Filling the Gap 115 Years after Ronald Ross: The Distribution of the *Anopheles coluzzii* and *Anopheles gambiae* s.s from Freetown and Monrovia, West Africa

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Abstract

It was in Freetown, Sierra Leone, that the malaria mosquito *Anopheles coastalis*, now known as *Anopheles gambiae*, was first discovered as the vector of malaria, in 1899. That discovery led to a pioneering vector research in Sierra Leone and neighbouring Liberia, where mosquito species were extensively characterized. Unfortunately, the decade long civil conflicts of the 1990s, in both countries, resulted in a stagnation of the once vibrant research on disease vectors. This paper attempts to fill in some of the gaps on what is now known of the distribution of the sibling species of the *An. gambiae* complex, and especially the *An. coluzzii* and *An. gambiae* s.s, formerly known as the *An. gambiae* molecular M and S forms respectively, in the cities of Freetown and Monrovia.

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Introduction

Anopheles gambiae (formally known An. coastalis) was first incriminated as a vector of malaria by Sir Ronald Ross in Freetown, Sierra Leone in 1899 [1]. Mosquitoes belonging to the Anopheles gambiae complex are the major vectors of malaria in West Africa. As such studying their distribution, ecology and population structure is essential for effective malaria control programs. In the majority of West African countries, various studies on these important vectors have been undertaken in details. However, there is a gap in knowledge of these vectors in post conflict countries such as Sierra Leone and Liberia where ecological studies could not be undertaken in the 1990s and early 2000s because of the civil war that engulfed both countries for more than a decade.

Prior to the civil unrest in these countries, the mosquito species and their distribution were largely documented. Sierra Leone primarily served as the centre of malaria vector research and control, being the place where human malarial parasites were first observed in wild-caught *An. gambiae* and *An. funestus*, in 1899 [1]. By the 1950s extensive studies on human biting mosquitoes in both Sierra Leone [1] and Liberia [2–5] had been undertaken. However, these studies were focussed on the morphological identification of these important vectors.

By the 1980s the study of *An. gambiae* vectors of malaria in West Africa had evolved from morphometric to cytotaxonomic studies [6,7]. The development of the PCR in 1990 [8] led to a further

evolution in the identification of the sibling species of the An. gambiae complex [9], the M and S molecular forms of the An. gambiae s.s. [10] and knockdown resistance markers to insecticides [11,12] using molecular methods. Following molecular and bionomical evidence, the An. gambiae molecular M form has subsequently been renamed An. coluzzii, whiles the An. gambiae molecular S form retained the name An. gambiae s.s [13]. These studies have been undertaken and enhanced malaria control activities in most countries in West Africa. Unfortunately, the civil conflict in Liberia (1989-1996, 1999-2003) and Sierra Leone (1991-2002) stagnated the once vibrant malaria vector research activities in the cities of Monrovia and Freetown. To date, no molecular identification of members of the An. gambiae complex had been undertaken/published in Sierra Leone, and in Liberia only one study [14] has described the distribution of the M and S forms of the An. gambiae s.s and their kdr frequencies in the Bomi County. In this short communication, we report the distribution of the An. coluzzii and the An. gambiae s.s., in Freetown and Monrovia the capital cities of Sierra Leone and Liberia respectively.

Methods

Study Sites

In Sierra Leone, samples were collected in November-December 2009, from 89 households in 5 communities around Freetown. In Liberia, samples were collected in July 2011, from 100

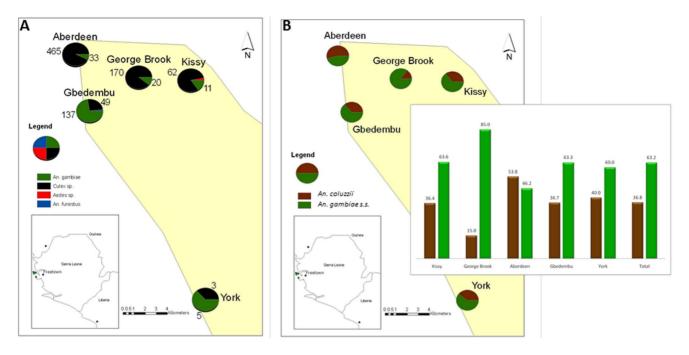


Figure 1. Distribution of the mosquito species, the *An. coluzzii* and the *An. gambiae* s.s from the sample communities in Freetown, Sierra Leone.

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households in 4 communities in Monrovia (See Figures 1 and 2 below). Samples were collected using the pyrethrum spray method. In each community, samples were collected early in the morning between 5 and 9 am, in different households, over a 3 days period.

The collected samples were identified morphologically and the *Anopheles* species were stored individually on silica gel, while the *Culex spp.* specimens were put together in pools of 20. Other mosquito species were also stored separately. The collected samples were brought to the Noguchi Memorial Institute for Medical Research, Ghana, for molecular identification.

