7.38 (m, 1H), 7.15 (d, *J* = 8.8 Hz, 2H), 4.89 (d, *J* = 48.0 Hz, 1H), 3.53 (m, 2H), 3.30 (m, 2H), 1.97 (m, 2H), 1.79 (m, 2H); 13C NMR (100 MHz, DMSO) C 175.2, 150.8, 128.5, 127.9, 127.3, 124.6, 115.4, 103.9, 89.9, 88.2, 79.6, 66.7, 45.2, 45.1, 31.0, 30.8, 15.5; MS (ES+) *m/z* 357 (M + H)+ HRMS calculated for 357.1170 C20H19N2OF35Cl, found 357.1159; Purity HPLC 95% (Method A) Rt = 2.15 min.

**General procedure for the preparation of compounds 15e-f.** Quinolone **14** (0.33 mmol) was added to DCM (15 mL) and MeOH (4 mL). NBS (58 mgs, 0.33 mmol) was added at room temperature and the resultant bright yellow solution was allowed to stir overnight. The solvent was removed *in vacuo* and the residue was dissolved in EtOAc (100 mL), followed by washing with water (50 mL) and brine (50 mL). The crude product was purified by column chromatography (eluting with 70 % EtOAc in *n*-hexanes) to afford the desired product.

*Preparation of 3-bromo-2-(4-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***15e***. White solid (63 %); 1H NMR (400 MHz, DMSO)  12.07 (bs, 1H), 8.15 (d, *J* = 8.1 Hz, 1H), 7.68 (m 2H), 7.49 (d, *J* = 8.8 Hz, 2H), 7.39 (ddd, *J* = 8.6, 7.9, 4.1 Hz, 1H), 7.08 (d, *J* = 8.8 Hz, 2H), 3.31 (m, 4H), 1.62 (m, 6H); 13C NMR (100 MHz, DMSO) C 172.1, 152.5, 150.4, 139.4, 132.3, 130.6, 125.6, 124.2, 124.0, 123.2, 118.8, 114.5, 105.5, 49.0, 25.3, 24.4; MS (ES+) *m/z* 383 (M + H)+ HRMS calculated for 383.0759 C20H20N2O79Br, found 383.0748; Purity HPLC 98% (Method A) Rt = 1.75 min.

*Preparation of 3-bromo-2-(4-(4-fluoropiperidin-1-yl)phenyl)quinolin-4(1H)-one* ***15f***. Light yellow solid (0.20g, 55 %). 1H NMR (400 MHz, DMSO) 12.26 (bs, 1H), 8.15 (d, *J* = 8.8 Hz, 1H), 7.69 (m, 1H), 7.50 (d, *J* = 8.8 Hz, 2H), 7.39 (m, 2H), 7.14 (d, *J* = 8.8 Hz, 2H), 4.89 (d, *J* = 48.0 Hz, 1H), 3.53 (m, 2H), 3.30 (m, 2H), 1.97 (m, 2H), 1.79 (m, 2H) . 13C NMR (100 MHz, DMSO) C 179.7, 150.3, 132.3, 130.2, 125.6, 124.5, 114.6, 105.6, 89.7, 88.1, 44.6, 44.5, 30.8, 15.5; MS (ES+) *m/z* 401 (M + H)+ HRMS calculated for 401.0665 C20H19N2OF79Br, found 401.0656; Purity HPLC 99% (Method A) Rt = 2.15 min.

*Preparation of 5,7-difluoro-3-methyl-2-(4-(4-(trifluoromethyl)piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***17a.*** White solid (32%); m.p. >350 °C. 1H NMR (400 MHz, DMSO) 11.52 (s, 1H), 7.39 (m, 2H), 7.17 (m, 3H), 7.00 (m, 1H), 3.95 (m, 2H), 2.85 (m, 2H), 1.91 (m, 2H), 1.87 (s, 3H), 1.55 (m, 2H); 13C NMR (100 MHz, DMSO) C not soluble in DMSO; MS (ES+) *m/z* 423 (M + H)+ HRMS calculated for 423.1496 C22H20N2OF5, found 423.1483; Anal. C22H19N2OF5 requires C 62.56%, H 4.53%, N 6.63%, found C 62.49%, H 4.52%, N 6.62%.

*Preparation of 5,7-difluoro-3-methyl-2-(4-(4-methylpiperidin-1-yl)phenyl)quinolin-4(1H)-one* ***17b****.* White solid (54%); m.p. decomposed at 310°C. NMR: 1H (400 MHz, DMSO) δ 11.50 (s, 1H), 7.36 (d, J = 8.8 Hz, 2H), 7.16 (d, J = 10.0 Hz, 1H), 7.08 (d, J = 8.9 Hz, 2H), 7.00 (ddd, J = 12.0, 9.6, 2.4 Hz, 1H), 3.83 (d, J = 12.8 Hz, 2H), 2.76 (td, J = 12.5, 2.4 Hz, 2H), 1.87 (s, 3H), 1.70 (d, J = 12.7 Hz, 2H), 1.63 – 1.49 (m, 1H), 1.21 (qd, J = 12.7, 4.0 Hz, 2H), 0.94 (d, J = 6.5 Hz, 3H); 13C (101 MHz, DMSO) δ 175.37, 163.51, 160.76, 152.01, 147.65, 142.84, 130.21, 123.60, 116.33, 114.91, 110.49, 99.41, 98.81, 48.41, 33.55, 30.65, 22.18, 12.51. ES HRMS: m/z found 369.1792, C22H23N2OF2 requires 369.1778; Anal. C22H22N2OF2 requires C 71.72%, H 6.02%, N 7.60%, found C 71.66%, H 5.95%, N 7.52%.

*Preparation of 2-(4-(6-azaspiro[2.5]octan-6-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one* ***17c.*** White solid (Yield 34%); m.p. > 300oC. 1H NMR (400 MHz, DMSO) δH 11.51 (s, 1H, NH), 7.38 (d, *J* = 8.7 Hz, 2H), 7.16 (d, *J* = 10.0 Hz, 1H), 7.11 (d, *J* = 8.8 Hz, 2H), 7.00 (ddd, *J* = 11.9, 9.6, 2.3 Hz, 1H), 3.38 – 3.35 (m, 4H), 1.88 (s, 3H, CH3), 1.53 – 1.36 (m, 4H), 0.35 (s, 4H); 13C NMR (101 MHz, DMSO) δ 175.52, 163.75 (d, *J* = 61.6 Hz), 161.38 (d, *J* = 77.0 Hz), 152.14, 147.75, 142.80 (dd, *J* = 14.7, 6.3 Hz), 130.34, 123.72, 116.45, 115.22, 110.59 (d, *J* = 2.4 Hz), 99.60 (dd, *J* = 24.9, 4.1 Hz), 98.95 (dd, *J* = 28.7, 25.6 Hz), 48.40, 34.32, 18.15, 12.61, 11.59. HRMS (ESI) C23H22N2OF23Na[M+Na]+ requires 403.1598, found 403.1612. Anal. C23H22N2OF requires C 72.61%, H 5.83%, N 7.36%, found C 72.41%, H 5.91%, N 7.31%.

*Preparation of 2-(4-(4,4-difluoropiperidin-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one* ***17d.*** White solid (0.30 g, 57 %). 1H NMR (400 MHz, DMSO) 7.38 (d, *J* = 8.8 Hz, 2H), 7.10 (d, *J* = 8.8 Hz, 2H), 7.07 (m, 1H), 6.82 (dd, *J* = 11.0, 10.6 Hz, 1H), 3.43 (m, 4H), 2.07 (m, 4H), 1.88 (s, 3H); 13C NMR (100 MHz, DMSO) δC 174.2, 149.5, 129.9, 122.8, 118.5, 115.3, 115.0, 45.3, 33.0, 32.8, 32.5, 12.6; MS (CI+) *m/z* 391 (M + H)+ HRMS calculated for 391.1428 C21H19N2OF4, found 391.1430; Purity HPLC 95% (Method A) Rt = 2.39 min.

*Preparation of 5,7-difluoro-2-(4-(3-fluoropiperidin-1-yl)phenyl)-3-methylquinolin-4(1H)-one* ***17e.*** Light brown solid (0.12 g, 27 %). 1H NMR (400 MHz, DMSO) 11.49 (bs, 1H), 7.38 (d, *J* = 8.8 Hz, 2H), 7.10 (d, *J* = 8.8 Hz, 2H), 7.07 (m, 1H), 6.99 (dd, *J* = 11.0, 10.6 Hz, 1H),4.82 (d, *J* = 48.8 Hz, 1H), 3.50-3.33 (m, 4H), 1.87 (s, 3H), 1.86-1.62 (m, 4H) ; 13C NMR (100 MHz, DMSO) C 175.2, 151.4, 147.1, 129.8, 123.6, 116.0, 114.6, 98.8, 88.1, 86.4, 51.8, 51.6, 47.3, 29.3, 29.1, 20.6, 20.5, 12.1; MS (EI+) *m/z* 373 (M + H)+ HRMS calculated for 373.1528 C21H20N2OF3, found 373.1524; Purity HPLC 97% (Method A) Rt = 2.42 min.

*Preparation of 5,7-difluoro-3-methyl-2-(4-(3-methylpiperidin-1-yl)phenyl)quinolin-4(1H)-one* ***17f.***White solid (45%). Melting point: 280~282°C. NMR: 1H (400 MHz, DMSO) δ 11.50 (s, 1H), 7.36 (d, J = 8.8 Hz, 2H), 7.16 (d, J = 9.0 Hz, 1H), 7.07 (d, J = 8.9 Hz, 2H), 7.00 (ddd, J = 12.0, 9.6, 2.4 Hz, 1H), 3.77 (t, J = 11.6 Hz, 2H), 2.72 (td, J = 12.3, 2.9 Hz, 1H), 2.42 (dd, J = 12.4, 10.7 Hz, 1H), 1.87 (s, 3H), 1.82 – 1.48 (m, 4H), 1.09 (ddd, J = 23.5, 12.4, 3.9 Hz, 1H), 0.93 (d, J = 6.6 Hz, 3H).13C (101 MHz, DMSO) δ 175.37, 164.10, 161.50, 152.00, 147.66, 142.69, 130.22, 123.46, 116.33, 114.81, 110.59, 99.40, 98.79, 55.93, 48.45, 32.93, 30.35, 24.72, 19.58, 12.50. ES HRMS: m/z found 369.1772, C22H23N2OF2 requires 369.1778; Anal. C22H22N2OF2 requires C 71.72%, H 6.02%, N 7.60%, found C 71.76%, H 5.94%, N 7.58%.

*Preparation of (R)-5,7-difluoro-3-methyl-2-(4-(3-methylpiperidin-1-yl)phenyl)quinolin-4(1H)-one* ***17g****.* White solid (43%). 1H and 13C NMR data is the same as the racemic analogue; ES HRMS: m/z found 369.1775, C22H23N2OF2 requires 369.1778; Anal. C22H22N2OF2 requires C 71.72%, H 6.02%, N 7.60%, found C 71.68%, H 6.06%, N 7.53%; the optical rotation was measured as [α]D22=+81.5°±0.9 (c=0.558g/100ml in MeOH).

*Preparation of (S)-5,7-difluoro-3-methyl-2-(4-(3-methylpiperidin-1-yl)phenyl)quinolin-4(1H)-one* ***17h.***White solid (40%). 1H and 13C NMR data is the same as the racemic analogue; ES HRMS: m/z found 369.1782, C22H23N2OF2 requires 369.1778; Anal. C22H22N2OF2 requires C 71.72%, H 6.02%, N 7.60%, found C 71.77%, H 6.0%, N 7.64%; the optical rotation was measured as [α]D22=-86.1°±0.7 (c=0.588g/100ml in MeOH).

*Preparation of 5,7-difluoro-3-methyl-2-(4-(4-methylpiperazin-1-yl)phenyl)quinolin-4(1H)-one* ***17i.*** White solid (39%); m.p. >350 °C. 1H NMR (400 MHz, DMSO) 11.50 (s, 1H), 7.89 (d, *J* = 9.0, 2H), 7.19 (m, 1H), 7.05 (m, 1H), 6.85 (d, *J* = 9.0, 2H), 3.35 (m, 4H), 2.55 (m, 4H), 2.31 (s, 3H), 1.90 (s, 3H); 13C NMR (100 MHz, DMSO) C not soluble in DMSO; MS (ES+) *m/z* 370 (M + H)+ HRMS calculated for 370.1717 C21H22N3OF2, found 370.1731; Purity HPLC 99% (Method A) Rt = 1.59 min.

*Preparation of 2-(4-(azepan-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one* ***17j.*** White solid (41%); m.p. >350 °C. 1H NMR (400 MHz, DMSO) 11.45 (s, 1H), 7.31 (d, *J* = 8.8, 2H), 7.19 (m, 1H), 7.00 (m, 1H), 6.85 (d, *J* = 8.9, 2H), 3.55 (m, 4H), 1.92 (s, 3H), 1.75 (bs, 4H), 1.45 (bs, 4H); 13C NMR (100 MHz, DMSO) C 175.4, 149.4, 147.8, 130.5, 120.5, 116.1, 110.8, 99.54, 98.9, 98.7, 49.1, 48.1, 47.9, 47.7, 47.5, 27.0, 26.6, 12.6; MS (ES+) *m/z* 369 (M + H)+ HRMS calculated for 369.1764 C22H23N2OF2, found 369.1778; Anal. C22H22N2OF2 requires C 71.72%, H 6.02%, N 7.60%, found C 71.36%, H 5.97%, N 7.39%.

*Preparation of* 2-(4-(benzylamino)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one ***17k.*** White solid (51%); m.p. 282-283°C.NMR: 1H (400 MHz, DMSO) δ 11.39 (s, 1H), 7.36 (dt, J = 15.1, 7.4 Hz, 4H), 7.27 – 7.21 (m, 3H), 7.13 (d, J = 9.0 Hz, 1H), 6.97 (ddd, J = 12.0, 9.8, 2.3 Hz, 1H), 6.84 (t, J = 6.1 Hz, 1H), 6.72 (d, J = 8.6 Hz, 2H), 4.36 (d, J = 6.1 Hz, 2H), 1.86 (s, 3H); 13C (101 MHz, DMSO) δ 175.36, 164.04, 161.44, 150.01, 148.03, 142.58, 141.46, 140.25, 130.20, 128.73, 127.47, 127.09, 121.61, 116.08, 112.05, 99.34, 98.71, 46.39, 12.56. ES HRMS: m/z found 377.1465, C23H19N2OF2 requires 377.1465; Anal. C23H18N2OF2 requires C 73.39%, H 4.82%, N 7.44%, found C 73.18%, H 4.74%, N 7.41%.

*Preparation of* 2-(4-(dimethylamino)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one ***17l.*** White solid (46%); m.p. 294°C. NMR: 1H (400 MHz, DMSO) δ 11.48 (s, 1H), 7.37 (d, J = 8.8 Hz, 2H), 7.17 (d, J = 10.1 Hz, 1H), 6.99 (ddd, J = 12.1, 9.6, 2.4 Hz, 1H), 6.86 (d, J = 8.9 Hz, 2H), 2.99 (s, 6H), 1.88 (s, 3H); 13C (101 MHz, DMSO) δ 175.39, 164.08, 160.75, 151.31, 147.89, 142.77, 130.17, 121.70, 116.20, 111.89, 110.45, 99.38, 98.77, 40.24, 12.55. ES HRMS: m/z found 315.1319, C18H17N2OF2 requires 315.1309; Anal. C18H16N2OF2 requires C 68.78%, H 5.13%, N 8.91%, found C 68.47%, H 5.14%, N 8.78%.

*Preparation of* 2-(4-(4-benzylpiperidin-1-yl)phenyl)-3-methylquinolin-4(1*H*)-one ***21a.*** White powder (Yield 33%); m.p 256-258 oC 1H NMR (400MHz, DMSO), δH 11.39 (s, 1H, NH), 8.10 (d, 1H, J = 7.7 Hz, Ar), 7.63-7.55 (m, 2H, AR), 7.37 (d, 2H, J = 8.9 Hz, Ar), 7.33-7.24 (m, 3H, Ar), 7.22-7.17 (m, 3H, Ar), 706 (d, 2H, J = 8.9 Hz, Ar), 3.82 (d, 2H, J = 12.8 Hz, CH2), 2.79-2.66 (m, 2H, CH2), 2.56 (d, 2H, J = 7.0 Hz, CH2Ar), 1.96 (s, 3H, CH3), 1.79-1.73 (m, 1H, CH), 1.67 (d, 2H, J = 12.9 Hz, CH2), 1.29 (qd, 2H, J = 12.6 Hz, 3.9 Hz, CH2) 13C NMR (100MHz, DMSO), δC 177.0, 151.9, 148.3, 140.5, 139.9, 131.3, 130.2, 129.4, 128.5, 126.2, 125.3, 124.5, 123.3, 122.7, 118.4, 115.0, 114.4, 48.7, 42.6, 37.7, 31.5, 12.8 MS (ES+), [M + H] + (100), 409.2, HRMS calculated for 409.2280 C28H29N2O, found 409.2289; Anal. C28H28N2O requires C 82.32%, H 6.91%, N 6.86%, found C 81.98%, H 6.92%, N 6.88%.

*Preparation of* 2-(4-(4-benzylpiperidin-1-yl)phenyl)-6-fluoro-3-methylquinolin-4(1*H*)-one ***21b.*** White powder (Yield 40%); m.p. 302-302 oC 1H NMR (400MHz, DMSO), δH 11.55 (s,1H, NH), 7.73 (dd, 1H, J = 9.5 Hz, 3.0 Hz, Ar), 7.68 (dd, 1H, J = 9.1 Hz, 4.7 Hz, Ar), 7.54-7.48 (m, 1H, Ar), 7.38 (d, 2H, J = 8.9 Hz, Ar), 7.33-7.27 (m, 2H, Ar), 7.23-7.17 (m, 3H, Ar), 7.06 (d, 2H, J = 8.9 Hz, Ar), 3.83 (d, 2H, J = 12.7 Hz, CH2), 2.79-2.67 (m, 2H, CH2), 2.55 (d, 2H, J = 7.0 Hz, CH2Ar), 1.94 (s, 3H, CH3), 1.80-1.68 (m, 1H, CH), 1.67 (d, 2H, J = 13.1 Hz, CH2), 1.28 (qd, 2H, J = 12.6 Hz, 3.9 Hz, CH2) 13C NMR (100MHz, DMSO), δC 176.2, 157.1, 152.0, 148.6, 140.5, 136.6, 130.2, 129.4, 128.5, 126.2, 124.3, 121.2, 120.4, 115.0, 113.9, 109.1, 48.4, 42.6, 37.7, 31.4, 12.7 MS (ES+), [M + H]+ (100), 427.2, HRMS calculated for 427.2186 C28H28N2O4F, found 427.2177; Anal. C28H27N2OF requires C 78.85%, H 6.38%, N 6.57%, found C 78.31%, H 6.35%, N 6.63%.

*Preparation of 2-(4-(4-benzylpiperidin-1-yl)phenyl)-7-methoxy-3-methylquinolin-4(1H)-one* ***21c.*** Light yellow powder (Yield 42 %); m.p. 218-220 oC 1H NMR (400MHz, DMSO), δH 11.21 (s, s, 1H, NH), 7.99 (d, 1H, J = 8.9 Hz, Ar), 7.36 (d, 2H, J = 8.7 Hz, Ar), 7.29 (d, 2H, J = 7.2 Hz, Ar), 7.20 (d, 3H, J = 6.4 Hz, Ar), 7.05 (d, 3H, J = 8.6 Hz, Ar), 6.87 (dd, 1H, J = 8.9 Hz, 2.4 Hz, Ar), 3.82 (s, 3H, OCH3), 2.71 (t, 2H, J = 11.5 Hz, CH2), 2.56 (d, 2H, J = 6.9 Hz, CH2Ar), 1.91 (s, 3H, CH3), 1.79-1.71 (m, 1H, CH), 1.29 (dt, 2H, J = 11.7 Hz, 8.9 Hz, CH2) 13C NMR (100MHz, DMSO), δC 176.7, 161.8, 151.8, 147.8,141.6, 140.5, 130.2, 129.4, 128.5, 127.1, 126.2, 124.6, 117.9, 115.0, 113.9, 113.0, 99.2, 55.6, 48.5, 42.6, 37.7, 31.5, 12.7 MS (ES+), [M + H] + (100), 439.2 HRMS calculated for 439.2386 C29H31N2O2, found 439.2386; Purity HPLC 95% (Method B) Rt = 2.43 min.

*Preparation of 2-(4-(4-benzylpiperazin-1-yl)phenyl)-3-methylquinolin-4(1H)-one* ***21d.*** White powder (Yield 30%); m.p. 258-260 oC 1H NMR (400MHz, DMSO), δH 11.40 (s, 1H, NH), 8.10 (d, 1H, J = 7.7 Hz, AR), 7.63-7.55 (m, 2H, Ar), 7.40 (d, 2H, J = 8.9 Hz, Ar), 7.37-7.33 (m, 3H, Ar), 7.27 (ddd, 2H, J = 10.3 Hz, 5.5 Hz, 2.5 Hz, Ar), 7.08 (d, 2H, J = 8.9 Hz, Ar), 3.54 (s, 2H, CH2Ar), 3.29-3.23 (m, 4H, NCH2), 2.58-2.52 (m, 4H, CH2N), 1.93 (s, 3H, CH3) 13C NMR (100MHz, DMSO), δC 177.0, 151.9, 148.2, 139.9, 138.4, 131.4, 130.2, 129.3, 128.6, 127.4, 125.3, 123.3, 122.8, 118.4, 114.9, 114.4, 62.4, 52.8, 49.0, 48.1, 12.7 MS (ES+), [M + H] + (100), 410.2, HRMS calculated for 410.2232 C27H28N3O, found 410.2234; Anal. C27H27N3O requires C 79.19%, H 6.65%, N 10.26%, found C 78.63%, H 6.66%, N 10.21%.

*Preparation of 2-(4-(4-benzylpiperazin-1-yl)phenyl)-6-fluoro-3-methylquinolin-4(1H)-one* ***21e.*** White powder (Yield 28%); m.p. 306-308 oC. 1H NMR (400MHz, DMSO), δH 11.69 (s, 1H, NH), 7.74-7.71 (m, 2H, Ar), 7.54-7.48 (m, 1H, Ar), 7.40 (d, 2H, J = 8.9 Hz, Ar), 7.37-7.33 (m, 4H, Ar), 7.08 (d, 2H, J = 8.9 Hz, Ar), 3.54 (s, 2H, CH2Ar), 3.30-3.22 (m, 4H, CH2N), 2.59-2.52 (m, 4H, NCH2), 1.94 (s, 3H, CH3) 13C NMR (100MHz, DMSO), δC 151.9, 148.6, 138.4, 136.7, 130.2, 129.3, 128.6, 127.4, 124.9, 124.2, 120.4, 114.8, 113.9, 109.0, 62.4, 55.3, 52.8, 48.0, 12.8 MS (ES+), [M + H] + (100), 428.2, HRMS calculated for 428.2138 C27H27N3OF, found 428.2138; Purity HPLC 98% (Method A) Rt = 1.82 min..

*Preparation of 7-methoxy-3-methyl-2-(4-(4-phenylpiperazin-1-yl)phenyl)quinolin-4(1H)-one* ***21f.*** White powder (Yield 30 %); m.p. 312-314 oC. 1H NMR (400MHz, DMSO), δH 11.25 (s, 1H, NH), 8.01 (d, 1H, J = 9.0 Hz, Ar), 7.43 (d, 2H, J = 8.8 Hz, Ar), 7.26 (dd, 2H, J = 8.4 Hz, Ar), 7.16 (d, 2H, J = 8.8 Hz, Ar), 7.05 (d, 1H, J = 2.4 Hz, Ar), 7.02 (d, 2H. J = 8.0 Hz, Ar), 6.88 (dd, 1H, J = 9.0 Hz, Ar), 6.83 (t, 1H, J = 7.3 Hz, Ar), 3.82 (s, 3H, OCH3), 3.41 (dd, 4H, J = 6.5 Hz, 3.5 Hz, NCH2), 3.31 (dd, 4H, J = 6.5 Hz, 3.5 Hz, CH2N), 1.92 (s, 3H, CH3) 13C NMR (100MHz, DMSO), δC 176.7, 161.8, 151.7, 151.3, 147.7, 141.6, 130.2, 129.4, 127.1, 125.6, 119.6, 117.9, 116.1, 115.1, 114.0, 113.0, 99.2, 55.7, 48.6, 48.1, 12.6 MS (ES+), [M + H] + (100), 426.2, HRMS calculated for 426.2182 C27H28N3O2, found 426.2184; Purity HPLC 91% (Method A) Rt = 1.80 min.