Molecular Studies

Genomic DNA was extracted from 2–3 legs from each mosquito, using the boil preparation method. Briefly, the legs were crushed in 100 μ l of distilled water and boiled at 95°C for 10 minutes. The supernatant was used as template for the PCR. The extracted DNA was used for species identification using Polymerase Chain Reaction, Restriction Fragment Length Polymorphism (PCR-RFLP) [10]. The determination of the knockdown mutation conferring resistance to pyrethroids was undertaken using the method of Martinez-Torres et al [11].

Ethics Statement

Approval for this study was obtained from the IRB of the Liverpool School of Tropical Medicine and the Ethics and Scientific Review Committees of the Ministries of Health in Sierra Leone and Liberia. The communities, where sampling was done, were informed on the project and consent sought from the local authorities within each community. Consent was also sought from the households where mosquito sampling was carried out.

Results and Discussion

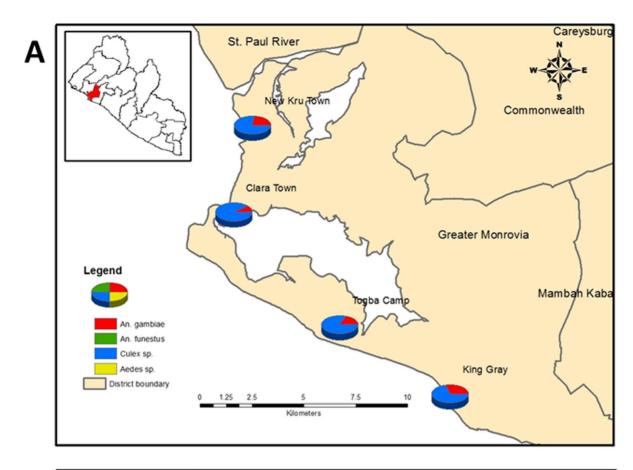
Specimen Collection and Morphological Identification

The number of collections from each community in Freetown is shown in Table 1. Overall, 206 *An. gambiae* and 749 *Culex* species were collected. In Monrovia, 161 and 381 *An. gambiae* and *Culex* species were collected respectively. *Culex* was the most common mosquito species collected, due to the polluted breeding waters found in the areas where the collections were made. *An. funestus* and *Aedes spp.* were also collected, but in very low numbers. The distribution of the mosquito species collected in Freetown and Monrovia is shown in Figure 1A and 2A respectively.

Molecular Identification and Distribution of the *An. coluzzii* and *An. gambiae* s.s

194 samples were analyzed for the molecular identification of the members of the An. gambiae complex in Freetown. 182 (93.8%) were successfully identified as either An. coluzzii or An. gambiae s.s. The remaining 12 samples failed to amplify. Further differentiation revealed 63.2% of the samples analyzed to be An. gambiae s.s., while the remaining 36.8% were An. coluzzii. The distribution of An. gambiae s.s. and An. coluzzii is shown in Figure 1B below. This study is the first to analyze the distribution of the An. gambiae s.s. and An. coluzzii in Freetown, Sierra Leone. Sierra Leone is a mountainous, forested region and this may explain the high proportion of An. gambiae s.s. compared to An. coluzzii, and is in accordance with a study [15] that showed that the An. gambiae molecular S form was dominant in mountainous, forested areas.

In Monrovia, 105 mosquitoes morphologically identified as An. gambiae were analyzed. Out of these, 90.5% were identified as An. coluzzü, and the remaining 9.5% as An. gambiae s.s. These results are consistent with the results of Temu and colleagues [14] which showed 98% of samples identified to be the An. gambiae molecular M form in the Bomi County. Figure 2B shows the distribution of An. gambiae s.s. and An. coluzzü in the collection sites in Monrovia.



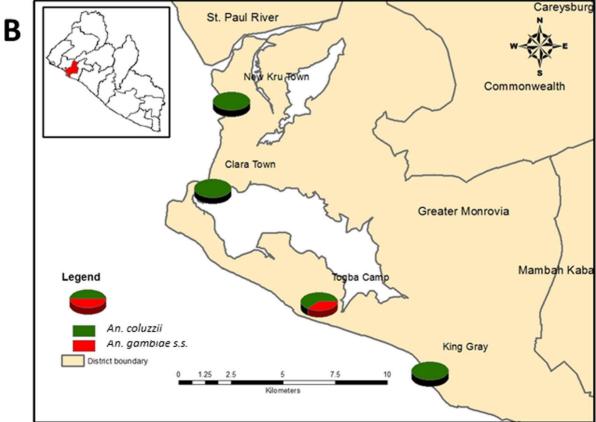


Figure 2. Distribution of the mosquito species, the *An. coluzzii* and the *An. gambiae* s.s from the sample communities in Monrovia, Liberia.

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Distribution of Knockdown Resistance Mutations

Very high proportions of kdr mutations (96.2%) were detected in the An. gambiae populations (Table 2). The mutation was present both in An. gambiae s.s. and An. coluzzü, but was more common in the An. gambiae s.s. Few hybrids were also detected. The presence of the kdr mutation was initially thought to be only present in the An. gambiae molecular S form. However, studies have shown its occurrence in the An. gambiae molecular M form [16–20]. This study showed the mutation in 91% of the An. coluzzü identified, similar to values recently reported in the An. gambiae molecular M form in Cote d'Ivoire [20]. It is suggested that the presence of the mutation in the M form may be a result of introgression from the S form [17]. However, observations from the Island of Bioko-Equatorial Guinea, where the mutation in the M form may have arisen independently due to intensive insecticide application.

The very high levels of kdr resistance to pyrethroids might be due to the high usage of insecticides (e.g. BHC and DDT) for malaria control activities in Sierra Leone, with possible cross resistance to pyrethroids. Studies conducted in the late 1950's, on the development of insecticide resistance in the An. gambiae in Freetown, showed the factor conferring resistance to BHC in 66% of the population [21]. Dieldrin resistance was observed in 91-100% of An. gambiae populations in neighboring Liberia in 1957 [22] and was reported in Freetown in 1966 [23]. HCH and DDT were widely used for house spraying and larviciding in Freetown after the World War II [1]. Temephos and malathion were also used for larviciding until it was realized that control activities were unsuccessful, with a possible resistance to malathion [1]. Despite these reports we cannot conclusively assign the observed high levels of kdr only to insecticide use in the 1950's and further studies will be required to assess this.

While the frequencies of *kdr* mutations were not determined for the Monrovia samples, it is believed that the situation may not be much different from Freetown, considering the mosquito control activities that were in place in the 1950s [1,24]. Also, recent studies in the Bomi County revealed *kdr* frequencies of over 90% [14].

Table 1. Number of mosquito collections from each community in Freetown, Sierra Leone.

Country	Community	An. gambiae	Culex sp.	Aedes sp.	An. funestus
Sierra Leone	Kissy	11	62	2	-
	George Brook	20	170	-	-
	Aberdeen	33	465	-	-
	Gbedembu	137	49	-	3
	York	5	3	-	-
	Total	206	749	2	3
Liberia	Clara Town	25	70	3	-
	New Kru Town	43	72	-	-
	King Gray	61	89	-	-
	Togba Camp	32	87	-	1
	Total	161	318	3	1

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Table 2. The distribution of the *kdr* allele in the *An. coluzzii* and the *An. gambiae* s.s in Freetown.

		<i>kdr</i> Allele			
Communities	An. spp	RR	SS	RS	
Aberdeen	An. coluzzii	85.7% (12)	14.3% (2)	-	
	An. gambiae s.s	100% (12)	-	-	
Gbedembu	An. coluzzii	93.2% (41)	6.8% (3)	-	
	An. gambiae s.s	97.4% (74)	-	2.6% (2)	
George Brook	An. coluzzii	66.7% (2)	33.3% (1)		
	An. gambiae s.s	88.2% (15)	5.9% (1)	5.9% (1)	
Kissi	An. coluzzii	100% (4)	-	-	
	An. gambiae s.s	100% (7)	-	-	
York	An. coluzzii	100% (2)	-	-	
	An. gambiae s.s	100% (3)	-	-	
Total	An. coluzzii	91.1% (61)	8.9% (6)	-	
	An. gambiae s.s	96.5% (111)	0.9% (1)	2.6% (3)	

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Although these results may represent initial information, it was recently demonstrated that there is no significant association between the presence of the 1014F *kdr* allele and ability to survive exposure to pyrethroid [20]. Thus insecticide susceptibility testing and bio-assay data are necessary to validate our findings.

Conclusions

The distribution of the An. coluzzii and An. gambiae s.s and prevalence of the kdr gene are described here for the first time in Sierra Leone, nearly 115 years after the incrimination of the Anopheles mosquito as the vector of malaria. This study showed that the An. gambiae s.s is the most dominant sibling species of An. gambiae complex in Freetown, while the An. coluzzii is the dominant species in Monrovia. A high prevalence of kdr mutation has been observed in both An. coluzzii and An. gambiae s.s in Freetown. There is however a limitation to these results, in that the determination of the kdr frequencies was not undertaken on surviving or dead mosquitoes exposed to pyrethroids through insecticide susceptibility testing. However, due to the high levels of kdr mutation in the An. coluzzii and An. gambiae s.s populations, it is recommended that insecticide susceptibility testing be done on non-pyrethroids insecticides (organophosphates and carbamates) in order to determine the insecticide of choice for future vector control activities

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Author Contributions

Conceived and designed the experiments: DKdS BGK FKB DAB MJB. Performed the experiments: DKdS. Analyzed the data: DKdS BGK MJB. Contributed reagents/materials/analysis tools: FKB DAB MJB. Wrote the paper: DKdS BGK FKB DAB MJB.

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