*Preparation of 2-(4-(4-benzylpiperazin-1-yl)phenyl)-7-methoxy-3-methylquinolin-4(1H)-one* ***21g.*** White powder (Yield 38%); m.p. 280-282 oC. 1H NMR (400MHz, DMSO), δH 11.22 (s, 1H, NH), 8.00 (d, 1H, J = 9.0 Hz, AR), 7.38 (d, 2H, J = 8.9 Hz, Ar), 7.37-7.33 (m, 4H, Ar), 7.31-7.24 (m, 1H, Ar), 7.07 (d, 2H, J = 8.9 Hz, Ar), 7.04 (d, 1H, J = 2.4 Hz, Ar), 6.87 (dd, 1H, J = 8.9 Hz, 2.4 Hz, Ar), 3.82 (s, 3H, OCH3), 3.54 (s, 2H, NCH2Ar), 3.29-3.23 (m, 4H, NCH2), 2.57-2.52 (m, 4H, CH2N), 1.91 (s, 3H, CH3) 13C NMR (100MHz, DMSO), δC 176.7, 161.8, 151.8, 147.7, 141.6, 138.4, 130.1, 129.3, 128.6, 127.4, 125.3, 117.9, 114.8, 113.0, 99.2, 62.4, 55.6, 52.8, 49.0, 48.0, 12.6 MS (ES+), [M + H] + (100), 440.2, HRMS calculated for 440.2338 C28H30N3O2, found 440.2344; Anal. C28H29N3O2 requires C 76.51%, H 6.65%, N 9.56%, found C 76.12%, H 6.63%, N 9.48%.

*Preparation of 5,7-difluoro-3-methyl-2-(3-(piperidin-1-yl)phenyl)quinolin-4(1H)-one* ***24.*** White solid (Yield 45%); m.p. 269 – 270oC. 1H NMR (400 MHz, DMSO) δH 11.63 (s, 1H, NH), 7.37 (t, *J* = 7.9 Hz, 1H, Ar), 7.16 (d, *J* = 9.8 Hz, 1H, Ar), 7.10 (dd, *J* = 8.4, 2.3 Hz, 1H, Ar), 7.06 – 6.97 (m, 2H, Ar), 6.86 (d, *J* = 7.5 Hz, 1H, Ar), 3.27 – 3.19 (m, 4H, CH2), 1.83 (s, 3H, CH3), 1.62 (d, *J* = 4.0 Hz, 4H, CH2), 1.59 – 1.50 (m, 2H, CH2); 13C NMR (101 MHz, DMSO) δ 175.48, 163.86 (dd, *J* = 65.8, 15.2 Hz), 161.33 (dd, *J* = 80.6, 14.7 Hz), 151.93, 148.14, 142.72 (dd, *J* = 14.7, 6.4 Hz), 135.62, 129.68, 118.86, 116.90, 116.66, 116.07, 110.74 (d, *J* = 10.7 Hz), 99.66 (dd, *J* = 24.4, 4.5 Hz), 99.06 (dd, *J* = 26.8, 25.8 Hz), 49.65, 25.57, 24.33, 12.44. HRMS (ESI) C21H20N2OF223Na [M+H]+ requires 377.1441, found 377.1448 (100%). Anal. C21H20N2OF2 requires C 71.17%, H 5.69%, N 7.90%, found C 70.78%, H 5.59%, N 7.64%.

*Preparation of 1-(4-(5,7-difluoro-3-methyl-4-oxo-1,4-dihydroquinolin-2-yl)phenyl)-1H-pyrrole-2-carbonitrile* ***32a.*** White solid (55%); m.p. 312°C. NMR: 1H (400 MHz, DMSO) δ 11.81 (s, 1H), 7.84 – 7.74 (m, 4H), 7.67 (dd, J = 2.8, 1.6 Hz, 1H), 7.31 (dd, J = 4.0, 1.6 Hz, 1H), 7.15 (d, J = 10.0 Hz, 1H), 7.06 (ddd, J = 12.0, 9.6, 2.4 Hz, 1H), 6.52 (dd, J = 3.9, 2.8 Hz, 1H), 1.86 (s, 3H); 13C (101 MHz, DMSO) δ 175.33, 161.19, 146.24, 142.77, 138.87, 134.57, 130.86, 128.96, 128.22, 124.59, 123.61, 117.07, 114.20, 111.64, 110.81, 103.09, 99.47, 99.21, 12.25; HRMS (ESI) C21H14N3OF2 [M+H]+ requires 362.1099, found 362.1108 (100%). Anal. C21H13N3OF2 requires C 69.80%, H 3.63%, N 11.63%, found C 69.67%, H 3.66%, N 11.38%.

*Preparation of 2-(4-(1H-indol-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one* ***32b*.** while solid (57%); m.p. decomposed at 325°C. NMR: 1H (400 MHz, DMSO) δ 11.79 (s, 1H), 7.84 (d, J = 8.5 Hz, 2H), 7.81 – 7.74 (m, 3H), 7.69 (t, J = 8.6 Hz, 2H), 7.26 (t, J = 7.7 Hz, 1H), 7.22 – 7.14 (m, 2H), 7.06 (ddd, J = 12.0, 9.7, 2.4 Hz, 1H), 6.78 (d, J = 3.3 Hz, 1H), 1.91 (s, 3H); 13C (101 MHz, DMSO) δ 175.38, 146.64, 142.71, 140.41, 137.96, 135.25, 132.43, 130.93, 129.74, 128.77, 123.87, 122.96, 121.51, 120.98, 117.40, 117.01, 110.75, 104.64, 99.47, 99.13, 96.43, 12.35; HRMS (ESI) C24H17N2OF2 [M+H]+ requires 387.1303, found 387.1300 (100%). Anal. C24H16N2OF2 requires C 74.60%, H 4.17%, N 7.25%, found C 74.21%, H 4.17%, N 7.24%.

*Preparation of 2-(4-(1H-pyrazol-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one* ***32c***. White solid (Yield 35%); m.p. 306oC. 1H NMR (400 MHz, DMSO) δH 11.73 (s, 1H, NH), 8.66 (d, *J* = 2.5 Hz, 1H), 8.07 (d, *J* = 8.6 Hz, 2H), 7.83 (d, *J* = 1.6 Hz, 1H), 7.70 (d, *J* = 8.5 Hz, 2H), 7.16 (d, *J* = 9.8 Hz, 1H), 7.05 (ddd, *J* = 11.9, 9.7, 2.3 Hz, 1H), 6.74 – 6.53 (m, 1H), 1.87 (s, 3H, CH3); 13C NMR (101 MHz, DMSO) δ 175.47 (C=O), 163.94 (dd, *J* = 72.7, 14.9 Hz, C-F), 161.40 (dd, *J* = 87.4, 15.3 Hz, C-F), 146.70, 142.81 (dd, *J* = 14.6, 6.2 Hz), 142.04, 140.83, 132.36, 130.81, 128.55, 118.62, 117.04, 110.80 (d, *J* = 8.8 Hz), 108.85, 99.70 (dd, *J* = 24.4, 4.5 Hz), 99.09 (d, *J* = 25.2 Hz), 12.40 (CH3); HRMS (ESI) C19H13N3OF223Na[M+Na]+ requires 360.0924, found 360.0935. Anal. C19H13N3OF2 requires C 67.65%, H 3.88%, N 12.46%, found C 67.26%, H 4.00%, N 12.24%.

*Preparation of 2-(4-(1H-pyrrol-1-yl)phenyl)-5-fluoro-3-methylquinolin-4(1H)-one* ***32d***. White solid (Yield 32%); m.p. >300oC. 1H NMR (400 MHz, DMSO) δH 11.66 (s, 1H, NH), 7.96 – 7.73 (m, 2H), 7.67 – 7.61 (m, 2H), 7.61 – 7.49 (m, 3H), 7.42 (d, *J* = 8.4 Hz, 1H), 6.97 (dd, *J* = 12.1, 7.9 Hz, 1H), 6.49 – 6.14 (m, 2H), 1.88 (s, 3H, CH3); 13C NMR (101 MHz, DMSO) δC 175.85 (C=O), 162.15, 159.57, 146.57, 142.23, 142.21 (d, *J* = 4.4 Hz), 140.89, 132.06 (d, *J* = 10.8 Hz), 131.60, 130.84, 119.35 (d, *J* = 12.6 Hz), 116.59, 114.53, 113.36 (d, *J* = 8.8 Hz), 111.37, 108.68 (d, *J* = 20.9 Hz), 12.46 (CH3). HRMS (ESI) C20H15N2OF23Na[M+Na]+ requires 341.1066, found 341.1080. Anal. C20H15N2OF requires C 75.46%, H 4.75%, N 8.80%, found C 75.23%, H 4.70%, N 8.72%.

*Preparation of 2-(4-(3,4-difluoro-1H-pyrrol-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one* ***32e***. White solid (38 mgs, 30 %). 1H NMR (400 MHz, DMSO) 11.78 (bs, 1H), 7.83 (d, *J* = 8.8 Hz, 2H), 7.72 (d, *J* = 8.8 Hz, 4H), 7.14 (d, *J* = 9.6 Hz, 1H), 7.11 (dd, *J* = 11.0, 10.6 Hz, 1H), 1.91 (s, 3H), 13C NMR (100 MHz, DMSO) C 175.1, 146.8, 140.3, 131.8, 130.8, 118.7, 116.9, 103.0, 12.3; MS (ES+) *m/z* 373 (M + H)+ HRMS calculated for 373.0964 C20H13N2OF4, found 373.0965; Purity HPLC 98% (Method A) Rt = 2.29 min.

*Preparation of 2-(3-chloro-4-(1H-pyrrol-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one* ***32f***. White solid (Yield 39%); m.p. 297 – 298oC. 1H NMR (400 MHz, DMSO) δH 11.79 (s, 1H, NH), 7.92 (s, 1H), 7.75 – 7.57 (m, 2H), 7.19 – 7.01 (m, 4H), 6.32 (t, *J* = 2.1 Hz, 2H), 1.87 (s, 3H, CH3); 13C NMR (101 MHz, DMSO) δ 175.42, 164.30, 161.92 (d, *J* = 14.8 Hz), 160.97 (d, *J* = 15.3 Hz), 145.27, 142.84, 139.25, 134.90, 131.52, 129.60, 128.42, 128.35, 122.63, 117.30, 110.23, 99.72 (d, *J* = 19.1 Hz), 99.24 (d, *J* = 26.0 Hz), 12.30; HRMS (ESI) C20H13N2OF235Cl23Na[M+Na]+ requires 393.0582, found 393.0592. Anal. C20H13N2OF requires C 64.79%, H 3.53%, N 7.56%, found C 64.66%, H3.69%, N 7.39%.

*Preparation of 5,7-difluoro-2-(2-fluoro-4-(1H-pyrrol-1-yl)phenyl)-3-methylquinolin-4(1H)-one* ***32g***. White solid (Yield 39%); m.p. 307oC. 1H NMR (400 MHz, DMSO) δH 11.82 (s, 1H, NH), 7.84 (dd, *J* = 11.8, 1.9 Hz, 1H), 7.77 – 7.64 (m, 2H), 7.62 – 7.55 (m, 2H), 7.18 – 6.99 (m, 2H), 6.40 – 6.23 (m, 2H), 1.79 (s, 3H, CH3); HRMS (ESI) C20H14N2OF3 [M+H]+ requires 355.1058, found 355.1074. Anal. C20H13N2OF3 requires C 67.79%, H 3.70%, N 7.91%, found C 66.94%, H 3.68%, N 7.73%.

*Preparation of (R)-5,7-difluoro-2-(4-(3-fluoropyrrolidin-1-yl)phenyl)-3-methylquinolin-4(1H)-one* ***38a****.* White solid (45%); m.p. 313-314°C. NMR: 1H (400 MHz, DMSO) δ 11.47 (s, 1H), 7.38 (d, J = 8.7 Hz, 2H), 7.18 (d, J = 9.3 Hz, 1H), 6.99 (ddd, J = 12.0, 9.6, 2.4 Hz, 1H), 6.73 (d, J = 8.7 Hz, 2H), 5.50 (d, J = 54.1 Hz, 1H), 3.71 – 3.36 (m, 4H), 2.38 – 2.12 (m, 2H), 1.89 (s, 3H); 13C (101 MHz, DMSO) δ 175.38, 148.31, 147.92, 142.70, 130.35, 121.60, 116.19, 111.70, 110.56, 99.39, 98.76, 94.49, 92.78, 54.48, 45.59, 32.14, 31.93, 12.58. ES HRMS: m/z found 359.1385, C20H18N2OF3 requires 359.1371; Anal. C20H17N2OF3 requires C 67.03%, H 4.78%, N 7.82%, found C 67.26%, H 4.73%, N 7.81%.

*Preparation of (S)-5,7-difluoro-2-(4-(3-fluoropyrrolidin-1-yl)phenyl)-3-methylquinolin-4(1H)-one* ***38b.***White solid (47%); m.p. 313-314°C. NMR: 1H (400 MHz, DMSO) δ 11.47 (s, 1H), 7.38 (d, J = 8.6 Hz, 2H), 7.18 (d, J = 9.2 Hz, 1H), 6.99 (ddd, J = 12.0, 9.7, 2.4 Hz, 1H), 6.73 (d, J = 8.7 Hz, 2H), 5.50 (d, J = 54.3 Hz, 1H), 3.69 – 3.36 (m, 4H), 2.36 – 2.13 (m, 2H), 1.89 (s, 3H); 13C (101 MHz, DMSO) δ 175.38, 148.32, 147.93, 142.78, 130.36, 121.60, 116.19, 111.70, 110.54, 99.36, 98.76, 94.49, 92.78, 54.48, 45.59, 32.14, 31.93, 12.58. ES HRMS: m/z found 359.1381, C20H18N2OF3 requires 359.1371; Anal. C20H17N2OF3 requires C 67.03%, H 4.78%, N 7.82%, found C 67.25%, H 4.67%, N 7.86%.

*Preparation of 2-(4-(3,3-difluoroazetidin-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one* ***38c****.* White solid (33%); m.p. 316-318°C. NMR: 1H (400 MHz, DMSO) δ 11.54 (s, 1H), 7.43 (d, J = 8.4 Hz, 2H), 7.16 (d, J = 9.6 Hz, 1H), 7.01 (t, J = 10.8 Hz, 1H), 6.74 (d, J = 8.5 Hz, 2H), 4.37 (t, J = 12.3 Hz, 4H), 1.86 (s, 3H).; 13C (101 MHz, DMSO) δ 175.39, 150.88, 147.53, 142.81, 130.18, 124.67, 117.01, 116.50, 112.70, 110.53, 99.41, 98.90, 90.56, 74.81, 63.29, 12.44. ES HRMS: m/z found 363.1130, C19H15N2OF4 requires 363.1121; Anal. C19H14N2OF4 requires C 62.98%, H 3.89%, N 7.73%, found C 63.03%, H 3.79%, N 7.71%.

*Preparation of 2-(4-(3,4-difluoro-1H-pyrrol-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one* ***38d.*** White solid (38 mgs, 30 %). 1H NMR (400 MHz, DMSO) 11.78 (bs, 1H), 7.83 (d, *J* = 8.8 Hz, 2H), 7.72 (d, *J* = 8.8 Hz, 4H), 7.14 (d, *J* = 9.6 Hz, 1H), 7.11 (dd, *J* = 11.0, 10.6 Hz, 1H), 1.91 (s, 3H), 13C NMR (100 MHz, DMSO) δC 175.1, 146.8, 140.3, 131.8, 130.8, 118.7, 116.9, 103.0, 12.3; MS (ES+) *m/z* 373 (M + H)+ HRMS calculated for 373.0964 C20H13N2OF4, found 373.0965; Purity HPLC 98% (Method A) Rt = 2.60 min..

*Preparation of 2-(4-(3,4-difluoro-1H-pyrrol-1-yl)phenyl)-7-methoxy-3-methylquinolin-4(1H)-one* ***38e.***White solid (0.12 g, 32 %). 1H NMR (400 MHz, DMSO) 11.48 (bs, 1H), 8.02 (d, *J* = 9.2 Hz, 1H), 7.75 (d, *J* = 8.8 Hz, 2H), 7.66 (m, 4H), 7.01 (s, 1H), 6.90 (d, *J* = 9.0 Hz, 1H), 3.82 (s, 3H), 1.90 (s, 3H); 13C NMR (100 MHz, DMSO) δC 176.5, 161.9, 141.8, 141.1, 140.0, 138.9, 138.7, 130.8, 127.2, 118.6, 118.0, 114.3, 113.3, 102.7, 102.5, 102.4, 99.2,. 55.7, 12.5; MS (ES+) *m/z* 367 (M + H)+ HRMS calculated for 367.1258 C21H17N2O2F2, found 367.1257; Purity HPLC 99+% (Method A) Rt = 2.09 min.

*Preparation of 6-chloro-2-(4-(3,4-difluoro-1H-pyrrol-1-yl)phenyl)-7-methoxy-3-methylquinolin-4(1H)-one* ***38f.***White solid (0.11 g, 30 %). 1H NMR (400 MHz, DMSO) 11.75 (bs, 1H), 8.03 (s, 1H), 7.71 (d, *J* = 8.8 Hz, 2H), 7.63 (m, 4H), 7.15 (s, 1H), 3.89 (s, 3H), 1.91 (s, 3H); 13C NMR (100 MHz, DMSO) δC 175.1, 156.3, 139.7, 138.7, 130.8, 126.0, 118.5, 114.2, 102.7, 102.5, 102.4, 56.5, 12.9; MS (ES+) *m/z* 401 (M + H)+ HRMS calculated for 401.0868 C21H16N2O2F235Cl, found 401.0870; Purity HPLC 97% (Method A) Rt = 2.35 min..

*Preparation of 5,7-difluoro-2-(4-(3-hydroxy-3-methylpiperidin-1-yl)phenyl)-3-methylquinolin-4(1H)-one* ***38g****.* While solid (48%); m.p. decomposed at 284°C. NMR: 1H (400 MHz, DMSO) δ 11.47 (s, 1H), 7.35 (d, J = 8.8 Hz, 2H), 7.16 (d, J = 9.2 Hz, 1H), 7.07 – 6.94 (m, 3H), 4.46 (s, 1H), 3.30 – 3.02 (m, 4H), 1.88 (s, 3H), 1.86 – 1.75 (m, 1H), 1.63 – 1.48 (m, 3H), 1.17 (s, 3H); 13C (101 MHz, DMSO) δ 175.37, 163.50, 161.49, 152.33, 147.68, 142.79, 130.15, 123.19, 116.27, 114.64, 110.56, 99.34, 98.82, 67.64, 59.76, 47.81, 37.73, 27.28, 22.10, 12.52. ES HRMS: m/z found 385.1738, C22H23N2O2F2 requires 385.1728; Anal. C22H22N2O2F2 requires C 68.74%, H 5.77%, N 7.29%, found C 68.49%, H 5.84%, N 7.39%.

*Preparation of 5,7-difluoro-2-(4-(3-hydroxy-3-methylpyrrolidin-1-yl)phenyl)-3-methylquinolin-4(1H)-one* ***38h.***White solid (50%); m.p. 288-290°C. NMR: 1H (400 MHz, DMSO) δ 11.43 (s, 1H), 7.35 (d, J = 8.7 Hz, 2H), 7.18 (d, J = 10.1 Hz, 1H), 6.98 (ddd, J = 12.0, 9.6, 2.5 Hz, 1H), 6.62 (d, J = 8.8 Hz, 2H), 4.85 (s, 1H), 3.48 – 3.36 (m, 2H), 3.24 (s, 2H), 2.01 – 1.92 (m, 2H), 1.89 (s, 3H), 1.37 (s, 3H); 13C (101 MHz, DMSO) δ 175.38, 160.89, 155.31, 148.73, 148.07, 130.29, 120.65, 116.06, 111.02, 99.38, 96.34, 94.24, 91.71, 75.74, 60.95, 55.28, 46.88, 26.29, 12.63. ES HRMS: m/z found 399.1391, C21H20N2O2F223Na requires 393.1391; Purity HPLC 98% (Method A) Rt = 2.25 min.

*Preparation of 5,7-difluoro-2-(4-(3-hydroxy-3-methylazetidin-1-yl)phenyl)-3-methylquinolin-4(1H)-one* ***38i***. White solid (43%); m.p. decomposed at 289°C.NMR: 1H (400 MHz, DMSO) δ 11.48 (s, 1H), 7.35 (d, J = 8.6 Hz, 2H), 7.16 (d, J = 9.1 Hz, 1H), 6.99 (ddd, J = 12.0, 9.6, 2.4 Hz, 1H), 6.58 (d, J = 8.6 Hz, 2H), 5.60 (s, 1H), 3.83 (d, J = 7.9 Hz, 2H), 3.69 (d, J = 7.7 Hz, 2H), 1.86 (s, 3H), 1.48 (s, 3H); 13C (101 MHz, DMSO) δ 175.38, 160.90, 152.74, 147.89, 142.69, 136.81, 134.24, 130.07, 122.72, 116.28, 111.45, 99.62, 98.81, 67.73, 66.17, 27.02, 12.52. ES HRMS: m/z found 379.1237, C20H18N2O2F223Na requires 379.1234; Anal. C20H18N2O2F2 requires C 67.41%, H 5.09%, N 7.86%, found C 67.18%, H 5.49%, N 7.24%.

*Preparation of (S)-2-(4-(2-((benzyloxy)methyl)pyrrolidin-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one* ***38j****.* Cream solid (0.10 g, 20 %). 1H NMR (400 MHz, DMSO) 10.60 (bs, 1H), 7.33 (m, 6H), 7.22 (d, *J* = 8.8 Hz, 2H), 6.56 (dd, *J* = 11.0, 10.6 Hz, 1H), 6.44 (d, *J* = 8.8 Hz, 2H), 4.52 (s, 2H), 3.84 (m, 1H), 3.51 (dd, *J* = 8.8, 4.5 Hz, 1H), 3.30 (m, 2H), 3.05 (m, 1H), 2.05 (m, 4H), 1.92 (s, 3H); 13C NMR (100 MHz, DMSO) δC 177.1, 148.8, 147.9, 138.1, 129.7, 128.4, 127.8, 127.6, 121.5, 117.2, 111.3, 99.2, 73.4, 70.0, 58.2, 48.3, 28.9, 23.2, 12.4; MS (ES+) *m/z* 461 (M + H)+ HRMS calculated for 461.2041 C28H27N2O2F2, found 461.2055.

**General procedure for the preparation of compounds 39a-c.**

*Preparation of (S)-5,7-difluoro-2-(4-(2-(hydroxymethyl)pyrrolidin-1-yl)phenyl)-3-methylquinolin-4(1H)-one* ***39a.*** Cream solid (50 mgs, 90 %). 1H NMR (400 MHz, DMSO) 11.45 (bs, 1H), 7.35 (d, *J* = 8.8 Hz, 2H), 7.20 (dd, *J* = 8.0, 4.5 Hz, 1H), 7.01 (dd, *J* = 11.0, 10.6 Hz, 1H), 6.75 (d, *J* = 8.8 Hz, 2H), 4.90 (m, 1H), 3.81 (m, 1H), 3.75 (m, 1H), 3.50 (m, 1H), 3.22 (m, 1H), 3.10 (m, 1H), 2.03 (m, 4H), 1.92 (s, 3H); 13C NMR (100 MHz, DMSO) C 175.4, 148.4, 148.0, 130.3, 121.1, 116.1, 111.7, 99.3, 61.3, 60.5, 48.5, 28.3, 23.0, 12.6; MS (ES+) *m/z* 371 (M + H)+ HRMS calculated for 371.1571 C21H21N2O2F2, found 371.1568; Purity HPLC 96% (Method A) Rt = 2.25 min.

*Preparation of (R)-5,7-difluoro-2-(4-(2-(hydroxymethyl)pyrrolidin-1-yl)phenyl)-3-methylquinolin-4(1H)-one* ***39b.*** Light yellow solid (0.065 g, 85 %). 1H NMR (400 MHz, DMSO) 11.44 (bs, 1H), 7.35 (d, *J* = 8.8 Hz, 2H), 7.17 (d, *J* = 8.0 Hz, 1H), 6.99 (dd, *J* = 11.0, 10.6 Hz, 1H), 6.75 (d, *J* = 8.8 Hz, 2H), 4.84 (dd, *J* = 5.8, 5.8 Hz, 1H), 3.77 (m, 1H), 3.51 (m, 1H), 3.42 (m, 1H), 3.25 (m, 1H), 3.08 (m, 1H), 1.98 (m, 4H), 1.89 (s, 3H); 13C NMR (100 MHz, DMSO) C 175.7, 148.8, 148.0, 130.3, 121.1, 116.2, 111.7, 99.9, 61.5, 60.5, 28.5, 23.6, 12.6; MS (ES+) *m/z* 371 (M + H)+ HRMS calculated for 371.1571 C21H21N2O2F2, found 371.1572; Purity HPLC 97% (Method A) Rt = 2.24 min.

*Preparation of (R)-2-(4-(3-(aminomethyl)pyrrolidin-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one* ***39c.*** White solid (21 mgs, 93 %). 1H NMR (400 MHz, DMSO) 11.48 (bs, 1H), 7.40 (m, 1H), 7.38 (d, *J* = 8.8 Hz, 2H), 7.21 (d, *J* = 8.4 Hz, 1H), 6.99 (dd, *J* = 11.0, 10.6 Hz, 1H), 6.86 (d, *J* = 8.8 Hz, 2H), 4.09 (m, 1H), 3.20 (m, 1H), 2.99 (m, 1H), 2.51 (d, *J* = 10.4 Hz, 1H), 2.31 (dd, *J* = 14.4, 10.9 Hz, 1H), 2.13 (m, 1H), 1.98 (s, 3H), 1.82 (m, 2H), 1.63 (m, 2H); 13C NMR (100 MHz, DMSO) C 175.4, 147.8, 130.4, 116.2, 112.0, 111.6, 99.7, 56.5, 56.2, 48.3, 34.6, 28.5, 12.6; MS (ES+) *m/z* 370 (M + H)+ HRMS calculated for 370.1731 C21H22N3OF2, found 370.1738; Purity HPLC 96% (Method A) Rt = 1.61 min.

*Preparation of 2-(4-(3,3-difluoropyrrolidin-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one* ***42a****.* White solid (56%); m.p. decomposed at 316°C. NMR: 1H (400 MHz, DMSO) δ 7.41 (d, J = 8.7 Hz, 2H), 7.17 (d, J = 9.0 Hz, 1H), 7.01 (ddd, J = 12.0, 9.6, 2.4 Hz, 1H), 6.78 (d, J = 8.8 Hz, 2H), 3.79 (t, J = 13.3 Hz, 1H), 3.56 (t, J = 7.2 Hz, 1H), 2.59 (tt, J = 14.5, 7.3 Hz, 1H), 1.87 (s, 1H); 13C (101 MHz, DMSO) δ 175.38, 164.09, 148.09, 147.72, 142.82, 130.34, 129.16, 126.71, 122.79, 116.32, 111.98, 111.61, 99.37, 98.82, 54.96, 45.75, 33.72, 12.54. ES HRMS: m/z found 399.1093, C20H16N2OF423Na requires 399.1096; Anal. C20H16N2OF4 requires C 63.83%, H 4.29%, N 7.44%, found C 63.49%, H 4.31%, N 7.28%.

*Preparation of 2-(4-(3,3-difluoropiperidin-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one* ***42b.***White solid (47%); m.p. decomposed at 297°C. NMR: 1H (400 MHz, DMSO) δ 11.54 (s, 1H), 7.39 (d, J = 8.8 Hz, 2H), 7.20 – 7.11 (m, 3H), 7.01 (ddd, J = 12.0, 9.6, 2.4 Hz, 1H), 3.65 (t, J = 11.9 Hz, 2H), 3.43 – 3.37 (m, 2H), 2.16 – 2.01 (m, 2H), 1.87 (s, 3H), 1.85 – 1.75 (m, 2H); 13C (101 MHz, DMSO) δ 175.38, 152.75, 150.88, 147.47, 142.69, 130.26, 124.51, 121.44, 116.43, 115.09, 113.88, 110.51, 99.67, 98.87, 53.21, 52.92, 46.93, 32.09, 21.59, 12.47. ES HRMS: m/z found 391.1441, C21H19N2OF4 requires 391.1434; Anal. C21H18N2OF4 requires C 64.61%, H 4.65%, N 7.18%, found C 64.06%, H 4.61%, N 7.05%.

*Preparation of (R)-N-(tert-butyl)-1-(4-(3-methyl-4-oxo-1,4-dihydroquinolin-2-yl)phenyl)pyrrolidine-2-carboxamide* ***45a***. Pale yellow powder (yield 20%); m.p. 164-166 oC 1H NMR (400 MHz, CDCl3-d6) δH 11.11 (s, 1H,NH), 7.43 (d, 2H, J = 8.6 Hz, Ar), 7.34 (d, 1H, J = 9.6 Hz, Ar), 6.71-6.61 (m, 1H, Ar), 6.55 (d, 2H, J = 8.6 Hz, Ar), 6.28 (s, 1H, NH), 3.59 (t, 1H, J = 7.2 Hz, CH), 2.99 (dd, 1H, J = 15.4 Hz, 8.9 Hz, CH2), 2.89 (d, 1H, J = 8.6 Hz, CH2), 2.03 (s, 3H, CH3), 1.92-1.65 (m, 4H, CH2), 1.34 (m, 9H, CH3) 13C NMR (100 MHz, CDCl3-d6) δc 173.1, 148.0, 130.1, 125.3, 117.7, 113.1, 64.6, 51.3, 49.8, 31.4, 28.6, 24.0, 12.3 MS (ES+), [M + Na] + (100) 462.2 HRMS calculated for 462.1969 C25H27O2N3F2Na, found 462.1955; Anal. C25H27N3O2F2 requires C 68.32%, H 6.19%, N 9.56%, found C 68.13%, H 6.10%, N 9.11%.

*Preparation of (R)-1-(4-(5,7-difluoro-3-methyl-4-oxo-1,4-dihydroquinolin-2-yl)phenyl)-N,N-dimethylpyrrolidine-2-carboxamide* ***45b***. Pale yellow powder (Yield 34%); m.p. 176-178 o C. 1H NMR (400 MHz, CDCl3-d6) δH 10.40 (s, 1H, NH), 7.24-7.22 (m, 1H,Ar), 7.12 (d, 2H, J = 8.6 Hz, Ar), 6.73-6.56 (m, 1H, Ar), 6.13 (d, 2H, J = 8.6 Hz, Ar), 4.22 (dd, 1H.J = 8.8 Hz, 2.1 Hz, CH), 3.46-3.39 (m, 1H, CH2), 3.25 (dd, 1H, J = 16.0 Hz, 8.4 Hz, CH2), 3.16 (s, 3H, NCH3), 2.85 (s, 3H, NCH3), 2.35-2.23 (m, 1H, CH2), 2.20-1.95 (m, 3H, CH2), 1.90 (s, 3H, CH3) 13C NMR (100 MHz, CDCl3-d6) δc 177.7, 172.7, 147.9, 129.6, 122.1, 117.1, 111.0, 58.6, 48.5, 36.9, 36.0, 30.5, 23.6, 15.3, 12.5 MS (ES+), [M + Na] + (100) 434.2 HRMS calculated for 434.1656 C23H23O2N3F2Na, found 434.1669; Purity HPLC 97% (Method B) Rt = 1.95 min.

*Preparation of (R)-1-(4-(5,7-difluoro-3-methyl-4-oxo-1,4-dihydroquinolin-2-yl)phenyl)-N-(tetrahydro-2H-pyran-4-yl)pyrrolidine-2-carboxamide* ***45c***. Pale yellow powder (yield 25%) m.p 228-230 oC 1H NMR (400 MHz, CDCl3-d6) δH 10.82 (s, 1H, NH), 7.36 (d, 2H, J = 8.7 Hz, Ar), 7.25 (d, 1H, J = 9.6 Hz, Ar), 6.68-6.59 (m, 1H, Ar), 6.56 (s, 1H, NH), 6.53 (d, 2H, J = 8.7 Hz, Ar), 4.06-3.82 (m, 2H, CH/CH2), 3.66-3.56 (m, 1H, CH2), 3.52-3.39 (m, 3H, CH2),3.30 (d, 1H, J = 6.7 Hz, CH2), 3.11-3.02 (m, 1H, CH2), 2.05-1.71 (m, 9H, CH2/CH3), 1.53-1.30 (m, 2H, CH2) 13C NMR (100 MHz, CDCl3-d6) δc 177.1, 173.1, 148.0, 147.2, 130.0, 124.9, 117.6, 112.9, 66.6, 65.9, 64.1, 49.7, 46.0, 32.9, 31.4, 24.1, 15.3, 12.3 MS (ES+), [M + Na] + (100) 490.2 HRMS calculated for 490.1018 C26H27O3N3F2Na, found 490.1932; Purity HPLC 93% (Method B) Rt = 1.92 min.

*Preparation of (R)-5,7-difluoro-3-methyl-2-(4-(2-(morpholine-4-carbonyl)pyrrolidin-1-yl)phenyl)quinolin-4(1H)-one* ***45d.*** Pale yellow powder (yield 18%); m.p. 236-238 oC. 1H NMR (400 MHz, CDCl3-d6) δH 10.26 (s, 1H, NH), 7.20 (d, 1H, J = 9.2 Hz, Ar), 7.14 (d, 2H, J = 8.6 Hz, Ar), 6.67-6.59 (m, 1H, Ar), 6.17 (d, 2H, J = 8.6 Hz, Ar), 4.44-4.37 (m, 1H, CH), 3.78 (dd, 1H, CH2), 3.74-3.55 (m, 6H, CH2), 3.46-3.35 (m, 2H, CH2), 3.27 (dd, 1H, J = 16.1 Hz, 8.3 Hz, CH2), 2.36-2.24 (m, 1H, CH2), 2.18-2.04 (m, 2H, CH2), 2.03-1.96 (m, 1H, CH2), 1.90 (s, 3H, CH3) 13C NMR (100 MHz, CDCl3-d6) δc 171.2, 147.7, 147.0, 129.6, 122.2, 117.2, 111.1, 67.0, 66.5, 58.7, 48.5, 45.8, 42.5, 30.8, 23.6, 12.4 MS (ES+), [M + Na] + (100) 476.2 HRMS calculated for 476.1762 C25H25O3N3F2Na, found 476.1778; Purity HPLC 93% (Method B) Rt = 1.90 min.

*Preparation of (R)-5,7-difluoro-2-(4-(2-(4-fluoropiperidine-1-carbonyl)pyrrolidin-1-yl)phenyl)-3-methylquinolin-4(1H)-one* ***45e.*** Pale yellow powder (yield 24%); m.p. 238-240 oC. 1H NMR (400 MHz, CDCl3-d6) δH 10.21 (s,1H, NH), 7.24-7.08 (m, 3H, Ar), 6.63 (t, 1H, Ar), 6.18 (dd, 2H, J = 7.7 Hz, 5.0 Hz, Ar), 5.04-4.81 (m, 1H, CHF), 4.44 (d, 1H, CH), 3.88-5.59 (m, 3H, CH2), 3.58-3.32 (m, 1H, CH2), 3.31-3.19 (m, 1H, CH2), 2.40-2.24 (m, 1H, CH2), 2.18-1.61 (m, 11H, CH2) 13C NMR (100 MHz, CDCl3-d6) δc 171.4, 148.0, 147.7, 129.6, 122.4, 116.9, 111.1, 65.9, 58.8, 48.5, 38.8, 30.9, 23.9, 12.4. MS (ES+), [M + Na] + (100) 492.2 HRMS calculated for 492.1875 C26H26O2N3F3Na, found 492.1872; Purity HPLC 96% (Method A) Rt = 2.20 min.

*Preparation of 4(R)-1-(4-(5,7-difluoro-3-methyl-4-oxo-1,4-dihydroquinolin-2-yl)phenyl)-N,N-dimethylazetidine-2-carboxamide* ***45f***. White solid (0.056 g, 14%). δH [400 MHz, (CD3)2SO] 1.87 (3 H, s, CH3C), 2.30-2.40, 2.60-2.75 (2 H, 2m, CCH2C),2.88, 2.94 (6 H, 2s, Me2N), 3.72, 3.93 (2 H, 2m, CH2N), 4.92 (1 H, approx. t, CHN), 6.51 (2 H, d, ArH), 7.00 (1 H, m, ArH), 7.17 (1 H, m, ArH), 7.33 (2 H, d, ArH) and 11.49 (1 H, br s, NH); δC [100 MHz, (CD3)2SO] 12.5, 22.1, 35.4, 35.8, 49.0, 63.2, 111.5, 116.3, 123.0, 129.8, 148.0, 151.9, 170.5 and 175.4; not all the aromatic carbons were seen; m/z (ES +ve mode) 398 (MH+, 100%); Found: m/z, 398.1667. C22H22N3O2F2 requires m/z, 398.1680; Anal. C22H21N3O2F2 requires C 66.49%, H 5.33%, N 10.57%, found C 66.15%, H 5.36%, N 9.88%.

*Preparation of (R)-N-(tert-butyl)-1-(4-(5,7-difluoro-3-methyl-4-oxo-1,4-dihydroquinolin-2-yl)phenyl)azetidine-2-carboxamide* ***45g***. Pale yellow powder (0.033 g, 12%). δH [400 MHz, CDCl3] 1.42 (9 H, s, Me3C), 2.00 (3 H, s, CH3C=), 2.20-2.30 (2 H, m, CCH2C), 3.26 (1 H, m), 3.58 (1 H, m), 3.95 (1 H, m), 6.52 (2 H, d, ArH), 6.60-6.70 (1 H, m, ArH), 7.19 (1 H, m, ArH), 7.40 (2 H, d, ArH) and 10.43 (1 H, br s, NH); m/z (CI, methane) 426 (MH+, base peak). Found: m/z, 426.1988. C24H26F2N3O2 requires m/z, 426.1986; Anal. C24H25N3O2F2 requires C 67.75%, H 5.92%, N 9.88%, found C 67.26%, H 5.88%, N 9.56%.

*Preparation of 2-(4-(3,3-difluoropyrrolidin-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4-yl acetate* ***46*.** To a suspension of 2-(4-(3,3-difluoropyrrolidin-1-yl)phenyl)-5,7-difluoro-3-methylquinolin-4(1H)-one (280mg, 0.74mmol) in THF (15ml), *t*BuOK (172mg, 1.5mmol) was added. The resulting mixture was kept stirring at room temperature for 1 hour. After that, excess acetyl chloride (0.2ml) was added and the reaction mixture was kept stirring for 3 hours at room temperature. After that, H2O (15ml) was used to quench the reaction and Et2O (50ml) was used to dilute the mixture. Organic layer was separated from the water layer, and DCM/MeOH (1:1, 20ml) was added to the organic layer to dissolve any precipitation. The organic solution was dried with MgSO4 and concentrated *in vacuo* to give the crude product. The crude product we purified by flash column chromatograph eluting with 20% EtOAc in hexane to give the title product a pale yellow solid (290mg, 94%). δH [400 MHz, CDCl3] 7.72 – 7.53 (m, 3H), 6.99 (dd, J = 15.1, 5.7 Hz, 1H), 6.66 (d, J = 8.6 Hz, 2H), 3.75 (t, J = 13.2 Hz, 2H), 3.61 (t, J = 7.1 Hz, 2H), 2.54 (ddd, J = 21.2, 14.0, 7.3 Hz, 2H), 2.46 (s, 3H), 2.32 (s, 3H); 13C NMR (101 MHz, CDCl3) δ 168.52, 163.55, 161.69 (dd, *J* = 249.3, 14.3 Hz), 157.00 (dd, *J* = 258.3, 14.3 Hz), 150.99 (t, *J* = 1.8 Hz), 149.05 (dd, *J* = 14.2, 2.6 Hz), 147.41, 130.57, 128.55, 128.04, 125.58, 121.90, 111.53, 109.74 (dd, *J* = 20.6, 5.0 Hz), 109.51 (dd, *J* = 9.3, 1.8 Hz), 103.05 (dd, *J* = 29.3, 25.9 Hz), 55.33 (t, *J* = 31.6 Hz), 45.54 (t, *J* = 3.2 Hz), 34.28 (t, *J* = 24.0 Hz), 20.71, 13.71; HRMS (ES) C22H18N2O2F323Na [M+Na]+ requires 441.1202, found 441.1212; Anal. C22H18N2O2F4 requires C 63.16%, H 4.34%, N 6.70%, found C 62.77%, H 4.29%, N 6.53%.

***Biology***

**Drug susceptibility assays using replicating and hypoxic Mtb -** For drug susceptibility assays, aerobic cultures of Mtb H37Rv were cultured as described previously 14. Cultures were grown until a mid-log growth phase was reached (Middlebrook 7H9 broth with addition of 10% albumin–dextrose–catalase solution (Becton Dickinson), 0.2% [vol/vol] glycerol and 0.05% [vol/vol] Tween 80). Hypoxic cultures of Mtb were produced using the same growth media but the method described by Wayne and Hayes was utilised 58, where oxygen supply was limited over six weeks and cultures were mixed using 8-mm Teflon-coated magnetic stirring bars (120 rpm, 37°C).

The effectiveness of test drugs to prevent Mtb growth was determined using a microplate AlamarBlue assay (MABA) as described previously 14. A range of test drug concentrations (10 µM to 0.08 µM, 2% DMSO) were co-incubated with replicating Mtb (OD 0.01, 7 days, 37°C) followed by a MABA. Measurements of well absorbance at 570 and 600 nm recorded using an Opsys MR plate reader were determined to calculate IC50 values for the inhibitors. For anaerobic cultures, co-incubations of hypoxic Mtb and test drug were performed as described for replicating Mtb, however the plates were sealed within GasPak EZ pouches containing an indicator to ensure anaerobic conditions were maintained. The plates were subsequently incubated anaerobically (7 days, 37°C) before being moved to an aerobic environment for a further 7 days. The IC50 values were calculated as described for aerobic cultures.

***In vitro* Metabolic Stability -** Mixed pools of microsomes from multiple donors were purchased from BD Biosciences, USA (Human, Rat and Mouse) (protein content 20 mg/mL). Compounds of interest were tested at 10, 1 and 0.1 µM with a final concentration of microsomal protein of 1 mg/mL. The reaction was initiated by the addition of NADPH (1 mM) and samples were incubated for up to 60 min at 37°C in a shaking incubator. The reaction was terminated at 0, 10, 30 and 60 min by the addition of ice cold ACN/MeOH (50:50) spiked with internal standard. Sample preparation for mass spectrometry involved the addition of an equivalent amount of water to each sample before extraction using ethyl acetate (3 x 500 µL). The organic layer was then dried under nitrogen before reconstitution in MeOH/H20 (50:50).

**Cytotoxicity assay in HEPG2 using MTT -** The cellular toxicity of test compounds were determined using the MTT assay, with modifications, using HEPG2 cells which were either resistant (cultured using glucose-containing media) or susceptible (cultured using galactose-containing media) to mitochondrial-toxicity-induced cell death 59, 60. Briefly, HepG2 cells cultured in glucose media (high-glucose Dulbecco’s modified Eagle’s medium (DMEM) containing 25 mM glucose and 1 mM sodium pyruvate, supplemented with 5 mM HEPES, 10% [vol/vol] fetal bovine serum (FBS), and 100 µg/mlpenicillin-streptomycin) or galactose media (glucose-free DMEM supplemented with 10 mM galactose, 5 mM HEPES, 10 % [vol/vol] FBS, 1 mM sodium pyruvate, and 100 µg/mlpenicillin-streptomycin) were added to 96-well plates (60 µl, 1 x 104 cells/well) and incubated for 24 hours. Log-range concentrations of each test compound (1-100 µM) were then added to the plates and a further incubation of 24 hours performed. Plates were subsequently incubated for 2 hours in the presence 1 mg/ml 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) solution. Cell lysis solution (50 µL, 50% [vol/vol] dimethylformamide in distilled water, 20 % [wt/vol] sodium dodecyl sulphate) was added to wells and plates were wrapped in metallic foil and mixed at 60 rpm for 2 hours at room temperature. Well absorbance at 560 nm was determined using a Varioskan plate reader (ThermoScientific) and were used to determine IC50 values using a four parameter logistic function using Prism 5 software. All incubations were performed at 37 ˚C in a CO2 incubator and compounds were solubilised in DMSO (1% [vol/vol] final concentration). The cytotoxic control compounds rotenone (0.001 µM – 1 µM, toxic to mitochondria) and tamoxifen (1-100 µM, no specific mitochondrial toxicity) were included as controls, as was a drug-free control containing 1% [vol/vol] DMSO.

**Caco-2 transepithelial drug transport -** Caco-2 monolayer experiments were performed as previously described 61, with modifications. When confluent, Caco-2 cells were seeded onto polycarbonate membrane transwells at a density of 2.6 X 105 cells/cm2 (DMEM, 15% [vol/vol] FCS) and incubated (37°C, 5% CO2) for 16 hours. Following this incubation, media was replaced to remove dead cells and to prevent the formation of multiple layers of cells settling on the filter. Plate media was changed every 48 hours and plates used in experiments 21 days from initial seeding. Monolayer integrity was checked using a MillicellERS instrument (Millipore) to determine the trans-epithelial electrical resistance (TEER) across the monolayer. A TEER of more than 400 Ω/cm2 was deemed acceptable.

On the day of the experiment, the TEER was assessed and the media replaced with warm transport buffer (HBSS, 25 mM HEPES, 0.1% [wt/vol] bovine serum albumin, pH 7) and allowed to equilibrate (37°C, 30 minutes). The transport buffer in the chambers was replaced with transport buffer containing either the test compound or the control drug verapamil (5 µM). Samples (50 µL) were taken from the receiver compartment at 0, 60, 120 and 180 minutes and replaced with an equal volume of transport buffer. Samples were analysed using LC-MS/MS. Data were used to determine apparent permeability (Papp, 10-6 cm/s) for each direction and efflux ratio (ratio of basolateral to apical Papp compared with apical to basolateral Papp). Papp was calculated using the following equation as described previously 62:

**Papp = (dQ / dt) x V**

**A x C0**

**dQ / dt** is the change in drug concentration in the receiver chamber over time (nM/s); **V** is the volume in the receiver compartment (mL); **A** is the total surface area of the transwell membrane (cm2); **C0** is the initial drug concentration in the donor compartment (nM); and **Papp** is the apparent permeability (x10-6 cm/s).

**Plasma protein binding using equilibrium dialysis -** The extent of plasma protein binding for each test compounds was determined by equilibrium dialysis. Test compound was added to human plasma which was mixed and heated (1 µM, 1% [vol/vol] DMSO, 37°C). Regenerated cellulose membranes (5000 Daltons, Harvard Apparatus) were soaked in phosphate buffer for 5 minutes and placed within Fast Micro-Equilibrium Dializers (Harvard Apparatus). One millilitre plasma containing the test drug was added to the first compartment, and 1 mL phosphate buffer (1% [vol/vol] DMSO, 37°C) was added to the second compartment. Equilibrium dialysis was undertaken by incubation (18 hours, 37°C) and samples were removed from each compartment for LC-MS/MS analysis.

**Plasma Stability -** Compounds were incubated in rat or human plasma (1 µM) at 37 °C for up to 3 h. At various time-points (0, 10, 30, 60, 120 and 180 min) an aliquot (100 µL) was taken and the reaction was terminated by the addition of ice cold ACN/MeOH (300 µL, 50%:50% [vol/vol]) spiked with internal standard. Samples underwent centrifugation to remove the protein precipitate and were analysed directly using LC-MS/MS analysis.

***In vitro* CYPP450 Inhibition -** CYPP450 VIVID® inhibition kits were purchased from Invitrogen Life Technologies™. Briefly, compounds were tested at a final concentration of 10, 1 and 0.1 µM alongside a relevant positive control for the isoform of interest and a solvent control. The assay utilised a substrate, specific to the isoform, which produced a fluorescent metabolite as it underwent oxidation by the P450 enzyme. Inhibition of the enzyme led to reduced fluorescent output. The assay was carried out in kinetics mode, with a reading being taken every minute for a total of 1 h.

**Pharmacokinetic Studies in Rats -** Male Wistar rats (180 – 250 g) (n=4) were purchased from Charles River Laboratories, UK and allowed to acclimatise for 1 week in controlled conditions (23 ± 3 °C; relative humidity 50 ± 10 %; light-dark cycle 12 h). Animals were provided with feed pellet and filtered water *ad libitum*. Each rat received an oral dose of the relevant compound (10 or 50 mg/kg) in PEG400 (100 %) (5 mL/kg) via gavage needle or an IV injection of the relevant compound (0.5 mg/kg) in 5% PEG400 and 5% Solutol in water. At various time-points the rats were anaesthetised using isoflurane and a blood sample (< 300 µL) was taken from a superficial vein in the tail. The blood was immediately stored on ice before undergoing centrifugation at 13,000 rpm, for 10 minutes. An aliquot of 100 µL plasma was removed and added to ACN/MeOH (300 µL, 50%:50% [vol/vol]) spiked with internal standard. Samples were then analysed using LC-MS/MS within 24 hours of obtaining the final sample.

PK data were modelled using the package Pmetrics® 63 utilising a one compartment gut absorption model. Separate doses were modelled separately to differentiate the effect of dose upon the pharmacokinetic profile of each compound.

**LC-MS/MS -** Drug concentration analyses were performed on a TSQ Quantum Access mass spectrometer (Thermo, UK). Chromatographic separation for all test compounds and control compounds was performed at 30˚C on a Fortis C-18 3 µm column (50 X 2.1 mm i.d., Fortis technologies, UK). Mobile phases were solution A (100% acetonitrile) and solution B (100% LC-MS/MS-grade water, 0.05% formic acid) and flow rate was 0.3 mL/min. Separation was achieved with a gradient elution beginning with 90% solution D and 10% solution A, which was maintained for 1 minute. Solution A was then gradually increased to 80% over 1.9 minutes and maintained for a further 1.4 minutes. Solution B was increased to 90% over 0.7 minutes and maintained for 0.2 minutes, giving a total run time of 5.2 minutes. Robustness of analyses were assessed using standard concentration curves and quality control concentrations, where concentration standard deviations were required to be within 20% for generated results to be accepted.

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Notes

The authors declare no competing financial interest.

ASSOCIATED CONTENT

**Supporting Information**.

Supporting information includes:

1. Quinolone screening summary
2. Full experimental for all intermediates.
3. Metabolite identification report for MTC420.
4. Molecular formula strings.

This material is available free of charge via the Internet at http://pubs.acs.org.

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ABBREVIATIONS

1. TB – tuberculosis, MDR – multi-drug resistant, XDR – extensively drug resistant, Mtb – *Mycobacterium tuberculosis*, NADH - Nicotinamide adenine dinucleotide, ETC – electron transport chain, ATP - Adenosine triphosphate, ETF – electron transferring flavoprotein, FRD – fumarate reductase, nar – nitrate reductase, HTS – high throughput screen, DMPK – drug metabolism and pharmacokinetics, SAR – structure activity relationship, DMF – dimethyl formamide, GSK – Glaxosmithkline, NBS – *N*-bromo succinamide, DCM – dichloromethane, PCC - pyridinium chlorochromate, EDC -1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide, NHS – *N*-hydroxy succinamide, GLU – glucose, PPB – plasma protein binding, CL - clearance, AUC- area under the curve, TI – therapeutic index, hERG - human Ether-à-go-go-Related Gene, NC – not calculated, ND – not determined, ID – identification, M – metabolite, SD – Sprague Dawley, HPLC – High performance liquid chromatography, TLC – thin layer chromatography, DMSO – dimethyl sulfoxide, NADPH - nicotinamide adenine dinucleotide phosphate, MTT - 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide, DMEM - Dulbecco's Modified Eagle's Medium, FBS – fetal bovine serum, HEPES - (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid, FCS – fetal calf serum, TEER - trans-epithelial electrical resistance, HBSS – Hank’s balance salt solution, LC-MS – Liquid chromatograph-mass spectrometry.

REFERENCES

1. World Health Organisation. *Global Tuberculosis Report 2015: Executive Summary*, 20th ed.; Geneva, Switzerland, 2015; pp 1-4.

2. Streicher, E. M.; Müller, B.; Chihota, V.; Mlambo, C.; Tait, M.; Pillay, M.; Trollip, A.; Hoek, K. G. P.; Sirgel, F. A.; Gey van Pittius, N. C.; van Helden, P. D.; Victor, T. C.; Warren, R. M. Emergence and treatment of multidrug resistant (MDR) and extensively drug-resistant (XDR) tuberculosis in South Africa. *Infect. Genet. Evol.* **2012,** *12*, 686-694.

3. Koul, A.; Arnoult, E.; Lounis, N.; Guillemont, J.; Andries, K. The challenge of new drug discovery for tuberculosis. *Nature* **2011,** *469*, 483-490.

4. Goel, D. Bedaquiline: A novel drug to combat multiple drug-resistant tuberculosis. *J. Pharmacol. Pharmacother.* **2014,** *5*, 76-78.

5. Guillemont, J.; Meyer, C.; Poncelet, A.; Bourdrez, X.; Andries, K. Diarylquinolines, synthesis pathways and quantitative structure–activity relationship studies leading to the discovery of TMC207. *Future Med. Chem.* **2011,** *3*, 1345-1360.

6. Xavier, A. S.; Lakshmanan, M. Delamanid: A new armor in combating drug-resistant tuberculosis. *J. Pharmacol. Pharmacother.* **2014,** *5*, 222-224.

7. Matsumoto, M.; Hashizume, H.; Tomishige, T.; Kawasaki, M.; Tsubouchi, H.; Sasaki, H.; Shimokawa, Y.; Komatsu, M. OPC-67683, a nitro-dihydro-imidazooxazole derivative with promising action against tuberculosis in vitro and in mice. *PLoS Med.* **2006,** *3*, e466.

8. Fox, G. J.; Menzies, D. A Review of the evidence for using bedaquiline (TMC207) to treat multi-drug resistant tuberculosis. *Infect. Dis. Ther.* **2013,** *2*, 123-144.

9. Weinstein, E. A.; Yano, T.; Li, L. S.; Avarbock, D.; Avarbock, A.; Helm, D.; McColm, A. A.; Duncan, K.; Lonsdale, J. T.; Rubin, H. Inhibitors of type II NADH:menaquinone oxidoreductase represent a class of antitubercular drugs. *Proc. Natl. Acad. Sci. U. S. A.* **2005,** *102*, 4548-4553.

10. Koul, A.; Dendouga, N.; Vergauwen, K.; Molenberghs, B.; Vranckx, L.; Willebrords, R.; Ristic, Z.; Lill, H.; Dorange, I.; Guillemont, J.; Bald, D.; Andries, K. Diarylquinolines target subunit c of mycobacterial ATP synthase. *Nat. Chem. Biol.* **2007,** *3*, 323-324.

11. Haagsma, A. C.; Abdillahi-Ibrahim, R.; Wagner, M. J.; Krab, K.; Vergauwen, K.; Guillemont, J.; Andries, K.; Lill, H.; Koul, A.; Bald, D. Selectivity of TMC207 towards mycobacterial ATP synthase compared with that towards the eukaryotic homologue. *Antimicrob. Agents Chemother.* **2009,** *53*, 1290-1292.

12. Koul, A.; Vranckx, L.; Dendouga, N.; Balemans, W.; Van den Wyngaert, I.; Vergauwen, K.; Gohlmann, H. W.; Willebrords, R.; Poncelet, A.; Guillemont, J.; Bald, D.; Andries, K. Diarylquinolines are bactericidal for dormant mycobacteria as a result of disturbed ATP homeostasis. *J. Biol. Chem.* **2008,** *283*, 25273-25280.

13. Diacon, A. H.; Pym, A.; Grobusch, M.; Patientia, R.; Rustomjee, R.; Page-Shipp, L.; Pistorius, C.; Krause, R.; Bogoshi, M.; Churchyard, G.; Venter, A.; Allen, J.; Palomino, J. C.; De Marez, T.; van Heeswijk, R. P.; Lounis, N.; Meyvisch, P.; Verbeeck, J.; Parys, W.; de Beule, K.; Andries, K.; Mc Neeley, D. F. The diarylquinoline TMC207 for multidrug-resistant tuberculosis. *N. Engl. J. Med.* **2009,** *360*, 2397-2405.

14. Warman, A. J.; Rito, T. S.; Fisher, N. E.; Moss, D. M.; Berry, N. G.; O'Neill, P. M.; Ward, S. A.; Biagini, G. A. Antitubercular pharmacodynamics of phenothiazines. *J. Antimicrob. Chemother.* **2013,** *68*, 869-880.

15. Pethe, K.; Bifani, P.; Jang, J.; Kang, S.; Park, S.; Ahn, S.; Jiricek, J.; Jung, J.; Jeon, H. K.; Cechetto, J.; Christophe, T.; Lee, H.; Kempf, M.; Jackson, M.; Lenaerts, A. J.; Pham, H.; Jones, V.; Seo, M. J.; Kim, Y. M.; Seo, M.; Seo, J. J.; Park, D.; Ko, Y.; Choi, I.; Kim, R.; Kim, S. Y.; Lim, S.; Yim, S.-A.; Nam, J.; Kang, H.; Kwon, H.; Oh, C.-T.; Cho, Y.; Jang, Y.; Kim, J.; Chua, A.; Tan, B. H.; Nanjundappa, M. B.; Rao, S. P. S.; Barnes, W. S.; Wintjens, R.; Walker, J. R.; Alonso, S.; Lee, S.; Kim, J.; Oh, S.; Oh, T.; Nehrbass, U.; Han, S.-J.; No, Z.; Lee, J.; Brodin, P.; Cho, S.-N.; Nam, K.; Kim, J. Discovery of Q203, a potent clinical candidate for the treatment of tuberculosis. *Nat. Med.* **2013,** *19*, 1157-1160.

16. Abrahams, K. A.; Cox, J. A.; Spivey, V. L.; Loman, N. J.; Pallen, M. J.; Constantinidou, C.; Fernandez, R.; Alemparte, C.; Remuinan, M. J.; Barros, D.; Ballell, L.; Besra, G. S. Identification of novel imidazo[1,2-a]pyridine inhibitors targeting M. tuberculosis QcrB. *PLoS One* **2012,** *7*, e52951.

17. Kana, B. D.; Machowski, E. E.; Schechter, N.; Shin, J.-T.; Rubin, H.; Mizrahi, V. Electron transport and respiration. In *Mycobacterium: Genomics and Molecular Biology* Parish, T.; Brown, A., Eds. Horizon Press, London, United Kingdom: 2009; pp 35-64.

18. Rao, S. P. S.; Alonso, S.; Rand, L.; Dick, T.; Pethe, K. The protonmotive force is required for maintaining ATP homeostasis and viability of hypoxic, nonreplicating Mycobacterium tuberculosis. *Proc. Natl. Acad. Sci. U. S. A.* **2008,** *105*, 11945-11950.

19. Lloyd, D.; Hayes, A. J. Vigor, vitality and viability of microorganisms. *FEMS Microbiol. Lett.* **1995,** *133*, 1 - 7.

20. Griffin, J. E.; Gawronski, J. D.; DeJesus, M. A.; Ioerger, T. R.; Akerley, B. J.; Sassetti, C. M. High-resolution phenotypic profiling defines genes essential for Mycobacterial growth and cholesterol catabolism. *PLoS Pathog.* **2011,** *7*, 9.

21. Awasthy, D.; Ambady, A.; Narayana, A.; Morayya, S.; Sharma, U. Roles of the two type II NADH dehydrogenases in the survival of Mycobacterium tuberculosis in vitro. *Gene* **2014,** *550*, 110-116.

22. Betts, J. C.; Lukey, P. T.; Robb, L. C.; McAdam, R. A.; Duncan, K. Evaluation of a nutrient starvation model of Mycobacterium tuberculosis persistence by gene and protein expression profiling. *Mol. Microbiol.* **2002,** *43*, 717-731.

23. Winder, F. G.; Collins, P. B. Inhibition by isoniazid of synthesis of mycolic acids in Mycobacterium tuberculosis. *Microbiology* **1970,** *63*, 41-48.

24. Brennan, P. J.; Crick, D. C. The cell-wall core of mycobacterium tuberculosis in the context of drug discovery. *Curr. Top. Med. Chem.* **2007,** *7*, 475-488.

25. Khisimuzi, M.; Zhenkun, M. Mycobacterium tuberculosis DNA Gyrase as a target for drug discovery. *Infect. Disord. Drug Targets* **2007,** *7*, 159-168.

26. Aubry, A.; Pan, X.-S.; Fisher, L. M.; Jarlier, V.; Cambau, E. Mycobacterium tuberculosis DNA Gyrase: Interaction with quinolones and correlation with antimycobacterial drug activity. *Antimicrob. Agents Chemother.* **2004,** *48*, 1281-1288.

27. Shi, W.; Zhang, X.; Jiang, X.; Yuan, H.; Lee, J. S.; Barry, C. E.; Wang, H.; Zhang, W.; Zhang, Y. Pyrazinamide inhibits trans-translation in Mycobacterium tuberculosis. *Science* **2011,** *333*, 1630-1632.

28. Campbell, E. A.; Korzheva, N.; Mustaev, A.; Murakami, K.; Nair, S.; Goldfarb, A.; Darst, S. A. Structural mechanism for rifampicin inhibition of bacterial RNA polymerase. *Cell* **2001,** *104*, 901-912.

29. Fisher, N.; Warman, A. J.; Ward, S. A.; Biagini, G. A. Chapter 17 Type II NADH: quinone oxidoreductases of Plasmodium falciparum and Mycobacterium tuberculosis kinetic and high-throughput assays. *Methods Enzymol.* **2009,** *456*, 303-320.

30. Warman, A. J.; Rito, T. S.; Fisher, N. E.; Moss, D. M.; Berry, N. G.; O'Neill, P. M.; Ward, S. A.; Biagini, G. A. Antitubercular pharmacodynamics of phenothiazines. *J. Antimicrob. Chemother.* **2013,** *68*, 869-880.

31. Kristiansen, J. E.; Dastidar, S. G.; Palchoudhuri, S.; Roy, D. S.; Das, S.; Hendricks, O.; Christensen, J. B. Phenothiazines as a solution for multidrug resistant tuberculosis: From the origin to present. *Int. Microbiol.* **2015,** *18*, 1-12.

32. Durant, J. L.; Leland, B. A.; Henry, D. R.; Nourse, J. G. Reoptimization of MDL keys for use in drug discovery. *J. Chem. Inf. Comput. Sci.* **2002,** *42*, 1273-1280.

33. Willett, P. Similarity-based virtual screening using 2D fingerprints. *Drug. Discov. Today* **2006,** *11*, 1046-1053.

34. Geppert, H.; Vogt, M.; Bajorath, J. Current trends in ligand-based virtual screening: molecular representations, data mining methods, new application areas, and performance evaluation. *J. Chem. Inf. Model* **2010,** *50*, 205-216.

35. Liu, K.; Feng, J.; Young, S. S. PowerMV: a software environment for molecular viewing, descriptor generation, data analysis and hit evaluation. *J. Chem. Inf. Model* **2005,** *45*, 515-522.

36. Lipinski, C. A. Drug-like properties and the causes of poor solubility and poor permeability. *J. Pharmacol. Toxicol. Methods* **2000,** *44*, 235-249.

37. Veber, D. F.; Johnson, S. R.; Cheng, H. Y.; Smith, B. R.; Ward, K. W.; Kopple, K. D. Molecular properties that influence the oral bioavailability of drug candidates. *J. Med. Chem.* **2002,** *45*, 2615-2623.

38. Gleeson, M. P. Generation of a set of simple, interpretable ADMET rules of thumb. *J. Med. Chem.* **2008,** *51*, 817-834.

39. Hughes, J. D.; Blagg, J.; Price, D. A.; Bailey, S.; DeCrescenzo, G. A.; Devraj, R. V.; Ellsworth, E.; Fobian, Y. M.; Gibbs, M. E.; Gilles, R. W.; Greene, N.; Huang, E.; Krieger-Burke, T.; Loesel, J.; Wager, T.; Whiteley, L.; Zhang, Y. Physiochemical drug properties associated with in vivo toxicological outcomes. *Bioorg. Med. Chem. Lett.* **2008,** *18*, 4872-4875.

40. Waring, M. J. Lipophilicity in drug discovery. *Expert Opin. Drug Discov.* **2010,** *5*, 235-248.

41. Pidathala, C.; Amewu, R.; Pacorel, B.; Nixon, G. L.; Gibbons, P.; Hong, W. D.; Leung, S. C.; Berry, N. G.; Sharma, R.; Stocks, P. A.; Srivastava, A.; Shone, A. E.; Charoensutthivarakul, S.; Taylor, L.; Berger, O.; Mbekeani, A.; Hill, A.; Fisher, N. E.; Warman, A. J.; Biagini, G. A.; Ward, S. A.; O’Neill, P. M. Identification, design and biological evaluation of bisaryl quinolones targeting Plasmodium falciparum Type II NADH:quinone oxidoreductase (PfNDH2). *J. Med. Chem.* **2012,** *55*, 1831-1843.

42. Leung, S. C.; Gibbons, P.; Amewu, R.; Nixon, G. L.; Pidathala, C.; Hong, W. D.; Pacorel, B.; Berry, N. G.; Sharma, R.; Stocks, P. A.; Srivastava, A.; Shone, A. E.; Charoensutthivarakul, S.; Taylor, L.; Berger, O.; Mbekeani, A.; Hill, A.; Fisher, N. E.; Warman, A. J.; Biagini, G. A.; Ward, S. A.; O’Neill, P. M. Identification, design and biological evaluation of heterocyclic quinolones targeting Plasmodium falciparum Type II NADH:quinone oxidoreductase (PfNDH2). *J. Med. Chem.* **2012,** *55*, 1844-1857.

43. Biagini, G. A.; Fisher, N.; Shone, A. E.; Mubaraki, M. A.; Srivastava, A.; Hill, A.; Antoine, T.; Warman, A. J.; Davies, J.; Pidathala, C.; Amewu, R. K.; Leung, S. C.; Sharma, R.; Gibbons, P.; Hong, D. W.; Pacorel, B.; Lawrenson, A. S.; Charoensutthivarakul, S.; Taylor, L.; Berger, O.; Mbekeani, A.; Stocks, P. A.; Nixon, G. L.; Chadwick, J.; Hemingway, J.; Delves, M. J.; Sinden, R. E.; Zeeman, A.-M.; Kocken, C. H. M.; Berry, N. G.; O’Neill, P. M.; Ward, S. A. Generation of quinolone antimalarials targeting the Plasmodium falciparum mitochondrial respiratory chain for the treatment and prophylaxis of malaria. *Proc. Nat. Acad. Sci. U. S. A.* **2012,** *109*, 8298-8303.

44. Sharma, R.; Lawrenson, A. S.; Fisher, N. E.; Warman, A. J.; Shone, A. E.; Hill, A.; Mbekeani, A.; Pidathala, C.; Amewu, R. K.; Leung, S.; Gibbons, P.; Hong, D. W.; Stocks, P.; Nixon, G. L.; Chadwick, J.; Shearer, J.; Gowers, I.; Cronk, D.; Parel, S. P.; O'Neill, P. M.; Ward, S. A.; Biagini, G. A.; Berry, N. G. Identification of novel antimalarial chemotypes via chemoinformatic compound selection methods for a high-throughput screening program against the novel malarial target, PfNDH2: Increasing hit rate via virtual screening methods. *J. Med. Chem.* **2012,** *55*, 3144-3154.

45. Nilsen, A.; LaCrue, A. N.; White, K. L.; Forquer, I. P.; Cross, R. M.; Marfurt, J.; Mather, M. W.; Delves, M. J.; Shackleford, D. M.; Saenz, F. E.; Morrisey, J. M.; Steuten, J.; Mutka, T.; Li, Y.; Wirjanata, G.; Ryan, E.; Duffy, S.; Kelly, J. X.; Sebayang, B. F.; Zeeman, A. M.; Noviyanti, R.; Sinden, R. E.; Kocken, C. H.; Price, R. N.; Avery, V. M.; Angulo-Barturen, I.; Jimenez-Diaz, M. B.; Ferrer, S.; Herreros, E.; Sanz, L. M.; Gamo, F. J.; Bathurst, I.; Burrows, J. N.; Siegl, P.; Guy, R. K.; Winter, R. W.; Vaidya, A. B.; Charman, S. A.; Kyle, D. E.; Manetsch, R.; Riscoe, M. K. Quinolone-3-diarylethers: a new class of antimalarial drug. *Sci. Transl. Med.* **2013,** *5*, 177ra37.

46. Nilsen, A.; Miley, G. P.; Forquer, I. P.; Mather, M. W.; Katneni, K.; Li, Y.; Pou, S.; Pershing, A. M.; Stickles, A. M.; Ryan, E.; Kelly, J. X.; Doggett, J. S.; White, K. L.; Hinrichs, D. J.; Winter, R. W.; Charman, S. A.; Zakharov, L. N.; Bathurst, I.; Burrows, J. N.; Vaidya, A. B.; Riscoe, M. K. Discovery, synthesis, and optimization of antimalarial 4(1H)-quinolone-3-diarylethers. *J. Med. Chem.* **2014,** *57*, 3818-3834.

47. Monastyrskyi, A.; Kyle, D. E.; Manetsch, R. 4(1H)-Pyridone and 4(1H)-quinolone derivatives as antimalarials with erythrocytic, exoerythrocytic, and transmission blocking activities. *Curr. Top. Med. Chem.* **2014,** *14*, 1693-1705.

48. Monastyrskyi, A.; LaCrue, A. N.; Mutka, T. S.; Sakhno, Y.; Kyle, D. E.; Manetsch, R. Synthesis and evaluation of 4(1H)-quinolone prodrugs targeting multi-drug resistance P. falciparum malaria. *Abstr. Pap. Am. Chem. Soc.* **2013,** 245.

49. Bueno, J. M.; Herreros, E.; Angulo-Barturen, I.; Ferrer, S.; Fiandor, J. M.; Gamo, F. J.; Gargallo-Viola, D.; Derimanov, G. Exploration of 4(1H)-pyridones as a novel family of potent antimalarial inhibitors of the plasmodial cytochrome bc1. *Future Med. Chem.* **2012,** *4*, 2311-2323.

50. Charoensutthivarakul, S.; David Hong, W.; Leung, S. C.; Gibbons, P. D.; Bedingfield, P. T. P.; Nixon, G. L.; Lawrenson, A. S.; Berry, N. G.; Ward, S. A.; Biagini, G. A.; O'Neill, P. M. 2-Pyridylquinolone antimalarials with improved antimalarial activity and physicochemical properties. *Med. Chem. Comm.* **2015,** *6*, 1252-1259.

51. Marwaha, J.; Palmer, M.; Hoffer, B.; Freedman, R.; Rice, K.; Paul, S.; Skolnick, P. Differential electrophysiological and behavioral responses to optically active derivatives of phencyclidine. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1981,** *315*, 203-209.

52. Antilla, J. C.; Baskin, J. M.; Barder, T. E.; Buchwald, S. L. Copper−diamine-catalyzed N-arylation of pyrroles, pyrazoles, indazoles, imidazoles, and triazoles. *J. Org. Chem.* **2004,** *69*, 5578-5587.

53. Antilla, J. C.; Klapars, A.; Buchwald, S. L. The copper-catalyzed N-arylation of indoles. *J. Am. Chem. Soc.* **2002,** *124*, 11684-11688.

54. Kerekes, A. D.; Esposite, S. J.; Doll, R. J.; Tagat, J. R.; Yu, T.; Xiao, Y.; Zhang, Y.; Prelusky, D. B.; Tevar, S.; Gray, K.; Terracina, G. A.; Lee, S.; Jones, J.; Liu, M.; Basso, A. D.; Smith, E. B. Aurora kinase inhibitors based on the imidazo[1,2-a]pyrazine core: Fluorine and deuterium incorporation improve oral absorption and exposure. *J. Med. Chem.* **2011,** *54*, 201-210.

55. Klapars, A.; Buchwald, S. L. Copper-catalyzed halogen exchange in aryl halides:  An aromatic Finkelstein reaction. *J Am. Chem. Soc.* **2002,** *124*, 14844-14845.

56. Taniguchi, T.; Kawada, A.; Kondo, M.; Quinn, J. F.; Kunitomo, J.; Yoshikawa, M.; Fushimi, M. Preparation of pyridazinone compounds as phosphodiesterase 10A inhibitors for preventing and treating schizophrenia. US20100197651A1, 2010.

57. Miley, G. P.; Pou, S.; Winter, R.; Nilsen, A.; Li, Y. X.; Kelly, J. X.; Stickles, A. M.; Mather, M. W.; Forquer, I. P.; Pershing, A. M.; White, K.; Shackleford, D.; Saunders, J.; Chen, G.; Ting, L. M.; Kim, K.; Zakharov, L. N.; Donini, C.; Burrows, J. N.; Vaidya, A. B.; Charman, S. A.; Riscoe, M. K. ELQ-300 prodrugs for enhanced delivery and single-dose cure of Malaria. *Antimicrob. Agents Chemother.* **2015,** *59*, 5555-5560.

58. Wayne, L. G.; Hayes, L. G. An in vitro model for sequential study of shiftdown of Mycobacterium tuberculosis through two stages of nonreplicating persistence. *Infect. Immun.* **1996,** *64*, 2062-2069.

59. Mosmann, T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J. Immunol. Methods* **1983,** *65*, 55-63.

60. Marroquin, L. D.; Hynes, J.; Dykens, J. A.; Jamieson, J. D.; Will, Y. Circumventing the Crabtree effect: replacing media glucose with galactose increases susceptibility of HepG2 cells to mitochondrial toxicants. *Toxicol. Sci.* **2007,** *97*, 539-547.

61. Moss, D. M.; Kwan, W. S.; Liptrott, N. J.; Smith, D. L.; Siccardi, M.; Khoo, S. H.; Back, D. J.; Owen, A. Raltegravir is a substrate for SLC22A6: a putative mechanism for the interaction between raltegravir and tenofovir. *Antimicrob. Agents Chemother.* **2011,** *55*, 879-887.

62. Elsby, R.; Surry, D. D.; Smith, V. N.; Gray, A. J. Validation and application of Caco-2 assays for the in vitro evaluation of development candidate drugs as substrates or inhibitors of P-glycoprotein to support regulatory submissions. *Xenobiotica* **2008,** *38*, 1140-1164.

63. Neely, M. N.; van Guilder, M. G.; Yamada, W. M.; Schumitzky, A.; Jelliffe, R. W. Accurate detection of outliers and subpopulations with Pmetrics, a nonparametric and parametric pharmacometric modeling and simulation package for R. *Ther. Drug Monit.* **2012,** *34*, 467-476.

Table of Contents Graphic

